

# Detailed Site Investigation

25-27 Leeds Street, Rhodes, NSW

**Project No. 22148**  
Version 4

9 September 2024

Reditus Consulting Pty Ltd  
ABN: 34 631 168 502



# Detailed Site Investigation

## 25-27 Leeds Street, Rhodes, NSW

### DOCUMENT CONTROL

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# Table of Contents

<b>Executive Summary</b>	i
<b>1 Introduction</b>	1
1.1 Commissioning	1
1.2 Background	1
1.3 Objectives	2
1.4 Scope of Works	2
<b>2 Site Details</b>	3
2.1 Site Identification	3
2.2 Site Condition	3
<b>3 Site Setting and Surrounding Environment</b>	5
<b>4 Site History</b>	7
<b>5 Previous Investigations</b>	8
<b>6 Preliminary Conceptual Site Model</b>	10
6.1 Potential Sources of Contamination	10
6.2 Potentially Affected Media	10
6.3 Potential Receptors	10
6.4 Potential Transport Mechanisms and Exposure Pathways	11
6.5 Source, Pathway and Receptor Linkages	11
<b>7 Sampling and Analysis Plan</b>	13
7.1 Data Quality Objectives	13
7.2 Schedule of Works	15
7.3 Sampling Analysis Plan and Sampling Rationale	15
7.4 Laboratory Analysis and Methods	19
7.5 Tier 1 Assessment Criteria	19
<b>8 Quality Assurance and Quality Control (QAQC)</b>	22
8.1 Field Quality Assurance	22
8.2 Laboratory QA/QC	22



8.3	Evaluation of the QA/QC Information Compared to the DQOs	23
<b>9 Results</b>		<b>25</b>
9.1	Field Observations	25
9.2	Analytical Results	26
9.3	Extent of Uncertainties in the Results	30
<b>10 Discussion</b>		<b>31</b>
<b>11 Refined Conceptual Site Model</b>		<b>32</b>
<b>12 Conclusions and Recommendations</b>		<b>33</b>
12.1	Conclusions	33
<b>13 Limitations</b>		<b>34</b>
<b>14 References</b>		<b>36</b>

## List of Tables

<b>Table 1. Site Identification .....</b>	<b>3</b>
<b>Table 2. Site Setting and Surrounding Environment .....</b>	<b>5</b>
<b>Table 3 Previous Investigations .....</b>	<b>8</b>
<b>Table 4. Potential Sources of Contamination .....</b>	<b>10</b>
<b>Table 5. Preliminary Exposure Pathway Assessment.....</b>	<b>11</b>
<b>Table 6. Data Quality Objectives.....</b>	<b>13</b>
<b>Table 7. Sampling Analysis Plan and Rationale.....</b>	<b>16</b>
<b>Table 8. Groundwater Physicochemical Stabilisation Criteria .....</b>	<b>18</b>
<b>Table 9. Stabilised Groundwater Physicochemical Parameters.....</b>	<b>25</b>
<b>Table 10. Soil Analytical Data Summary .....</b>	<b>26</b>
<b>Table 11. Acid Sulfate Soil Data.....</b>	<b>28</b>
<b>Table 12. Groundwater Analytical Data Summary .....</b>	<b>28</b>
<b>Table 13. Refined Conceptual Site Model.....</b>	<b>32</b>



# Appendices

**Appendix A – Figures**

**Appendix B – Photoboard**

**Appendix C – Analytical Results Summary Tables**

**Appendix D – Borehole Logs**

**Appendix E – Field Sheets**

**Appendix F – Laboratory Reports**

**Appendix G – EIL Calculation Spreadsheet**



## Abbreviations

TERM	DEFINITION
<b>ACM</b>	Asbestos Containing Material
<b>AF/FA</b>	Asbestos Fines / Fibrous Asbestos
<b>AHD</b>	Australian Height Datum
<b>ANZG</b>	Australia and New Zealand Guidelines
<b>ASC</b>	Assessment of Site Contamination
<b>ASS</b>	Acid Sulfate Soil
<b>ASSMP</b>	Acid Sulfate Soil Management Plan
<b>bgl</b>	below ground level
<b>BTEX</b>	Benzene Toluene Ethylbenzene Xylene
<b>CEC</b>	Cation Exchange Capacity
<b>COC</b>	Chain of Custody
<b>COPC</b>	Contaminants of Potential Concern
<b>CSM</b>	Conceptual Site Model
<b>DO</b>	Dissolved Oxygen
<b>DPI</b>	Department of Primary Industries
<b>DQO</b>	Data Quality Objective
<b>DSI</b>	Detailed Site Investigation
<b>EIL</b>	Ecological Investigation Level
<b>EPA</b>	Environment Protection Authority
<b>ESL</b>	Ecological Screening Level
<b>HEPA</b>	Heads of Environment Protection Agencies
<b>HIL</b>	Health Investigation Level
<b>HSL</b>	Health Screening Level
<b>LOR</b>	Limit-Of-Reporting
<b>ML</b>	Management Limits
<b>NATA</b>	National Association of Testing Authorities
<b>NEPC</b>	National Environment Protection Council
<b>NEPM</b>	National Environment Protection Measure (2013)
<b>NSW</b>	New South Wales
<b>OC</b>	Organic Carbon
<b>OCP</b>	Organochloride Pesticides
<b>OPP</b>	Organophosphorus Pesticides



TERM	DEFINITION
<b>PAH</b>	Polycyclic Aromatic Hydrocarbons
<b>PCB</b>	Polychlorinated Biphenyls
<b>PFAS</b>	Per- and polyfluoroalkyl Substances
<b>PFHxS</b>	Perfluorohexanesulphonic acid
<b>PFOA</b>	Perfluorooctanoic acid
<b>PFOS</b>	Perfluorooctanesulfonic acid
<b>PID</b>	Photoionisation Detector
<b>PPM</b>	Parts Per Million
<b>PQL</b>	Practical Quantification Limit
<b>QA/QC</b>	Quality Assurance / Quality Control
<b>RPD</b>	Relative Percentage Difference
<b>RTK</b>	Real-time kinematic positioning
<b>SAC</b>	Soil Assessment Criteria
<b>SD</b>	Standard Deviation
<b>SEARs</b>	Secretary's Environmental Assessment Requirements
<b>SEPP</b>	State Environmental Planning Policy
<b>SPR</b>	Source-Receptor-Pathway
<b>TOC</b>	Top-of-Casing
<b>TPH</b>	Total Petroleum hydrocarbons
<b>TRH</b>	Total Recoverable Hydrocarbons
<b>UCL</b>	Upper Confidence Limit
<b>USCS</b>	Unified Soil Classification System
<b>VOC</b>	Volatile Organic Compounds



# Executive Summary

Reditus Consulting Pty Ltd (Reditus) was engaged by Billbergia Pty Ltd (the client) to complete a Detailed Site Investigation (DSI) for the proposed residential development located at 25-27 Leeds Street, Rhodes, NSW (the site).

The site is currently occupied by a large commercial building tenanted by various businesses. It is understood that the site is proposed to be redeveloped to include multi-storey residential development with basement car parking.

The objectives of the DSI were to evaluate the possibility for contamination to be present at the site as a result of current and former land use activities.

The objectives of this investigation were achieved by completing the following:

- Desktop review of the site setting and site history.
- Review of previous environmental investigations and supplement those investigations with further assessment.
- Drilling of 15 soil borings across the site and installation of 5 groundwater monitoring wells.
- Collection of 30 soil samples and analysis at a National Association of Testing Authorities (NATA) accredited laboratory for a range of contaminants of potential concern (CoPC).
- Collection of groundwater samples from the 5 new and 5 existing groundwater monitoring wells at the site and analysis for a range of CoPC.
- Preparation of this DSI report.

Based on a review of the site history, previous investigation, observations made during fieldwork, results of laboratory analysis and the proposed land use scenario, Reditus concludes the following:

- Fill materials at the site are suitable to remain onsite, from a contamination perspective, for the proposed residential land use.
- Residual natural soils are also suitable for the proposed residential land use. However, excavation of natural material and/or dewatering will require management of acid sulfate soils (ASS).
- Exceedances to the ecological criteria in soil are not seen to be an issue as this material will be removed during excavation of the basement.
- The groundwater data was consistent with previous investigations which identified heavy metals, namely copper, lead, nickel, and zinc, and Perfluorooctanesulfonic acid (PFOS) at concentrations exceeding ecological criteria. The presence of metals in groundwater appears to be a regional issue given the presence in up-gradient wells and the lack of source areas on the site. PFOS in groundwater also appears to be related to an offset source given the reduction in concentrations by over two orders of magnitude across the site in the direction of groundwater flow.
- The findings of this investigation were consistent with previous assessments completed at the site.

**Based on the above, Reditus considers that the site is suitable for the proposed development (mixed use high density residential with basement carparking) and the northern section (the foreshore area within the site boundary) is suitable to be dedicated to Council for use as a park.**

**The scope of work completed during this DSI has satisfied Items 13, 16, and 17 of the SEARs and Item 3 of the SEARs cover letter.**

Given the elevated concentrations of heavy metals and PFOS in groundwater it is recommended that groundwater not be abstracted for beneficial use at the site. As dewatering will be required to facilitate the basement excavation a dewatering management plan and an acid sulfate soil management plan will be required to ensure protection of the surrounding environment during the works. If excavation and/or groundwater extraction is required within the foreshore area of the site, the same controls will be required.

**Reditus considers that the consent authority may be satisfied that the required considerations of Clause 4.6 of State Environmental Planning Policy (Resilience and Hazards) 2021 are satisfied for the following reasons:**

- This DSI (Reditus) and previous investigations (Jacobs (2016) and ADE (2022)) completed on the development site has adequately assessed the site history to identify potential sources of contamination, and has suitability



assessed the potential risk posed by contaminants to health and the environment. As such, Clause 4.6(1)(a) of SEPP (Resilience and Hazards) 2021 has been successfully achieved.

- This DSI has determined that the land can be made suitable in its current state for the purposes for which the development is proposed to be carried out (as mixed use high density residential). As such, Clause 4.6(1) (b) and (c) of SEPP (Resilience and Hazards) 2021 have been successfully achieved.
- The DSI has been completed in accordance with the contaminated land planning guidelines, satisfying Clause 4.6(2) and 4.6(3) of SEPP (Resilience and Hazards) 2021.



# 1 Introduction

## 1.1 Commissioning

Reditus Consulting Pty Ltd (Reditus) was engaged by Billbergia Pty Ltd (the client) to complete a Detailed Site Investigation (DSI) for the proposed residential development located at 25-27 Leeds Street, Rhodes, NSW (the site). The location and layout of the site is presented in **Figure 1, Appendix A**.

## 1.2 Background

The site is occupied by a large warehouse with ground level and first floor car parking. A range of commercial tenants currently lease various parts of the building for uses including pet supplies, golf cart repairs, floorboard distribution, and furniture display.

A development application (DA2023/0235) was lodged with the City of Canada Bay (CCB) in October 2023 for the *demolition of all buildings onsite and site preparation; excavation for 2 basement levels with vehicular access via Blaxland Road; construction of a mixed-use development containing six (6) buildings ranging from 9 to 13 storeys and comprising: 7 retail premises, 249 apartments, landscape works including tree removal, through site links, communal and private open space and a foreshore park.*

On 9 February 2024 a memo was issued by the Department of Planning, Housing and Infrastructure (DPHI) relating to "Infill Affordable Housing Leeds Street Rhodes (SSD-67419241)". The Planning Secretary's Environmental Assessment Requirements (SEARs) for the SSD required the following regarding contamination at the site. These requirements from the industry specific SEARs and the SEARs cover letter.

ISSUE	DETAIL
<b>Cover Letter Requirements</b>	
<b>Item 3</b>	<i>Additional contamination assessment and remediation methods specific to the foreshore land that is proposed to be dedicated.</i>
<b>Industry Specific Requirements</b>	
<b>Issue 13</b>	<i>Ground and Water Conditions which requires a Salinity Management Plan and/or Acid Sulfate Soils Management Plan. Similarly, City of Canada Bay Council have requested A preliminary assessment carried out in accordance with the ASSMAC Acid Sulphate Soil Manual which identifies the depth of the present of ASS to what the site is proposed to excavate and fill in the data gaps which have been identified by the DESI, dated June 2023.</i>
<b>Issue 16</b>	<i>Contamination and Remediation which states in accordance with Chapter 4 of SEPP (Resilience and Hazards) 2021, assess and quantify any soil and groundwater contamination and demonstrate that the site is suitable (or will be suitable, after remediation) for the development.</i>
<b>Issue 17</b>	<i>Waste Management which states If buildings are proposed to be demolished or altered, provide a hazardous materials survey</i>

This DSI was completed to support the State Significant Development Application (SSDA) and SEARs in accordance with State Environmental Planning Policy (Resilience and Hazards) 2021 requirements. The DSI includes assessment to define the nature, extent, and degree of contamination (if any); to assess potential risk posed by contaminants to health and the environment (if any); and to obtain sufficient information to assess environmental risk and inform recommendations to render the site suitable (if required) for the proposed redevelopment.

Reditus notes that this report, including its conclusions and recommendations, must be read in conjunction with the Statement of Limitations provided in **Section 13**.



## 1.3 Objectives

The objective of the DSI was to determine whether the site is suitable for the proposed development (including the future foreshore dedication to council) in its current form or if further investigation/remediation/management is required.".

## 1.4 Scope of Works

To achieve the objectives outlined above, Reditus completed the following:

- Desktop review of the site history and site setting including review of previous investigations.
- Completion of an intrusive investigation including:
  - Location of underground services using an accredited cable locator at each proposed location.
  - Drilling of 15 boreholes, to supplement the 9 existing boreholes at the site to supplement the previous investigation locations conducted onsite.
  - Soil samples were collected from each bore near the surface (0.1-03m bgl), at 0.5 bgl, and at regular 1 metre intervals. Additional samples were taken at depths where the geology changed, and in zones of gross contamination.
  - Each soil sample location was logged in general accordance with the Unified Soil Classification System (USCS). Each soil sample was screened with a Photoionisation Detector (PID) to detect the potential presence of volatile organic compounds (VOCs).
  - Collection of soil samples from each borehole, nominally one of the fill/near surface material and one representative soil sample from underlying material. Soil samples were submitted to a NATA accredited laboratory for analysis of a range of analytes associated with the site history.
  - Conversion of 5 of the new boreholes into groundwater monitoring wells.
  - Groundwater sampling of the existing and newly installed groundwater monitoring wells using low-flow sampling methods.
  - Groundwater samples were submitted to the NATA accredited laboratory for analysis for a range of analytes associated with the site history.
  - The analytical data was compared against the human health and ecological guidelines for the proposed development.
- Preparation of a DSI report in general accordance with the ASC NEPM, NSW EPA (2020) and other relevant NSW EPA reporting guidelines.



## 2 Site Details

### 2.1 Site Identification

The site identification details have been prepared in general accordance with the NSW EPA (2020) Consultants Reporting on Contaminated Land and the ASC NEPM (2013) Field Checklist for 'site Information'. The site identification information is summarised in **Table 1** below.

**Table 1. Site Identification**

ITEM	DETAIL
<b>Address</b>	25-27 Leeds Street, Rhodes, NSW <ul style="list-style-type: none"><li>• 25 Leeds St:<ul style="list-style-type: none"><li>– Lot C DP367132</li><li>– Lot A DP329241</li></ul></li><li>• 27 Leeds St: Lot 2 DP1192949</li></ul>
<b>Title and Land Information</b>	
<b>Site Area</b>	Approximately 1.16ha
<b>Local Government Area</b>	City of Canada Bay Council
<b>Site Coordinates to the approximate centre of the site (GDA2020 MGA Zone 56)</b>	Easting: 323198.33 Northing: 6255760.83
<b>Zoning</b>	MU1 Mixed Use City of Canada Bay Local Environment Plan 2013
<b>Current Owner</b>	Billbergia
<b>Current Land Use</b>	Commercial.
<b>Future Land Use</b>	Mixed use Commercial/Residential.
<b>Trigger for Assessment</b>	To provide information to satisfy requirements of the SEARs for the SSD (see Section 1.2)  The land uses currently surrounding the site include: <ul style="list-style-type: none"><li>• <b>North:</b> Parramatta River.</li><li>• <b>South:</b> Leeds Street and low density residential.</li><li>• <b>East:</b> Commercial.</li><li>• <b>West:</b> Blaxland Road, John Whitton Reserve, and rail bridge.</li></ul>
<b>Site Layout</b>	<b>Figure 2, Appendix A</b>

### 2.2 Site Condition

The following site observations were recorded during a site visit by Reditus' Principal Environmental Engineer, Toby Scrivener on 2 May 2024:

- The site was sealed with concrete and asphalt hardstand.
- 25 Leeds Street was occupied by a building which was tenanted by a furniture display and commercial kitchen.



- 27 Leeds Street comprised a large warehouse which was tenanted by various commercial users including a pet supplies, golf cart repairer and floorboard distributer.
- An upper level carpark was constructed over the southern portion of the site.
- The hardstand was generally in good condition.
- Odours and other indicators or contamination such as staining were not observed.
- Vegetation on the boundary of the site appeared in good condition.

Photographs from the site inspection are provided in **Appendix B**.

## 3 Site Setting and Surrounding Environment

The following details regarding the site setting and surrounding environment has been taken from the previous investigations (see Section 4).

**Table 2. Site Setting and Surrounding Environment**

ITEM	DETAIL
<b>Topography and Hydrology</b>	<p>There is a high point with an approximate elevation of 20 m above the Australian Height Datum (AHD) around the central western part of the Investigation Area. The ground then slopes down to the Parramatta River. There is a rock cutting around the properties at the eastern end of Leeds Street on the southern side of the street.</p> <p>Surface water across the Investigation Area would generally flow to the Council drainage network before discharging to Parramatta River at numerous discharge points. Besides Parramatta River, there are no other natural surface water bodies within or adjoining the Investigation Area (Jacobs 2016).</p>
<b>Regional geology</b>	<p>The "Sydney Soil Landscape Series Sheet 9130" map produced by the Soil Conservation Service of NSW indicates that soils within the Investigation Area are within the Blacktown soil landscape grouping. The landscape in this grouping is characterised as gently undulating rises to steep low hills on Wianamatta Group shales and Hawkesbury sandstone. Soils are characterised as shallow to moderately deep (&lt;100cm) Red and Brown Podzolic Soils on crests, upper slopes and well drained areas; deep (150-300cm) Yellow Podzolic Soils and Soloths on lower slopes and in areas of poor drainage.</p> <p>The "Sydney Geological Series Sheet 9130, 1:100,000 scale" map produced by the Geological Survey of NSW indicates that geology in the Investigation Area consists of Wianamatta Group Hawkesbury Sandstone (Rh) which is overlain by Ashfield Shale (Rwa) in the more elevated areas away from the Parramatta River (Jacobs 2016).</p>
<b>Site Soil Description</b>	<p>The generalised site stratigraphy was summarised by Jacobs (2016) as follows:</p> <ol style="list-style-type: none"><li>1) Fill material varying in depth from approximately 1 metre in the southern portion of the site to approximately 4 metres in the areas close to the Parramatta River. Fill material was observed to consist of silty clay or sand soils with some areas of gravel and crushed rock. Building material waste such as bricks, timber was observed in fill material in some locations.</li><li>2) Residual silty clay and clay soil with a thickness varying from not present in the south-eastern portion of the site to approximately 1 – 1.5 metres in the central portion and greater than 5 metres in north-eastern portion of the site near Parramatta River. Shells were observed in the soil at several locations adjacent to Parramatta River.</li><li>3) Sandstone was encountered at depths ranging from 1 metre bgl in the south-eastern portion of the site to 6 to 7 metres bgl in the central and northern portions of the site. There was some variation in the boreholes adjacent to Parramatta River, potentially due to the reclamation of land in this area.</li></ol>

ITEM	DETAIL
<b>Acid Sulfate Soils (ASS)</b>	<p>A review of the Canada Bay Local Environmental Plan 2009 acid sulfate soil risk map indicated that the majority of the site is located within a Class 2 ASS risk area, whilst the southwestern portion is located within a Class 5 ASS risk area. Acid sulfate soils in a Class 2 areas are likely to be found below the natural ground surface. Acid sulfate soils are not usually found in Class 5 areas, however, are within 500m of adjacent Class 1, 2, 3 or 4 soils.</p> <p>Previous investigations have identified the presence of ASS to 2 metres bgl however further assessment was required to assess ASS below 2 metres to the depth of the proposed development excavation.</p>
<b>Registered Groundwater Bore Search</b>	<p>A review of the Department of Primary Industries (DPI) register of water bores determined that there were no registered bores in the Investigation Area. The nearest registered bores to the Investigation Area were:</p> <ul style="list-style-type: none"> <li>• A cluster of bores registered as monitoring wells located in Anderson Park, close to the ferry wharf on the opposite side of Parramatta River, approximately 300 metres northeast of the Investigation Area boundary.</li> <li>• Several registered bores at Wentworth Point, approximately 750 metres west of the Investigation Area boundary. These are noted to be monitoring wells.</li> <li>• A registered bore to the west of the intersection of Oulton Ave and Homebush Bay Drive approximately 800 metres southwest of the boundary of the Investigation Area. This is noted as a test irrigation well, installed to a depth of 180 metres bgl.</li> </ul> <p>Sydney Water supplies potable water throughout the Investigation Area. None of the information reviewed by Jacobs indicates that there is any beneficial use of groundwater in the Investigation Area (Jacobs 2016).</p>
<b>Regional Hydrogeology</b>	<p>A review of hydrogeology map of Australia (Geoscience Australia) indicated the aquifer type beneath the site is a fractured or fissured, extensive aquifer of low to moderate productivity.</p> <p>Groundwater was mapped by Jacobs (2016) to flow in a northerly direction toward Parramatta River.</p>
<b>Depth to Water Table</b>	<p>Groundwater was generally encountered by Jacobs (2016) in the fill / soil layer at approximately 3 metres bgl in the boreholes located adjacent to Parramatta River and between 2.5 and 5 metres bgl in the central portion of the site.</p>
<b>Yield and Inferred Groundwater Quality</b>	<p>There is no available information on the yield and groundwater quality in the surrounding area.</p>
<b>Groundwater Embargoes</b>	No groundwater embargoes apply to the site.
<b>Sensitive Environments</b>	The nearest sensitive environment includes the Parramatta River on the northern boundary of the site.

The observations made by Reditus during this investigation are similar to that made by Jacobs (2016) and others during previous investigations.

## 4 Site History

Jacobs (2016) conducted a detailed review of the site history through interrogation of Canada Bay Council files, historical aerial photographs, property titles, business directories, contaminated land records, and previous investigations.

Industrial development of the Rhodes peninsula commenced with the establishment of the NSW State Sawmill in the early 1900s. This was located to the north of Leeds Street (and appears to comprise the site). The northern portion of the site and land to the east was reclaimed from the Parramatta River in the 1920s.

Historical industry surrounding the site included:

- The John Darling & Son flour mill to the west of the rail bridge, this later became Allied Feeds.
- Timbrol Limited, a timber preservative manufacturer was located to the south of Allied Feeds. Timbrol later merged with Union Carbon to produce chemicals such as pesticides, herbicides and polyethylene film.
- State Power Station on Uhr's Point.
- Iron works to the south of the site, this later became Rhodes Corporate Park.

Most of the industry located outside of the Investigation Area closed operations from the 1980s. Large scale remediation of the areas on the western side of the railway and to the south of the Investigation Area was performed from the 1980s to 2000s in order to enable commercial and residential development. This included the remediation of dioxin contaminated waste that was used to reclaim land from Homebush Bay as well as the partial remediation of dioxin contaminated sediments in Homebush Bay.

## 5 Previous Investigations

Previous investigations made available to Reditus include the following:

- Jacobs (2016) Rhodes East Priority Investigation Area Contamination and Acid Sulphate Soils Report.
- ADE Consulting Group (2020) PFAS Investigation of Rhodes East – Leeds Foreshore Rhodes East Site 1 – Leeds Foreshore
- ADE Consulting Group (2023) Detailed Site Investigation 25-27 Leeds Street, Rhodes, NSW
- ADE Consulting Group (2023) Acid Sulfate Soil Management Plan 25 – 27 Leeds Street, Rhodes NSW

The following table summarise the details from the above reports.

**Table 3 Previous Investigations**

REPORT	KEY ITEMS
<b>Jacobs (2016) Rhodes East Priority Investigation Area Contamination and Acid Sulphate Soils Report.</b>	<p>Jacobs completed:</p> <ul style="list-style-type: none"><li>• a desktop review of the site history and site setting</li><li>• a soil and groundwater assessment.</li></ul> <p>The investigation included 25-27 Leeds St and land to the east. The investigation area is referred to as "the Rhodes East Priority Investigation Area"</p> <p>The investigation identified seven areas of potential concern:</p> <ul style="list-style-type: none"><li>• AEI 1: Reclaimed land north of Leeds Street prior to 1928</li><li>• AEI 2: Sawmill and timber activities in the current industrial area from approximately 1912 to 1972</li><li>• AEI 3: Historical power station at Uhrs Point from 1914 to sometime between 1961 and 1972</li><li>• AEI 4: Reclaimed land in the northern portion of 1 – 3 Leeds Street between 1961 and 1972</li><li>• AEI 5: Industrial activities in the current industrial area after approximately 1972</li><li>• AEI 6: Industrial activities at 14 Cavell Avenue</li><li>• AEI 7: Land contamination from asbestos containing materials</li></ul> <p>Of these, AEI 1, 2, 3, 5 and 7 are relevant to the site.</p> <p>The soil and groundwater assessment included drilling 16 soil borings across the investigation area and installation of 8 groundwater monitoring wells.</p> <p>Soil and groundwater samples were collected and analysed for a range of CoPC.</p> <p>The analytical data identified Total Recoverable Hydrocarbons (TRH), Polycyclic Aromatic Hydrocarbons (PAH) and dioxins in soil at concentrations exceeding the human health assessment criteria in locations outside of the site. Copper, lead and nickel was detected in groundwater at concentrations exceeding the assessment criteria on and offsite.</p> <p>Groundwater was identified at depths ranging 2.5-5 metres bgl and was inferred to flow to the north toward Parramatta River.</p> <p>Jacobs reviewed acid sulfate soil mapping and a report from a nearby site (1-3 Leeds St). From this Jacobs concluded that an acid sulfate soil management plan would be required where intrusive works or dewatering were to occur.</p>

REPORT	KEY ITEMS
<b>ADE Consulting Group (2020) PFAS Investigation of Rhodes East – Leeds Foreshore Rhodes East Site 1 – Leeds Foreshore</b>	<p>The investigation included the site and area to the east "Site 1 – Leeds Foreshore" and a parcel of land to the south of Leeds St "Site 2 – Education Land".</p> <p>5 soil borings were drilled to a depth of 6 metres bgl and soil samples collected from fill and natural materials.</p> <p>3 existing groundwater monitoring wells sampled.</p> <p>2 surface water samples were collected.</p> <p>The results indicated low levels of Per- and polyfluoroalkyl substances (PFAS) were present in the sampled media.</p>
<b>ADE Consulting Group (2022a) Detailed Site Investigation 25-27 Leeds Street, Rhodes, NSW</b>	<p>ADE completed soil and groundwater sampling across the greater Rhodes East site.</p> <p>The investigation identified hotspots of PFAS within soils in the greater investigation area, but not within the current site footprint.</p> <p>ADE confirmed the presence of acid sulfate soils and recommended the need for an Acid Sulfate Soil Management Plan (ASSMP).</p>
<b>ADE Consulting Group (2023) Acid Sulfate Soil Management Plan 25 – 27 Leeds Street, Rhodes NSW</b>	<p>The ASSMP is specific to the site at 25-27 Leeds St. The ASSMP provides a range of management strategies including excavation and liming to reduce the pH and direct excavation and disposal.</p>

# 6 Preliminary Conceptual Site Model

Based on the information presented in Sections 1-4 of this report, a Conceptual site Model (CSM) has been prepared for the site. The ASC NEPM (2013) defines a CSM as:

***"A representation of site-related information regarding contamination sources, receptors and exposure pathways between those sources and receptors"***

The essential elements of the CSM, as required by the ASC NEPM (2013), include an understanding of:

- Known and potential sources of contamination and contaminants of concern including the mechanism(s) of contamination (e.g., 'top down' spill or sub-surface release from corroded tank or pipe).
- Potentially affected media (soil, sediment, groundwater, surface water, indoor and ambient air) and human and ecological receptors.
- Potential and complete exposure pathways.

A tabular CSM provided in **Table 5** preliminarily identifies complete and potential Source, Pathway, Receptor (SPR) linkages.

## 6.1 Potential Sources of Contamination

The potential sources of contamination identified during the DSI are summarised in **Table 4**.

**Table 4. Potential Sources of Contamination**

SOURCE	AREAS OF CONCERN	CONTAMINANTS OF POTENTIAL CONCERN
<b>Historical use of fill</b>	Potential importation of fill from an unknown source used for reclaiming foreshore.	Metals, TRH, BTEX, PAHs, OCP, OPP, Asbestos, Dixons/furans
<b>Historical land uses including timber milling/treatment,</b>	Entire site	Metals, pesticides
<b>Historical surrounding land use including chemical manufacture/storage</b>	Groundwater migrating onto the site	Pesticides, metals, VOCs, PFAS
<b>Weathering of potentially hazardous building materials</b>	Surface soils surrounding former structure onsite along eastern site boundary.	Asbestos and lead.
<b>Historical use of pesticides</b>	Weed and onsite vermin control.	OCPs and OPPs.

## 6.2 Potentially Affected Media

The potentially affected media at the site includes soil and groundwater.

## 6.3 Potential Receptors

It is understood that the proposed residential development will feature some landscaped areas with accessible soils.

Based on the proposed land use, the potential receptors at and near the site include the following:

- Construction/maintenance workers.
- Future site users.

- Offsite residents.
- Parramatta River.

## 6.4 Potential Transport Mechanisms and Exposure Pathways

Potential transport mechanisms, exposure pathways and receptors relevant to the site may include:

- Direct contact with potentially contaminated soils during near or sub-surface works at the site (i.e., trenching and/or excavation works during construction or by future occupants).
- Inhalation/ingestion of disturbed soil/dust.
- Inhalation of vapours.
- Surface water run-off discharging into stormwater or running off directly to the Parramatta River.
- Groundwater migrating onto the site and leaving the site and mixing with surface water of the Parramatta River.
- Plant uptake/ingestion my microorganisms and biomagnification/bioaccumulation.

## 6.5 Source, Pathway and Receptor Linkages

A preliminary tabular CSM has been prepared for the site based on the outcomes of the desktop assessment in **Table 5** below. The tabular CSM describes potential linkages and assesses each of the linkages as possible or unlikely based on the likelihood of occurrence and availability of data.

**Table 5. Preliminary Exposure Pathway Assessment**

SOURCE	EXPOSURE PATHWAY	RECEPTOR	EXPOSURE
<b>Fill materials of unknown origin.</b>	<ul style="list-style-type: none"> <li>• Direct contact with potentially contaminated soils during sub-surface works at the site</li> </ul>	<ul style="list-style-type: none"> <li>• Construction Workers.</li> <li>• Future site users.</li> <li>• Maintenance workers<sup>1</sup>.</li> </ul>	<b>Possible</b> The SPR linkage is potentially complete.
<b>Historical land uses including timber milling/treatment</b>	<ul style="list-style-type: none"> <li>• Inhalation of dust and vapours.</li> <li>• Ingestion of mobilised dust.</li> <li>• Surface water run-off.</li> </ul>	<ul style="list-style-type: none"> <li>• Offsite Maintenance workers<sup>1</sup>.</li> <li>• Offsite receptors.</li> </ul>	Assessment of CoPCs in surface and sub-surface soils is warranted.
<b>Historical surrounding land use including chemical manufacture/storage</b>	<ul style="list-style-type: none"> <li>• Groundwater/surface water interaction.</li> <li>• Vapour intrusion into building airspace.</li> </ul>	<ul style="list-style-type: none"> <li>• Maintenance workers<sup>1</sup>.</li> <li>• Construction Workers.</li> <li>• Future site users.</li> <li>• Offsite receptors.</li> </ul>	<b>Possible</b> The SPR linkage is potentially complete. Assessment of CoPCs in groundwater is warranted.
<b>Weathering of potentially hazardous building materials.</b>	<ul style="list-style-type: none"> <li>• Direct contact with potentially contaminated soil during surface and sub-surface works or by futures users of the area.</li> <li>• Inhalation of dust.</li> <li>• Ingestion of potentially contaminated soil.</li> <li>• Surface water run-off.</li> </ul>	<ul style="list-style-type: none"> <li>• Construction Workers.</li> <li>• Future site users.</li> <li>• Maintenance workers<sup>1</sup>.</li> <li>• Offsite receptors.</li> </ul>	<b>Possible</b> The SPR linkage is potentially complete. Assessment of CoPCs in surface and sub-surface soils.
<b>Historical use of pesticides</b>	<ul style="list-style-type: none"> <li>• Direct contact with contaminated soil.</li> </ul>	<ul style="list-style-type: none"> <li>• Future site users.</li> </ul>	<b>Possible</b>

SOURCE	EXPOSURE PATHWAY	RECEPTOR	EXPOSURE
	<ul style="list-style-type: none"><li>• Inhalation dust.</li><li>• Surface water run-off .</li></ul>	<ul style="list-style-type: none"><li>• Construction workers.</li><li>• Maintenance workers<sup>1</sup>.</li><li>• Offsite receptors.</li></ul>	<p>The SPR linkage is potentially complete.</p> <p>Assessment of CoPCs in surface and sub-surface soils.</p>

Notes:

1. Maintenance workers attending the site, particularly if works require excavation, trenching or any activities applicable to the sub-surface of the site.

# 7 Sampling and Analysis Plan

## 7.1 Data Quality Objectives

The Data Quality Objective (DQO) process is a systematic planning tool based on the scientific method for establishing criteria for data quality and for developing data collection designs. The DQO defines the experimental process required to test a hypothesis. The DQO process has been developed to ensure that efforts relating to data collection are cost effective, by eliminating unnecessary, duplicative or overly precise data whilst at the same time, ensuring the data collected is of sufficient quality and quantity to support defensible decision making.

It is recognised that the most efficient way to accomplish these goals is to establish criteria for defensible decision making before data collection begins and develop a data collection design based on these criteria. By using the DQO process to plan the investigation effort, the relevant parties can improve the effectiveness, efficiency and defensibility of a decision in a resource and cost-effective manner.

The DQO process consists of seven steps, which are designed to clarify the study objectives, define the appropriate type of data and specify tolerable levels of potential decision errors. The seven-step DQO process adopted for this DSIR can be summarised as:

- **Step 1: State the Problem** – concisely describe the problem to be studied. Review prior studies and existing information to gain a sufficient understanding to define the problem.
- **Step 2: Identify the Decision** – identify what questions the study will attempt to resolve, and what actions may result.
- **Step 3: Identify the Inputs to the Decision** – identify the information that needs to be obtained and the measurements that need to be taken to resolve the decision statement.
- **Step 4: Define the Study Boundaries** – specify the time periods and spatial area to which decisions will apply. Determine when and where data should be collected.
- **Step 5: Develop a Decision Rule** – define the statistical parameter of interest, specify the action level, and integrate the previous DQO outputs into a single statement that describes the logical basis for choosing among alternative actions.
- **Step 6: Specify Tolerable Limits on Decision Errors** – define the decision maker's tolerable decision error rates based on a consideration of the consequences of making an incorrect decision; and
- **Step 7: Optimise the Design** – evaluate information from the previous steps and generate alternative data collection designs. Choose the most resource-effective design that meets all DQOs.

The DQOs are provided in **Table 6** below and were derived in accordance with Australian Standard 4482.1-2005.

**Table 6. Data Quality Objectives**

ITEM	DETAIL
<b>Step 1: State the problem</b>	The site is proposed to be redeveloped for residential land use. Previous investigations have identified the presence of contaminants of potential concern in soil and groundwater at the site at concentrations that have the potential to pose an unacceptable risk to human health and ecological receptors under the proposed land use. Delineation of the nature and extent of contamination at the site is required to inform future remediation and/or management decisions.

ITEM	DETAIL
<b>Step 2: Identify the decision/goal of the study</b>	If elevated concentrations of CoPCs are identified at the site: <ul style="list-style-type: none"> <li>• What is the extent of the impact?</li> <li>• Does any CoPC at the site occur at concentrations that pose or may pose an unacceptable liability or risk to the environment and/or human health to persons who will utilise the future development?</li> <li>• If so, what is the order of priority to minimise the risk and what additional measures are required to mitigate, remediate, or manage the risk?</li> <li>• Is the site suitable for the proposed land use setting, in the context of land contamination?</li> </ul>
<b>Step 3: Identify the information inputs</b>	Key data required to resolve the project problem included: <ul style="list-style-type: none"> <li>• Site history and setting</li> <li>• Historical soil and groundwater data</li> <li>• Site observations during field works.</li> <li>• The CSM</li> <li>• The proposed land use and development plans</li> <li>• Published Tier 1 assessment criteria.</li> </ul>
<b>Step 4: Define the boundaries of the study</b>	This investigation was restricted to the physical site boundaries, as shown in <b>Figure 2, Appendix A</b> . The vertical extent of the study boundaries was limited to a maximum depth of 12 metres bgl.
<b>Step 5: Develop a decision rule</b>	If the concentrations of CoPC in soil are reported to be below the relevant adopted tier 1 assessment guidelines, then the site will be deemed suitable, and no management/remediation options will be proposed for the proposed land use at the site.  If, however, the concentration of one or more CoPC are greater than the guidelines, then further investigation will be required to horizontally and vertically delineate the extent of the impact and/or recommendations made for the remediation/management of contamination to render the site suitable for the proposed development and future land dedication to council of the foreshore park.

ITEM	DETAIL
<b>Step 6: Specify tolerable limits on decision errors</b>	<p>The laboratory data quality indicators are as follows:</p> <ul style="list-style-type: none"> <li>• Relative Percentage Difference (RPD) for laboratory duplicates for TPH and BTEX analysis is less than 60%; and</li> <li>• Recovery of matrix spikes and surrogate spikes is as per the laboratory's Quality Assurance targets accepted under their NATA accreditation.</li> </ul> <p>Precision is measured using the Standard Deviation (SD) or RPD. Replicate data for field duplicates of organics is expected to be as follows:</p> <ul style="list-style-type: none"> <li>• RPD criteria of 50% or less, for concentrations greater than or equal to 10 times practical quantitation limits (PQL).</li> <li>• RPD criteria of 75% or less, for concentrations between 5 and 10 times the PQL; and</li> <li>• RPD criteria of 100% or less, for concentrations less than 5 times PQL.</li> </ul> <p>Replicate data for field duplicates for inorganics, including metals is expected to be as follows:</p> <ul style="list-style-type: none"> <li>• RPD criteria of 30% or less, for concentrations &gt; or = 10 times PQL.</li> <li>• RPD criteria of 75% or less, for concentrations between 5 and 10 times the PQL; and</li> <li>• RPD criteria of 100% or less, for concentrations &lt; 5 times PQL.</li> </ul> <p>Where acceptable limits for field duplicates are not met, a discussion on low biased error will be provided.</p> <p>For this investigation, a decision error of 5% will be considered acceptable. This error rate is in accordance with Appendix B of Schedule B(2) of the ASC NEPM (2013). In order to achieve this level of confidence, the investigation has been designed as described below.</p>
<b>Step 7: Optimise the design</b>	<p>Sampling locations were selected based on site access restrictions and infilling of areas not previously assessed.</p> <p>Groundwater monitoring wells were located to target upgradient and downgradient locations.</p> <p>Soil samples were collected at relevant intervals, changes in geology or in zones of gross contamination and locations selected for efficient and representative sampling.</p> <p>Soil samples for acid sulfate soil analysis were selected based on depth above and below the proposed excavation extent and also on field observations.</p> <p>All media sampling was completed in accordance with Reditus standard operating procedures and relevant industry guidelines.</p>

## 7.2 Schedule of Works

Fieldworks including borehole drilling, soil sampling, monitoring well installation and well development were completed on 20 – 23 May 2024 by Reditus' Senior Environmental Scientist Renee Ashton.

Groundwater gauging, sampling and GPS surveying were completed on 30 and 31 May 2024 by Reditus' Environmental Scientist Kyle Sier.

## 7.3 Sampling Analysis Plan and Sampling Rationale

The intention of the sampling plan was to attain the objectives stated in **Section 1.3**. To achieve this, a judgemental (targeted) sampling design program was adopted for this investigation. The sampling plan was based on a review of the previous investigations, the site history, the site walkover and NSW EPA (2022) Sampling Design Guidelines, and

targeted the location of potentially contaminating infrastructure, inferred groundwater flow direction, and had the goal of providing sufficient data to allow for definition, assessment and characterisation of soil and groundwater.

The sampling plan, rationale and analysis undertaken is summarised in **Table 7** and locations of soil and groundwater sample locations are presented in **Figure 3, Appendix A**.

**Table 7. Sampling Analysis Plan and Rationale**

SAMPLE LOCATIONS	MEDIA	RATIONALE	DEPTH (m)
NEW LOCATIONS			
<b>BH101 / RMW1</b>	Soil, Groundwater	Data gap in existing dataset in northwest of site and assessment of foreshore area.	6m – estimated depth to top of bedrock
<b>BH102 / RMW2</b>	Soil, Groundwater	Data gap in existing dataset in northwest of site and assessment of foreshore area. Deep well for dewatering assessment	10m – to assess for groundwater dewatering
<b>BH103 / RMW3</b>	Soil, Groundwater	Data gap in existing dataset in centre/north of site and assessment of foreshore area.	6m – estimated depth to top of bedrock
<b>BH104 / RMW4</b>	Soil, Groundwater	Data gap in existing dataset in centre/north of site and assessment of foreshore area. Deep well for dewatering assessment.	10m – to assess for groundwater dewatering
<b>BH105</b>	Soil	Data gap in existing dataset in west of site	6m – estimated depth to top of bedrock
<b>BH106</b>	Soil	Data gap in existing dataset in centre/west of site	6m – estimated depth to top of bedrock
<b>BH107</b>	Soil	Data gap in existing dataset in centre/east of site	6m – estimated depth to top of bedrock
<b>BH108</b>	Soil	Data gap in existing dataset in west of site	6m – estimated depth to top of bedrock
<b>BH109</b>	Soil	Data gap in existing dataset in centre of site	4m – estimated depth to top of bedrock
<b>BH110</b>	Soil	Data gap in existing dataset in central/south of site	4m – estimated depth to top of bedrock
<b>BH111</b>	Soil	Data gap in existing dataset in east of site	4m – estimated depth to top of bedrock
<b>BH112</b>	Soil	Data gap in existing dataset in central/south of site	4m – estimated depth to top of bedrock
<b>BH113</b>	Soil	Data gap in existing dataset on eastern side of site	4m – estimated depth to top of bedrock
<b>BH114</b>	Soil	Data gap in existing dataset in south-eastern corner of site	4m – estimated depth to top of bedrock
<b>BH115 / RMW05</b>	Soil/Groundwater	Upgradient location, deep well for dewatering	10m – to assess for groundwater dewatering

SAMPLE LOCATIONS	MEDIA	RATIONALE	DEPTH (m)
NEW LOCATIONS			
EXISTING LOCATIONS (ADE 2022)			
<b>MW1-B</b>	Groundwater	To obtain current data point	Depth (6.15m) Sample depth: Mid-point of water column in screened interval
<b>MW2-B</b>	Groundwater	To obtain current data point	Depth (11m) Sample depth: Mid-point of water column in screened interval
<b>MW3-B</b>	Groundwater	To obtain current data point	Depth (5.93m) Sample depth: Mid-point of water column in screened interval
<b>MW4-B</b>	Groundwater	To obtain current data point	Depth (5.7m) Sample depth: Mid-point of water column in screened interval

### 7.3.1 SOIL SAMPLING

Soil sampling was completed as follows:

- All soil borings were advanced with a drill rig utilising push tubes to allow for the collection of soil samples, followed by solid flight augers to facilitate well installation (where required).
- To achieve the target depth for the deep monitoring wells, once refusal was met on sandstone the drilling method was changed to coring. Where zones of weathered material were observed, samples were collected in accordance with the method described below.
- Soil samples were generally collected from the surface (0.1 metres), 0.5 metres bgl, 1 metre bgl, and every metre thereafter until the target depth was reached.
- The soil samples were collected directly from disposable push tube sleeves, the auger tip or from weathered zones within the core sample using dedicated nitrile gloves.
- Efforts were made to minimise disturbance of the material being sampled to the extent practicable. Such techniques included taking the sample directly from the auger tip to prevent cross-contamination and minimise the potential for loss of VOCs.
- Each soil sample was described in general accordance with the USCS and details of any discolouration, staining, odours or other indicators of contamination were also noted. Borehole logs are provided in **Appendix D**.
- Part of the soil sample was placed into a snap lock plastic bag for screening with a PID and part being placed directly into a laboratory prepared 250 mL glass jar with the details of the sample, including the sample name, the job number, the date of the sample and the sample depth.

To assess the presence, form and concentration of asbestos in soil, the following methodology was followed:

- Soil samples were collected in accordance with the ASC NEPM (2013) methodology for asbestos gravimetric sampling and submitted for asbestos in soil quantification analysis, which included:
  - A 500 mL soil sample was collected from each test-pit and placed into snap lock plastic bag (double bagged), for laboratory analysis of asbestos fines and friable asbestos (AF and FA).
  - A 10 L sample was collected from the fill material at each test-pit was collected. Each 10 L sample was spread over a contrasting surface (i.e., tarpaulin) and inspected for bonded Asbestos Containing Material (ACM)fragments. Any ACM or FA identified was collected, bagged and submitted to the laboratory for asbestos identification and the weight of asbestos fragments collected.

The percentage soil asbestos from collected ACM fragments was calculated as follows:

$$\%Soil\ Asbestos = \frac{\%Asbestos\ Content \times ACM\ (kg)}{Soil\ Volume\ (L) \times Soil\ Density\ (kg/L)}$$

where:

- % Asbestos Content by weight (within asbestos cement materials) = 10% to 15% (Reditus has adopted 15% for the sake of conservatism)
- Soil Density (for sand material) = 1.65 kg/L

### 7.3.2 WELL DEVELOPMENT AND GROUNDWATER SAMPLING

5 of the soil borings were installed to either the top of rock or to 10 metres bgl and converted to groundwater monitoring wells. 2 of the 5 newly installed wells (RMW01 and RMW04) were screened from at least 1 metre above the saturated zone to the bottom of the borehole (totalling approximately 3m). The remaining 3 newly installed wells (RMW02, RMW03 and RMW05) were installed into the deeper sandstone bedrock with a 1 metre screen length. The groundwater monitoring wells were constructed using 50 mm internal diameter, Class 18 flush jointed unplasticised polyvinyl chloride (uPVC) with machine slots of 0.5 mm to 1.0 mm width. 1-2 mm washed graded gravel was then packed from the base of the well up to 0.5 metres above the screen. A minimum 1 metre thick bentonite seal was installed above the gravel pack, with cement grout finishing the well to the surface. Each well was finished with a trafficable Gatic cover.

All monitoring wells were developed using a stainless-steel bailer and 12V groundwater development pump after installation and at least one week before groundwater sampling. The process was used to disturb the water column within the well annulus to remove any groundwater and well debris that may have been introduced since installation. Where practicable, a minimum quantity of three casing volumes of water was removed or until purged dry, whichever came first. The well was then left to stabilise for at least one week prior to purging and sampling.

During the groundwater monitoring event, each groundwater monitoring well was gauged for depth to water, depth to product, and depth to base from the top of casing (TOC) using an oil/water interface probe prior to purging and sampling. If product was identified during gauging, a clear plastic bailer was inserted into the well and the extent, appearance, apparent age and odour of the product was recorded.

Following gauging, each well was then purged using a low-flow peristaltic pump until groundwater quality parameters had stabilised to within the groundwater stabilisation criteria listed in **Table 8**. Physicochemical parameters were monitored using a calibrated HANNA water quality meter placed within a flow cell. Once the parameters were within the stabilisation criteria, the sample was collected.

**Table 8. Groundwater Physicochemical Stabilisation Criteria**

PHYSICOCHEMICAL PARAMETER	PURGING STABILISATION CRITERIA
<b>Dissolved Oxygen</b>	+/- 10%
<b>Electrical Conductivity</b>	+/- 3%
<b>Oxidation-Reduction Potential</b>	+/- 10%
<b>pH</b>	+/- 0.1
<b>Temperature</b>	+/- 10%

Groundwater samples were collected directly into appropriately preserved laboratory supplied sampling containers and labelled with the date, sample name, sampler name and project number. Samples for dissolved metals were field filtered using a 0.45-micron Millipore bell filter.

Fresh high-density polyethylene tubing was used at each monitoring well, and the interface probe was thoroughly decontaminated between gauging events with a phosphate free detergent and rinsed with deionised water followed by potable water.

Field forms from the groundwater monitoring event are provided in **Appendix E**.

### 7.3.3 SURVEYING

The TOC height relative to AHD was obtained for each well using a Real-time kinematic positioning (RTK) GPS (Trimble) to allow for the interpolation of groundwater contours and approximation of groundwater flow direction.

TOC measurements were collected in the GDA2020 MGA Zone 56 coordinate reference system. Where GPS signal was poor, laser level measurements were utilised.

#### 7.3.4 SAMPLING HANDLING

Sample handling and transportation was completed as follows:

- Each sample was collected in laboratory supplied and preserved sample containers.
- The sample containers were filled to ensure no headspace for volatile to off-gas.
- The sample containers were labelled with the project number, sampler, sample location and depth, and sampling date.
- The samples were immediately stored in an ice cooled esky to keep them chilled, prior to being couriered to the laboratory under a signed chain of custody (COC) form filled out with the required analysis (provided in Appendix F).
- The eskies were couriered to the laboratories within holding times.

### 7.4 Laboratory Analysis and Methods

Laboratory analytical methods and analyte PQLs are presented in the analytical laboratory certificates provided in **Appendix F** and were considered appropriate for the quantification of the CoPC identified in the CSM.

### 7.5 Tier 1 Assessment Criteria

Tier 1 assessment involves the comparison of monitoring data to published guideline criteria (typically presented as screening levels). Relevant criteria are selected based on the identified viable exposure pathways and CoPCs and proposed land use.

In Australia, appropriate HILs (including interim HILs for vapour intrusion and, where applicable, HSLs for petroleum hydrocarbons and assessment criteria for asbestos) are used for Tier 1 screening to provide a rapid assessment of whether the site contamination may be of concern with respect to human health. Should contaminant concentrations at a site occur at levels that are below the Tier 1 levels, this implies that for the majority of the people in the population there is no significant health risk from contamination and that remedial action may not be required to protect human health.

Exceedances of the tier 1 HILs should be identified and considered. Tier 1 HIL exceedances do not imply that a risk is necessarily present, but that further assessment may be justified. Tier 1 HILs are not intended to indicate a clear demarcation between acceptable and unacceptable. Marginal exceedances may not require quantitative Tier 2 risk assessment to conclude that further assessment is not necessary. The magnitude of the exceedance should be considered in the context of the CSM (that is, whether the exposure pathways are plausible and whether exposure will result in harm).

Tier 1 screening criteria (including HILs and HSLs) should only be used where there has been adequate characterisation of a site (that is, appropriate representative sampling has been carried out). For this investigation, the maximum reported concentrations for each sample and analyte will be compared against the tier 1 criteria. Should any individual sample exceedance of the tier 1 criteria exist, the 95% Upper Confidence Limit (UCL) of the analyte for the site data set was calculated and compared to relevant Tier 1 screening criteria. However, the implications of localised elevated values should also be considered. In order to adopt the 95% UCL result, the analyte data set must also meet the following criteria:

- The SD of the results should be less than 50% of the Tier 1 screening criteria.
- No single value exceeds 250% of the relevant Tier 1 screening criteria (characterised as a 'hot-spot').

Where site data exceeds the screening levels or suitable screening levels cannot be identified, further consideration (Tier 2 assessment) is required.

The tier 1 assessment criteria were adopted from:

- National Environment Protection Council (NEPC) 1999, 'Schedule B(1) Guideline on Investigation Levels for Soil and Groundwater, National Environment Protection (Assessment of site Contamination) Measure (NEPM), as amended in 2013'.

### 7.5.1 SOIL

The soil assessment criteria (SAC) adopted for this assessment were:

- ASC NEPM (2013) HIL-B criteria values have been adopted to assess concentrations of CoPC in soil for site suitability and human health in a high density residential land use scenario.
- ASC NEPM (2013) HSL-A/B guidelines for vapour intrusion have been adopted to evaluate the risk posed from vapour intrusion in a mixed-use high-density residential land use scenario. The soil HSLs are based on depth of impacts, overlying soil type and land use. After a review of subsurface conditions, the guidelines for sand were conservatively selected.
- ASC NEPM (2013) ESLs and EILs were both selected to determine the risk of potential contamination to identified ecological receptors on the site, in the context of an urban residential/public open space. Both ESL and EIL criteria values are applicable for the first 2 metres of the soil profile.
- ASC NEPM (2013) health screening levels for asbestos contamination in soil, which are based on specific land use exposure scenarios, for three forms of asbestos: bonded ACM, FA and AF. The laboratory method for analysis of asbestos in bulk materials is based on AS 4964-2004. Consequently, a practical quantification limit equal to or less than 0.001% by weight cannot be reported as a NATA accredited limit which is required to compare against the AF/FA assessment criteria. The NATA accredited limit is 0.1g/kg (equivalent to 0.01% w/w). For this assessment both the NATA and non-NATA accredited results have been reported. The health screening level for high density residential land use have been adopted for this assessment
- EIL criteria values were derived using site measured values and the NEPM EIL Interactive Calculation Worksheet available at <http://www.nepc.gov.au/nepms/assessment-site-contamination/toolbox>. The output from the EIL calculation is available in **Appendix G**. The following input parameters were reported from soil samples collected from the site:
  - **Cation exchange capacity (CEC)**: 3 cmol/kg
  - **Clay (%)**: 25 %
  - **pH**: 4.6
  - **Soil organic carbon (OC %)**: 2.8 %
- PFAS assessment criteria were taken from the Heads of Environmental Protection Authorities (HEPA, 2020) PFAS NEMP v2.0 for a residential with minimal access to soil land use scenario.
- In lieu of Australian guidelines, the dioxins and furans were assessed with respect to the tier 1 published criteria presented in CCME (2002).

Soil analytical results and guideline criteria are tabulated in **Table 1, Appendix C**.

### 7.5.2 GROUNDWATER

Selection of the Groundwater Assessment Criteria is based on the proposed development of the site to a residential land use and proximity of the site to identified receptors.

The tier 1 assessment criteria were adopted from:

- ASC NEPM (2013).
- Australian and New Zealand Governments Guidelines for Fresh and Marine Water Quality 2018.
- HEPA (2020)
- National Health and Medical Research Council (NHMRC) Australian Drinking Water Guidelines 6, 2011, Version 3.8 Updated September 2022.

The groundwater analytical data were compared against the following adopted Tier 1 screening criteria:

- ANZG (2018) 95% species protection default guideline values (DGVs) (supersedes the ASC NEPM (2013) Groundwater Investigation Levels (GIL)) for Freshwater.
  - The selection of Marine Water criteria is based on the proximity of the Site to the Parramatta River, which is likely to accept both surface water and potentially groundwater discharge from the Site.

- Use of the 95% protection level assumes that the surrounding watercourses are moderately disturbed ecosystems. This is as defined in Section 3.1 of the guidelines as receiving road and storm water runoff from adjacent industry and residential properties, consistent with the setting and environment surrounding the site.
  - The ANZG (2018) is a revision of the Australian and New Zealand Guidelines for Fresh and Marine Water Quality (ANZECC & ARMCANZ 2000) presented as an online platform, to improve usability and facilitate updates as new information becomes available.
  - Revisions to DGVs since the ANZECC & ARMCANZ (2000) guidelines have been provided for the 'aquatic ecosystem' community value. DGVs have been revised for physical and chemical (PC) stressors based on increased understanding, broader monitoring data collected since 2000.
  - The NEPM ASC (2013) GILs were derived using the ANZECC & ARMCANZ (2000), which were subsequently revised (superseded) in ANZG (2018). As such, the ANZG (2018) DGVs will be used as an initial screening in place of the NEPM ASC (2013) GIL criteria as they provide concentrations which once exceeded require further investigation into receptors and points of discharge.
- HEPA (2020) PFAS NEMP 2.0 Table 1 Recreational Water guideline criteria have been adopted to assess concentrations of PFAS compounds in groundwater which may discharge into the Parramatta River.
  - HEPA (2020) PFAS NEMP 2.0 Table 5, Marine Water 95% species protection - slightly to moderately disturbed systems exposure scenario guideline criteria have been adopted to assess concentrations of PFAS in groundwater which may discharge into Parramatta River and be ingested by ecological aquatic receptors.
  - HEPA (2020) PFAS NEMP 2.0 Table 5, Marine Water 99% species protection – high conservation value systems exposure scenario guideline criteria have been adopted to assess concentrations of PFAS in groundwater to allow for biomagnification.
  - NEPM ASC (2013) Groundwater HSL for vapour intrusion HSL-A&B (GW HSL-A&B) for residential land use with groundwater depth 2-<4m within sand have been adopted to assess vapour intrusion in a residential land use setting. The 'coarse' soil texture has been selected following a review of predominant subsurface conditions.
  - ANZG (2023) Toxicant default guideline values for aquatic ecosystem protection: Dioxins in freshwater (adopted in the absence of marine water guidelines) Toxicant default guideline values, 2,3,7,8-TCDD in freshwater, moderate reliability for 95 % protection of species.

Groundwater is not abstracted at the site or down-gradient for beneficial use such as drinking water or irrigation. As such the respective criteria for these have not been considered further.

# 8 Quality Assurance and Quality Control (QAQC)

## 8.1 Field Quality Assurance

### 8.1.1 DETAILS OF SAMPLING TEAM

Fieldworks including the site inspection and soil sampling were completed by Reditus' personnel who are suitably qualified and experienced in the collection of environmental samples.

### 8.1.2 DECONTAMINATION PROCEDURES

Single use equipment was utilised where possible. Equipment that required decontamination, including the oil-water interface probe, low flow pump were washed in a Liquinox then triple rinsed in clean water.

### 8.1.3 CHAIN OF CUSTODY DETAILS

All samples were transported to the laboratory under a COC. Information on the COC included the sampler, sample identifier, sample matrix, collection date, analyses to be performed, sample preservation method, sample release date and sample received date. COCs are provided in **Appendix F** along with the laboratory reports.

### 8.1.4 FIELD DUPLICATE SAMPLES

#### **Sampling Splitting Techniques**

Soil duplicates and triplicate samples were collected by taking representative samples of the soil at the same depth interval. Due to the potential loss of volatiles, samples were not mixed or homogenised during collection or splitting.

#### **Statement of Duplicate Frequency**

Field duplicates and triplicates were collected at a rate of 1:15 (soil) and 1:10 (groundwater). This complies with the Australian Standard 4482.1-2005 and Reditus' QA frequency ranges.

The following soil QA/QC samples were collected:

- DUP02 and TRIP02 were respectively soil intra-laboratory and inter-laboratory duplicates of sample BH102\_2m.
- DUP03 and TRIP03 were respectively soil intra-laboratory and inter-laboratory duplicates of sample BH107\_3m.

The following groundwater QA/QC samples were collected:

- DUP1 and TRIP1 were respectively soil intra-laboratory and inter-laboratory duplicates of sample MW1-B.
- DUP2 and TRIP2 were respectively soil intra-laboratory and inter-laboratory duplicates of sample EW1.

### 8.1.5 TRIP BLANKS AND TRIP SPIKES

Trip spikes are used to assess contamination from field conditions at the time of the sampling event and that the sampling procedures are adequate in preventing cross contamination of VOCs during sample transport and storage.

The results of trip spikes and trip blanks were within acceptable ranges as shown in **Appendix F**.

### 8.1.6 RINSATE BLANKS

Rinsate blanks are used to assess the potential for cross contamination between sampling to ensure decontamination procedures are adequate throughout the investigation.

The results of rinsate sampling were within acceptable ranges and are provided in **Appendix F**.

## 8.2 Laboratory QA/QC

The following is a summary of the internal laboratory QA/QC procedures. Detailed laboratory QA/QC is provided in **Appendix F**.

### 8.2.1 SAMPLE HOLDING TIMES

All holding times were reported as being within specified ranges with the exception of the chromium suite. As such the acid sulfate soil data will be seen as an estimate of the true value. It is noted that the Scr results exceeded the criteria and therefore the management decisions for the site are unaffected by the holding time non-conformance.

### 8.2.2 LABORATORY ACCREDITATION AND ANALYTICAL METHODS USED

The primary laboratory used for samples was Envirolab Services Pty Ltd (Envirolab), while the secondary laboratory was Australian Laboratory Services Pty Ltd (ALS).

Envirolab is accredited by NATA to ISO/IEC 17025 with the accreditation number 1261, while ALS is also accredited to this standard with the accreditation number 825.

### 8.2.3 LABORATORY CONTROL SAMPLES

Laboratory control samples were reported within acceptable ranges.

### 8.2.4 LABORATORY RELATIVE PERCENT DIFFERENCE (RPD)

All internal laboratory RPDs met acceptance criteria with the exception of samples 352254-1 for lead and zinc and 352254-17 which exceeded the criteria for zinc. As such, triplicate results were issued (laboratory sample numbers 352254-42 and 352254-43).

## 8.3 Evaluation of the QA/QC Information Compared to the DQOs

### 8.3.1 DOCUMENTATION COMPLETENESS

- Soil logs, chain-of-custody forms, calibration were complete and appropriate.

Data completeness:

- All samples were received by the laboratories and analytical results reported including laboratory QA/QC.

### 8.3.2 DATA COMPARABILITY

- Reditus standard operating procedures, Australian Standards and industry best practice were followed during sampling.
- Consistent field conditions and similarly trained staff were used during sampling.
- The limits of reporting are appropriate and generally consistent from each laboratory.

### 8.3.3 DATA REPRESENTATIVENESS

- Reditus is confident that cross contamination has not occurred, and primary samples are representative of actual soil conditions.
- The frequency of laboratory blanks was acceptable, and the results were within specified ranges.

### 8.3.4 PRECISION

- Intra-laboratory and inter-laboratory duplicates were collected at the following rates:
  - Soil intra-laboratory and inter-laboratory duplicates frequency were collected at a rate of 1:15 primary samples.
  - Groundwater intra-laboratory and inter-laboratory duplicates frequency were collected at a rate of 1:10 primary samples.
- QA/QC sample collection rate follows the guidance provided in AS4482.1-2005.

### 8.3.5 RELATIVE PERCENT DIFFERENCE

Refer to **Table 4 and Table 5, Appendix B** for Relative Percent Differences (RPDs) calculations. Reditus notes that RPDs were only calculated for groups of compounds with detections above the laboratory detection limits. RPDs for soil were reported within acceptable ranges, with exception to the following:

- BH102\_2m and TRIP02

- Chromium (total) – 40%
- Lead – 32%
- BH107\_3m and TRIP03
  - Copper – 43%
  - Lead – 63%

Reditus considers that these RPD exceedances are likely to be due to the heterogeneity of sampled soil. Reditus hence does not consider that these RPD exceedances have affected the integrity of the analytical results.

All groundwater RPDs were within acceptable ranges.

# 9 Results

## 9.1 Field Observations

### 9.1.1 SOIL

Soil samples collected from the boreholes were logged in general accordance with the USCS. Borehole logs are provided as **Appendix D**. The soils encountered during the intrusive investigation are summarised below:

#### Fill

- Fill material was encountered at depths up to 2.5 metres bgl and comprised gravelly sand and silty sand. The greatest depth of fill was observed in the northern portion of the site (i.e. the foreshore area).

#### Natural

- Natural material comprised silty sand and sandy clay.
- Estuarine mud with shell inclusions was observed in bores drilled on the northern side of the site (foreshore area) at depths of 3 metres bgl.
- Weathered sandstone was observed at all locations on top of competent sandstone bedrock.
- Depth to rock ranged between 1.6 metres bgl in the south of the site to 7 metres bgl in the north.

### 9.1.2 GROUNDWATER

Groundwater was encountered in soil borings at depths greater than 3 metres bgl in residual soils and in bedrock.

Groundwater field sheets are included in **Appendix E**. A summary of the results is provided below:

**Table 9. Stabilised Groundwater Physicochemical Parameters**

ID	pH	TEMP (°C)	EC (mS/cm)	DO (ppm)	REDOX (mV)
Shallow Wells					
<b>RMW01*</b>	4.33	21.1	17.58	3.78.	-122.5
<b>RMW03*</b>	4.35	21.60	21.82	5.73	212.1
<b>EW1</b>	5.66	21.4	7.65	4.31	23.3
<b>MW1-B</b>	6.26	22.2	7.23	3.30	-118.4
<b>MW2-B</b>	6.25	20.8	10.76	3.64	-147.6
<b>MW3-B*</b>	6.74	21.3	38.10	3.91	91.6
<b>MW4-B</b>	4.63	21.4	0.603	3.14	91.7
Deep Wells					
<b>RMW02* (deep)</b>	4.18	19.9	17.70	4.33	-132.1
<b>RMW04* (deep)</b>	5.58	22.1	12.53	5.02	48.3
<b>RMW05 (deep)</b>	5.62	21.2	4.816	4.55	55.9

\*located in the foreshore area

The following can be seen from the above:

- The water in the shallow and deep wells is acidic.
- Depending on location at the site, the groundwater is fresh (MW4-B) to brackish. The EC at MW3-B is representative of seawater.

- The DO indicates well oxygenated water in the shallow and deep wells.
- The redox values indicate variable reducing and oxidizing environments depending on the location at the site.

There does not appear to be a difference in physico-chemical parameters between the shallow and deep wells which may indicated interconnectivity between the water bearing zones.

Groundwater was observed at depths between 1.8 metres bgl and 4.3 metres bgl and was inferred to flow to the north/northeast (see **Figure 4, Appendix A**).

## 9.2 Analytical Results

### 9.2.1 SOIL – CONTAMINATION ASSESSMENT

A summary of the soil analytical results can be found in **Table 1, Appendix C**. The following section outlines the key findings of the comparison between laboratory results and the site assessment criteria.

Soil sample locations are presented on **Figure 3, Appendix A**.

**Table 10. Soil Analytical Data Summary**

ANALYTE	NUMBER OF RESULTS	NUMBER OF DETECTS	MINIMUM CONCENTRATION (mg/kg)	MAXIMUM CONCENTRATION (mg/kg)	EXCEEDANCES
<b>BTEXN</b>					
<b>Benzene</b>	30	0	<0.2	<0.2	None
<b>Toluene</b>	30	0	<0.5	<0.5	None
<b>Ethylbenzene</b>	30	0	<0.5	<1	None
<b>Xylene Total</b>	30	0	<0.5	<1	None
<b>Naphthalene</b>	30	0	<0.1	<0.5	None
<b>TRH</b>					
<b>F1</b>	30	0	<10	<25	None
<b>F2</b>	30	0	<50	<50	None
<b>F3</b>	30	1	<100	230	None
<b>F4</b>	30	0	<100	<100	None
<b>PAH</b>					
<b>BaP</b>	30	2	<0.05	<0.5	None
<b>BaP TEQ</b>	30	2	<0.5	1.2	None
<b>PAHs (total)</b>	28	4	<0.05	0.4	None
<b>Metals</b>					
<b>Arsenic</b>	32	17	<4	21	None
<b>Cadmium</b>	32	0	<0.4	<1	None
<b>Chromium (III+VI)</b>	32	32	6	36	None

ANALYTE	NUMBER OF RESULTS	NUMBER OF DETECTS	MINIMUM CONCENTRATION (mg/kg)	MAXIMUM CONCENTRATION (mg/kg)	EXCEEDANCES
<b>Copper</b>	32	31	1	470	2 x EIL BH108 (0.3m), BH113 (0.5m)
<b>Lead</b>	32	32	2	220	None
<b>Mercury</b>	32	0	<0.1	<0.1	None
<b>Nickel</b>	32	25	<1	22	3 x EIL BH108 (0.3m), BH109 (0.3m), BH111 (0.5m)
<b>Zinc</b>	32	29	<1	600	1 x EIL BH113 (0.5m)
<b>Pesticides</b>					
<b>OCP</b>	13	0	<0.1	<0.1	None
<b>OPP</b>	13	0	<0.1	<0.1	None
PCBs (Sum of total)	13	0	<0.1	<0.1	
<b>PFAS</b>					
<b>PFOA</b>	10	0	<0.0001	<0.0001	None
<b>PFOS</b>	10	1	<0.0001	0.0002	None
<b>PFHxS</b>	10	0	<0.0001	<0.0001	None
<b>PFHxS+PFOS</b>	10	1	<0.0001	0.0002	None
<b>Asbestos</b>					
<b>Asbestos (&lt;2mm AF/FA)</b>	12	0	<0.001	<0.001	None
<b>Asbestos (ACM &gt;7mm)</b>	12	0	<0.01	<0.01	None
<b>Total Asbestos</b>	12	0	<0.1	<0.1	None
Dioxins/Furans TEQ (pg/g)	6	0	0	3.178	None

### 9.2.2 SOIL – ACID SULFATE SOIL ASSESSMENT

A summary of the soil ASS analytical results can be found in **Table 2, Appendix C**. The following section outlines the key findings of the comparison between laboratory results and the site assessment criteria.

Soil sample locations are presented on **Figure 3, Appendix A**.

**Table 11. Acid Sulfate Soil Data**

FIELD ID	DEPTH (m bgl)	NET ACIDITY (ACIDITY UNITS) MOLE H <sup>+</sup> /T	NET ACIDITY (SULFUR UNITS) %S
ACID SULFATE SOIL MANAGEMENT LIMITS (ASSMAC 1998)		18	0.03
<b>BH102</b>	<b>4</b>	500	0.80
<b>BH102</b>	<b>6</b>	150	0.24
<b>BH103</b>	<b>4</b>	<5	<0.005
<b>BH103</b>	<b>6</b>	30	0.050
<b>BH105</b>	<b>4</b>	340	0.55
<b>BH106</b>	<b>3</b>	87	0.14
<b>BH106</b>	<b>6</b>	27	0.044
<b>BH107</b>	<b>4</b>	240	0.38
<b>BH109</b>	<b>2</b>	<5	0.0060
<b>BH114</b>	<b>1</b>	110	0.18
<b>BH115</b>	<b>5.1</b>	22	0.040
<b>BH115</b>	<b>10.7</b>	130	0.21

It can be seen from the table above that ASS is present in 10 of the 12 samples analysed ranging in depth from 1 metre bgl to 10.7 metres bgl. Therefore, an ASSMP is required to be prepared and implemented during the excavation and dewatering works for the proposed development.

### 9.2.3 GROUNDWATER

A summary of the soil analytical results can be found in **Table 3, Appendix C**. The following section outlines the key findings of the comparison between laboratory results and the site assessment criteria.

Soil sample locations are presented on **Figure 3, Appendix A**.

**Table 12. Groundwater Analytical Data Summary**

ANALYTE	NO SAMPLES	NO. DETECTS	MIN (ug/L)	MAX (ug/L)	EXCEEDENCES
<b>BTEX</b>					
<b>Benzene</b>	10	0	<1	<1	None
<b>Toluene</b>	10	1	<1	46	None
<b>Ethylbenzene</b>	10	0	<1	<1	None
<b>m+p-xylene</b>	10	0	<2	<2	None
<b>o-xylene</b>	10	0	<1	<1	None
<b>Naphthalene</b>	10	0	<1	<1	None
<b>TRH</b>					



ANALYTE	NO SAMPLES	NO. DETECTS	MIN (ug/L)	MAX (ug/L)	EXCEEDENCES
<b>F1</b>	10	2	<10	50	None
<b>F2</b>	10	0	<50	<50	None
PAH					
<b>Benzo(a)pyrene</b>	10	0	<0.1	<0.1	None
Metals					
<b>Arsenic-Dissolved</b>	10	6	<1	9	None
<b>Cadmium-Dissolved</b>	10	4	<0.1	1.1	None
<b>Chromium-Dissolved</b>	10	3	<11	3	None
<b>Cobalt-Dissolved</b>	5	5	25	92	5 x Marine: RMW01, RMW02, RMW03, RMW04, RMW05
<b>Copper-Dissolved</b>	10	4	<1	120	3 x Marine: MW4-B, RMW02, RMW03
<b>Lead-Dissolved</b>	10	3	<3	44	2 x Marine: RMW02, RMW03
<b>Mercury-Dissolved</b>	10	0	<0.05	<0.05	None
<b>Nickel-Dissolved</b>	10	8	<1	130	3 x Marine: EW1, RMW03, RMW05
<b>Zinc-Dissolved</b>	10	10	4	620	8 x Marine: EW1, MW1-B, MW3-B, RMW01, RMW02, RMW03, RMW04, RMW05
PFAS					
<b>PFHxS</b>	7	5	<0.0002	0.068	None
<b>PFOS</b>	7	6	<0.0002	0.013	6 x 99% LoP: MW1-B, MW2-B, MW3-B, MW4-B, RMW01, RMW04

ANALYTE	NO SAMPLES	NO. DETECTS	MIN (ug/L)	MAX (ug/L)	EXCEEDENCES
<b>PFOA</b>	7	6	<0.0002	0.018	None
<b>PFHxS+PFOS</b>	7	6	<0.0002	0.071	None
Dioxins/Furans TEQ (pg/L)	3	3	ND	0.036	None

### 9.3 Extent of Uncertainties in the Results

The sampling methodologies used by Reditus during this investigation has been designed to limit uncertainty in the results. Reditus is confident that the results of this investigation give an accurate representation of the current status of the site but note that in all subsurface investigations the potential remains for variability between sampling points and for conditions to be different onsite from the conditions reported herein.

## 10 Discussion

The previous investigations completed at the site did not identify the presence of contamination in soil however the sampling density was less than required by NSW EPA (2022). The soil sampling completed as part of the current investigation increased the sampling density to meet the requirements of NSW EPA (2022), i.e. 23 locations for the site area of 1.16ha.

Observations made during the current site works indicated:

- The current land use has limited potential to impact soil and groundwater.
- The soil profile observed was consistent with previous assessments. The depth to sandstone rock was shallow (<1 metres bgl) in the southern portion, tending deeper to the north and was deepest on the northern boundary in areas known to be reclaimed land (>6 metres bgl).
- The depth to groundwater was consistent with previous investigations and was identified on average at 2.5 metres bgl below ground level and flowed to the north toward the Parramatta River.

The soil analytical data shows the following:

- The concentrations of all CoPC in soil were less than the human health assessment criteria, indicating the site is suitable for the proposed residential land use and the foreshore area is suitable for to be dedicated to council (from a contamination perspective).
- Exceedances to ecological assessment criteria were observed in the basement area, however these will be removed during excavation of the basement.
- Asbestos was not detected in any sample analysed.
- Acid sulfate soils are present at the site which will require management during excavation and dewatering.

The groundwater analytical data shows the following:

- Risks to human health from a vapour intrusion pathway are not present onsite.
- CoPC including PAH, BTEX, TRH, and PFAS were not detected at concentrations exceeding human health assessment criteria.
- PFOS was detected at concentrations exceeding the interim marine 99% level of protection assessment criteria across the site including the foreshore area. This indicates a potential risk to ecological receptors where groundwater and surface water interface. However, given the significant dilution expected this is unlikely to pose an unacceptable risk.
- The greatest PFOS concentrations were observed in the southeast (Leeds St side) and northeast (foreshore area) corners of the site and decreased by two orders of magnitude at the western and northern (foreshore) boundaries of the site. This indicates source of PFOS is not related to the site and is likely to be related to offsite uses to the east. Refer to concentration contour provided in **Figure 5, Appendix A**.
- Heavy metals including copper, lead, nickel and zinc were detected at concentrations exceeding ecological criteria. As for PFAS, the potential exists for ecological receptors of the Parramatta River to be impacted from the groundwater. Dilution of the groundwater at the surface water interface is likely to reduce the risk to ecological receptors. The source of heavy metals is from an upgradient off-site receptor and not attributed to current or historical land use.

Refer to **Figure 6a & 6b, Appendix A**, showing soil and groundwater exceedances respectively from Reditus's investigation, and **Figure 7, Appendix A**, for groundwater exceedances from previous investigations.

# 11 Refined Conceptual Site Model

Based on the results of the investigation and the preliminary CSM presented in **Section 5** has been refined to identify complete and potential pathways between the known or potential source(s) and the receptor(s).

**Table 13. Refined Conceptual Site Model**

SOURCE	EXPOSURE PATHWAY	RECEPTOR	EXPOSURE
<b>Fill materials of unknown origin.</b>	<ul style="list-style-type: none"> <li>Direct contact with potentially contaminated soils during sub-surface works at the site</li> <li>Inhalation of dust and vapours.</li> <li>Ingestion of mobilised dust.</li> <li>Surface water run-off.</li> </ul>	<ul style="list-style-type: none"> <li>Construction Workers.</li> <li>Future site users.</li> <li>Maintenance workers<sup>1</sup>.</li> <li>Offsite Maintenance workers<sup>1</sup>.</li> <li>Offsite receptors.</li> </ul>	<b>Unlikely</b> Soil results indicate that a source of significant soil contamination is not present.
<b>Historical land uses including timber milling/treatment</b>			<b>Possible</b> The SPR linkage is potentially complete if an exposure path to groundwater is created during future site development. The potential exists for groundwater to contaminate receiving waters, however dilution at the interface is likely to significantly reduce the concentrations and therefore reduce the risk to receptors.
<b>Historical surrounding land use including chemical manufacture/storage</b>	<ul style="list-style-type: none"> <li>Groundwater/surface water interaction.</li> <li>Vapour intrusion into building airspace.</li> </ul>	<ul style="list-style-type: none"> <li>Maintenance workers<sup>1</sup>.</li> <li>Construction Workers.</li> <li>Future site users.</li> <li>Offsite receptors.</li> </ul>	
<b>Weathering of potentially hazardous building materials.</b>	<ul style="list-style-type: none"> <li>Direct contact with potentially contaminated soil during surface and sub-surface works or by futures users of the area.</li> <li>Inhalation of dust.</li> <li>Ingestion of potentially contaminated soil.</li> <li>Surface water run-off.</li> </ul>	<ul style="list-style-type: none"> <li>Construction Workers.</li> <li>Future site users.</li> <li>Maintenance workers<sup>1</sup>.</li> <li>Offsite receptors.</li> </ul>	<b>Unlikely</b> CoPC associated with hazardous building materials such as asbestos and lead were not detected in soil at concentrations exceeding the land use criteria.
<b>Historical use of pesticides</b>	<ul style="list-style-type: none"> <li>Direct contact with contaminated soil.</li> <li>Inhalation dust.</li> <li>Surface water run-off .</li> </ul>	<ul style="list-style-type: none"> <li>Future site users.</li> <li>Construction workers.</li> <li>Maintenance workers<sup>1</sup>.</li> <li>Offsite receptors.</li> </ul>	<b>Unlikely</b> Soil results indicate that a source of significant soil contamination is not present.

Notes:

Maintenance workers attending the site, particularly if works require excavation, trenching or any activities applicable to the sub-surface of the site.

# 12 Conclusions and Recommendations

## 12.1 Conclusions

Based on a review of the site history, observations made during fieldwork, results of laboratory analysis and the proposed land use scenario, Reditus concludes the following:

- Fill materials at the site are suitable to remain onsite, from a contamination perspective, for the proposed residential land use.
- Residual natural soils are also suitable for the proposed residential land use. However, excavation of natural material and/or dewatering will require management of acid sulfate soils.
- Exceedances to the ecological criteria in soil are not seen to be an issue as this material will be removed during excavation of the basement.
- The groundwater data was consistent with previous investigations which identified heavy metals, namely copper, lead, nickel, and zinc, and PFOS at concentrations exceeding ecological criteria. The presence of metals in groundwater appears to be a regional issue given the presence in up-gradient wells and the lack of source areas on the site. PFOS in groundwater also appears to be related to an offset source given the reduction in concentrations by over two orders of magnitude across the site in the direction of groundwater flow.
- The findings of this investigation were consistent with previous assessments completed at the site.

**Based on the above, Reditus considers that the site is suitable for the proposed development (mixed use high density residential with basement carparking) and the northern section (the foreshore area within the site boundary) is suitable to be dedicated to Council for use as a park.**

**The scope of work completed during this DSI has satisfied Items 13, 16, and 17 of the SEARs and Item 3 of the SEARs cover letter.**

Given the elevated concentrations of heavy metals and PFOS in groundwater it is recommended that groundwater not be abstracted for beneficial use at the site. As dewatering will be required to facilitate the basement excavation a dewatering management plan and an acid sulfate soil management plan will be required to ensure protection of the surrounding environment during the works. If excavation and/or groundwater extraction is required within the foreshore area of the site, the same controls will be required.

**Reditus considers that the consent authority may be satisfied that the required considerations of Clause 4.6 of State Environmental Planning Policy (Resilience and Hazards) 2021 are satisfied for the following reasons:**

- This DSI (Reditus) and previous investigations (Jacobs (2016) and ADE (2022)) completed on the development site has adequately assessed the site history to identify potential sources of contamination, and has suitability assessed the potential risk posed by contaminants to health and the environment. As such, Clause 4.6(1)(a) of SEPP (Resilience and Hazards) 2021 has been successfully achieved.
- This DSI has determined that the land can be made suitable in its current state for the purposes for which the development is proposed to be carried out (as mixed use high density residential). As such, Clause 4.6(1) (b) and (c) of SEPP (Resilience and Hazards) 2021 have been successfully achieved.
- The DSI has been completed in accordance with the contaminated land planning guidelines, satisfying Clause 4.6(2) and 4.6(3) of SEPP (Resilience and Hazards) 2021.



## 13 Limitations

This report has been prepared in accordance with the scope of services described in the **Section 1.3**. The letter has been prepared for the sole use of the client and has been prepared in accordance with a scope of work agreed by the client.

The report or document does not purport to provide legal advice, and any conclusions or recommendations made should not be relied upon as a substitute for such advice.

The report does not constitute a recommendation by Reditus for the client or any other party to engage in any commercial or financial transaction and any decision by the client or other party to engage in such activities is strictly a matter for the client.

The report relies upon data, surveys, measurements and results taken at or under the Site at particular times and conditions specified herein. Any findings, conclusions or recommendations only apply to the aforementioned circumstances and no greater reliance should be assumed or drawn by the client. Furthermore, the report has been prepared solely for use by the client and Reditus accepts no responsibility for its use by other parties. The client agrees that Reditus' report or associated correspondence will not be used or reproduced in full or in part for promotional purposes and cannot be used or relied upon by any other individual, party, group or company in any prospectus or offering. Any individual, party, group or company seeking to rely on this report cannot do so and should seek their own independent advice.

No warranties, express or implied, are made. Subject to the scope of work undertaken, Reditus assessment is limited strictly to identifying typical environmental conditions associated with the subject property based on the scope of work and testing undertaken and does not include and evaluation of the structural conditions of any buildings on the subject property or any other issues that relate to the operation of the Site and operational compliance of the Site with state or federal laws, guidelines, standards or other industry recommendations or best practice. Scope of work undertaken for assessments are agreed in advance with the client and may not necessarily comply with state or federal laws or industry guidelines for the type of assessment conducted.

Additionally, unless otherwise stated Reditus did not conduct soil, air or wastewater analyses including asbestos or perform contaminated sampling of any kind. Nor did Reditus investigate any waste material from the property that may have been disposed off-site or undertake and assessment or review of related site waste management practices.

The results of this assessment are based upon (if undertaken as part of the scope work) a site inspection conducted by Reditus personnel and/or information from interviews with people who have knowledge of site conditions and/or information provided by regulatory agencies. All conclusions and recommendations regarding the property are the professional opinions of the Reditus personnel involved with the project, subject to the qualifications made above.

While normal assessments of data reliability have been made, Reditus assumes no responsibility or liability for errors in any data obtained from regulatory agencies, statements from sources outside of Reditus, or developments resulting from situations outside the scope of this project/assessment.

Reditus is not engaged in environmental auditing and/or reporting of any kind for the purpose of advertising sales promoting, or endorsement of any client's interests, including raising investment capital, recommending investment decisions, or other publicity purposes. Reditus assumes no responsibility or liability for errors in any data obtained from regulatory agencies, statements from sources outside of Reditus, or developments resulting from situations outside the scope of this project.

In relation the conduct of asbestos inspections or the preparation of hazardous materials reports Reditus has conducted inspections and the identification of hazardous material within the constraints presented by the property. Whilst efforts are made to access areas not normally accessed during normal use of the Site to identify the presence of asbestos or other hazardous material, unless explicitly tested no guarantee can be provided that such material is or is not present.

Reditus' professional opinions are based upon its professional judgment, experience, and training. These opinions are also based upon data derived from the limited testing and analysis described in this report or reports reviewed. It is possible that additional testing and analysis might produce different results and/or different opinions or other opinions. Reditus has limited its investigation(s) to the scope agreed upon with its client. Reditus believes that its opinions are reasonably supported by the testing and analysis that has been undertaken (if any), and that those opinions have been developed according to the professional standard of care for the environmental consulting profession in this area at this time. Other opinions and interpretations may be possible. That standard of care may



change and new methods and practices of exploration, testing and analysis may develop in the future, which might produce different results.

# 14 References

## Previous Investigations

- Jacobs (2016) Rhodes East Priority Investigation Area Contamination and Acid Sulphate Soils Report.
- ADE Consulting Group (2020) PFAS Investigation of Rhodes East – Leeds Foreshore Rhodes East Site 1 – Leeds Foreshore
- ADE Consulting Group (2023) Detailed Site Investigation 25-27 Leeds Street, Rhodes, NSW
- ADE Consulting Group (2023) Acid Sulfate Soil Management Plan 25 – 27 Leeds Street, Rhodes NSW

## Environmental Planning

- NSW Environmental Planning and Assessment Act (the EP&A Act 1979).
- NSW State Environmental Planning Policy Number (SEPP) Resilience and Hazards 2021. Superseding SEPP55 – Remediation of Land, 1998.

## Site Contamination

- CCME (2002) Canadian Soil Quality Guidelines for the Protection of Environmental and Human Health Polychlorinated Dibenz-P-Dioxins and Polychlorinated Dibenzofurans (PCDD/Fs)
- CRC Care (2011) *Health screening levels for petroleum hydrocarbons in soil and groundwater. Part 1: Technical development document*.
- NSW Contaminated Land Management Act (the CLM Act 1997).
- NSW EPA statutory guidelines made or approved under section 105 of the CLM Act, including:
- NSW EPA Guidelines for the NSW Site Auditor Scheme (3rd Edition), 2017.
  - NSW EPA Guidelines for Consultants Reporting on Contaminated Land, 2020.
  - NSW EPA Sampling Design Guidelines, August 2022.
  - NSW EPA Guidelines for the Assessment and Management of Groundwater Contamination, 2007.
  - NSW EPA Guidelines for Assessment and Management of Hazardous Ground Gases, 2020.

National Environment Protection Council (1999, Revised 2013) National Environment Protection (Assessment of Site Contamination) Amendment Measure 2013 – Schedule B1 Guideline on Investigation levels for Soil and Groundwater (NEPC, 2013).

Australian and New Zealand Governments Guidelines for Fresh and Marine Water Quality 2018.

- ANZG (2023) Toxicant default guideline values for aquatic ecosystem protection: Dioxins in freshwater (adopted in the absence of marine water guidelines).

Australian Standard AS4482.1-2005. Guide to the Investigation and Sampling of sites with Potentially Contaminated Soil. Part 1: Non-volatile and Semi-volatile Compounds, 2005.

Australian Standard AS4482.2-1999. Guide to the Investigation and Sampling of sites with Potentially Contaminated Soil. Part 2: Volatile Substances, 1999.

Heads of Environment Protection Authorities (2020) PFAS National Environmental Management Plan version 2.0 (the PFAS NEMP 2.0), 2020.

## Acid Sulfate Soils

The Acid Sulfate Soil Management Advisory Committee (ASSMAC) Acid Sulfate Soils Assessment Guidelines 1998 (Also referred to as the "Acid Sulfate Soils Manual").

Soil Conservation Service of NSW "Sydney Soil Landscape Series Sheet 9130, 1989.

Geological Survey of NSW "Sydney Geological Series Sheet 9130, 1:100,000 scale" map (First Edition 1983)".

Geoscience Australia "Hydrogeology Map of Australia", 1987.



## **Waste**

*NSW Protection of the Environment Operations (Waste) Regulations 2014.*

*NSW EPA Waste Classification Guidelines, Part 1 Classifying Waste, 2014.*

*NSW EPA Resource Recovery Order, Excavated Natural Material Order under Part 9, Clause 93 of POEO Waste Regulation 2014 (the ENM Order 2014).*

## **Asbestos**

*NSW Work Health and Safety Regulations, 2017 (WHS Reg 2017), Chapter 8 Asbestos, 2017 (NSW WHS Reg 2017).*

*NSW EPA Managing Asbestos in or on Soil, 2014 (NSW EPA 2014).*

*Western Australia Department of Health Guidelines for the Assessment Remediation and Management of Asbestos Contaminated Sites in Western Australia 2009 (WA DoH, 2009) as referred to by NEPM 2013*

# A

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## Figures

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**REDITUS**



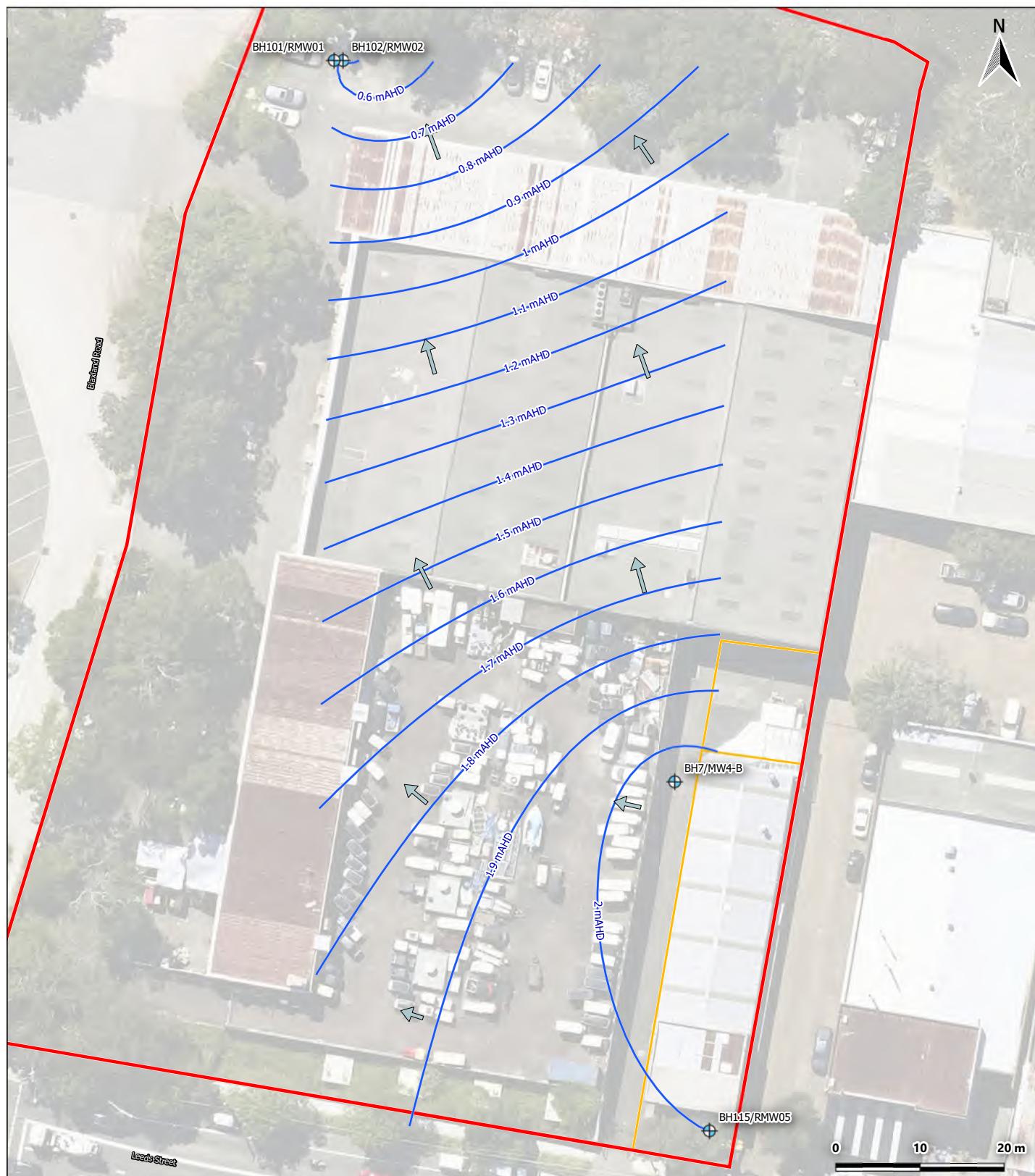
Map		<b>Legend</b>	<b>Watercourses</b>		<b>Figure 1 - Site Location</b>
22148_rp01_f01_siteloc_v01	Date of Export 07/06/2024				
Author JP	Approver TS		Rivers		25-27 Leeds Street, Rhodes NSW 2138
Data Source	Metromap, Google Maps, Open Street Map, NSW Government		Stream (Perennial)		22148 - Detailed Site Investigation
			Stream (Non-Perennial)		
			Unnamed Stream (Non-Perennial)		
			Other Channels		



Map		<b>Legend</b>	<b>Figure 2 - Site Layout</b>
Date of Export	Map Scale (approx. at A3)		
19/06/2024	1:650		25-27 Leeds Street, Rhodes NSW 2138
<b>Author</b>	<b>Approver</b>		22148 - Detailed Site Investigation
JP	TS		
<b>Data Source</b>		Billbergia Pty Ltd	
Metromap, Google Maps, Open Street Map, Geoscience Australia			



Map		Legend	Sample Locations - ADE, 2020-2022		Figure 3 - Sample Locations
22148_rp01_f03_samplelocs_v01			Borehole	Groundwater Monitoring Well	
Date of Export	Map Scale (approx. at A3)	Site Boundary	Lot Boundaries	Sample Locations - Reditus, 2024	25-27 Leeds Street, Rhodes NSW 2138
24/06/2024	1:650	Borehole	Groundwater Monitoring Well	Borehole	22148 - Detailed Site Investigation
Author	Approver	● Borehole	● Groundwater Monitoring Well	● Borehole	Billbergia Pty Ltd
JP	TS	● Borehole	● Groundwater Monitoring Well	● Borehole	
Data Source		Sample Locations - Jacobs, 2016			
Metromap, Google Maps, Open Street Map, Geoscience Australia		Borehole	Groundwater Monitoring Well		



Map		<b>Legend</b>
22148_rp01_f04_gwcont_v01		
Date of Export	Map Scale (approx. at A3)	
19/06/2024	1:550	
Author	Approver	
JP	TS	
Data Source		
Metromap, Google Maps, Open Street Map, Geoscience Australia		

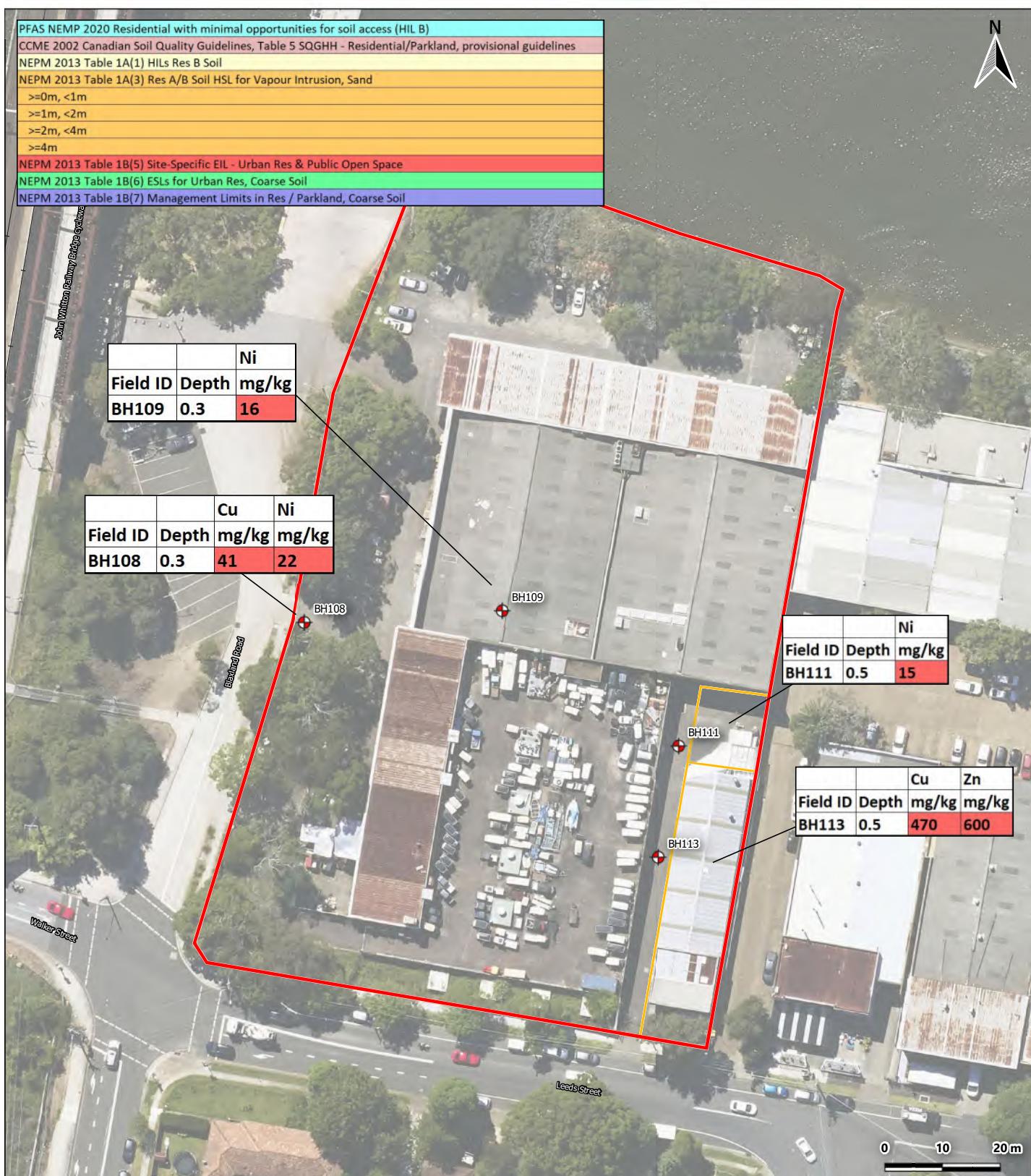
**Figure 4 - Groundwater Elevation Contours**

25-27 Leeds Street, Rhodes NSW 2138

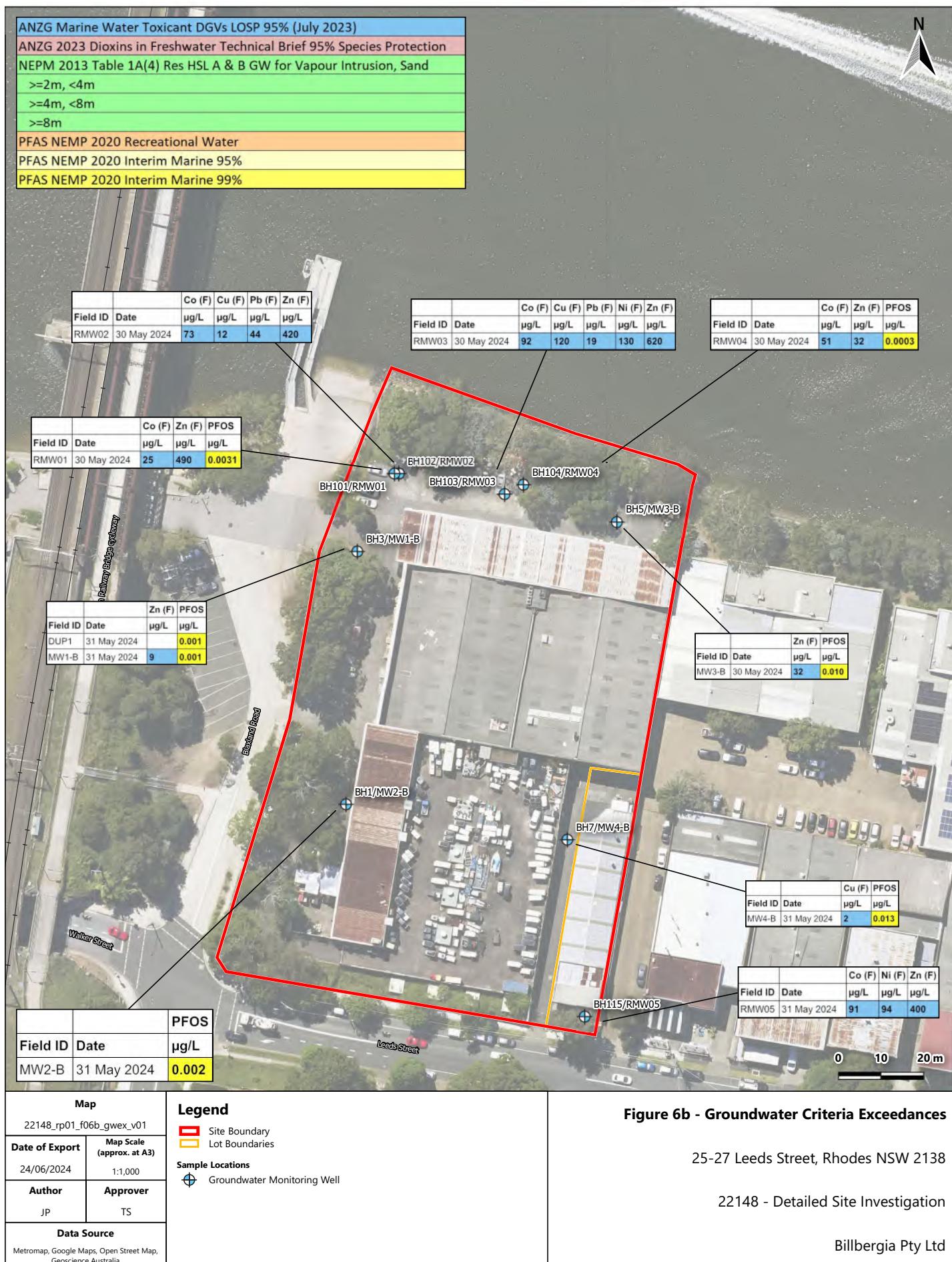
22148 - Detailed Site Investigation

Billbergia Pty Ltd





Map		Legend		Figure 6a - Soil Criteria Exceedances
Date of Export	Map Scale (approx. at A3)			
24/06/2024	1:800			25-27 Leeds Street, Rhodes NSW 2138
Author	Approver			22148 - Detailed Site Investigation
JP	TS			Billbergia Pty Ltd
Data Source				
Metromap, Google Maps, Open Street Map, Geoscience Australia				





Map		Legend	Figure 7 - Historical Groundwater Criteria Exceedances 25-27 Leeds Street, Rhodes NSW 2138 22148 - Detailed Site Investigation Billbergia Pty Ltd
Date of Export	Map Scale (approx. at A3)		
24/06/2024	1:1,000	<ul style="list-style-type: none"> <li>■ Site Boundary</li> <li>■ Lot Boundaries</li> </ul> <p>Sample Locations</p> <ul style="list-style-type: none"> <li>● Groundwater Monitoring Well</li> </ul>	
Author	Approver		
JP	TS		
Data Source			
Metromap, Google Maps, Open Street Map, Geoscience Australia			

# B

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## Photoboard

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**APPENDIX B**  
**SITE PHOTOGRAPHS**

**Report Title**  
Detailed Site Investigation



<b>Client Name</b> Billbergia Pty Ltd	<b>Site Location</b> 25-27 Leeds Street, Rhodes NSW	<b>Project Number</b> 22148
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<b>Photo No.</b> 1	<b>Date</b> 20/5/2024	<b>Direction Facing</b> -	<b>Description</b> Drilling within golf cart workshop	
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<b>Photo No.</b> 2	<b>Date</b> 20/5/2024	<b>Direction Facing</b> -	<b>Description</b> Drilling within golf cart workshop	
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**APPENDIX B**  
**SITE PHOTOGRAPHS**

**Report Title**  
Detailed Site Investigation



**Client Name**  
Billbergia Pty Ltd

**Site Location**  
25-27 Leeds Street, Rhodes NSW

**Project Number**  
22148

Photo No.	Date	
3	20/5/2024	
Direction Facing	N	
Description	Drilling near NW corner of site	

Photo No.	Date	
4	20/5/2024	
Direction Facing	N	
Description	Drilling within driveway of 25 Leeds Street	

**APPENDIX B**  
**SITE PHOTOGRAPHS**

**Report Title**  
Detailed Site Investigation

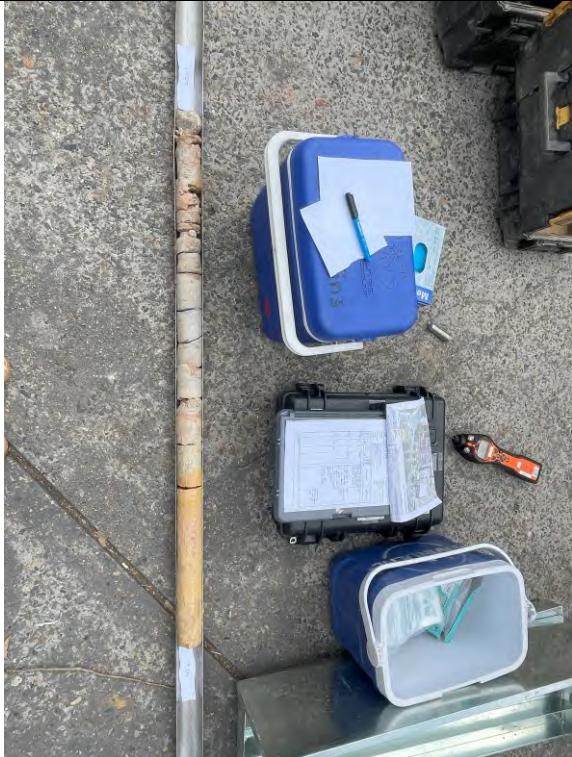


**Client Name**  
Billbergia Pty Ltd

**Site Location**  
25-27 Leeds Street, Rhodes NSW

**Project Number**  
22148

Photo No.	Date	
5	21/5/2024	
Direction Facing	N	
Description	Drilling near NW corner of site	

Photo No.	Date	
6	21/5/2024	
Direction Facing	-	
Description	Sandstone core	

**APPENDIX B**  
**SITE PHOTOGRAPHS**

**Report Title**  
Detailed Site Investigation



**Client Name**  
Billbergia Pty Ltd

**Site Location**  
25-27 Leeds Street, Rhodes NSW

**Project Number**  
22148

Photo No.	Date	
7	21/5/2024	
Direction Facing	-	

Description
Sandstone core

Photo No.	Date	
8	21/5/2024	
Direction Facing	NW	

Description
Monitoring well installation

**APPENDIX B**  
**SITE PHOTOGRAPHS**

**Report Title**  
Detailed Site Investigation



**Client Name**  
Billbergia Pty Ltd

**Site Location**  
25-27 Leeds Street, Rhodes NSW

**Project Number**  
22148

Photo No.	Date	
9	21/5/2024	
<b>Direction Facing</b>		
S		

**Description**  
Drilling within flooring warehouse

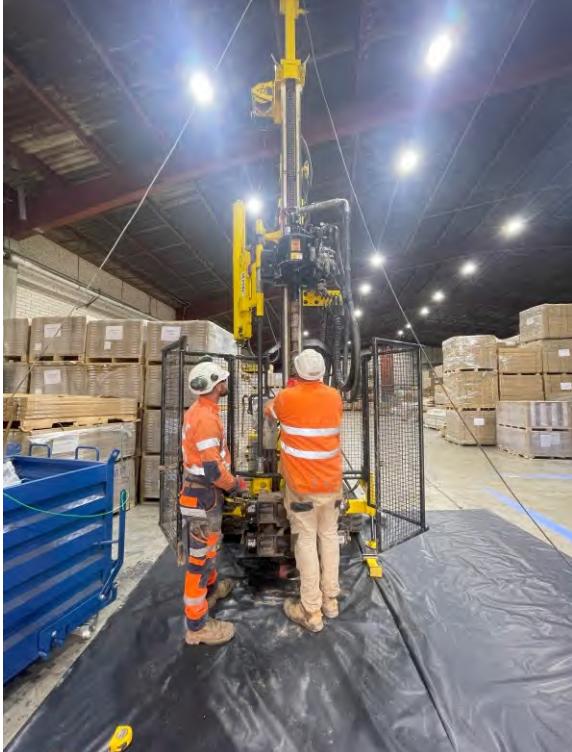


Photo No.	Date	
10	22/5/2024	
<b>Direction Facing</b>		
W		

**Description**  
Drilling towards southern site boundary



**APPENDIX B**  
**SITE PHOTOGRAPHS**

**Report Title**  
Detailed Site Investigation



**Client Name**  
Billbergia Pty Ltd

**Site Location**  
25-27 Leeds Street, Rhodes NSW

**Project Number**  
22148

Photo No.	Date	
11	23/05/2024	
Direction Facing		
NE		
Description		
Drilling at 25 Leeds Street		

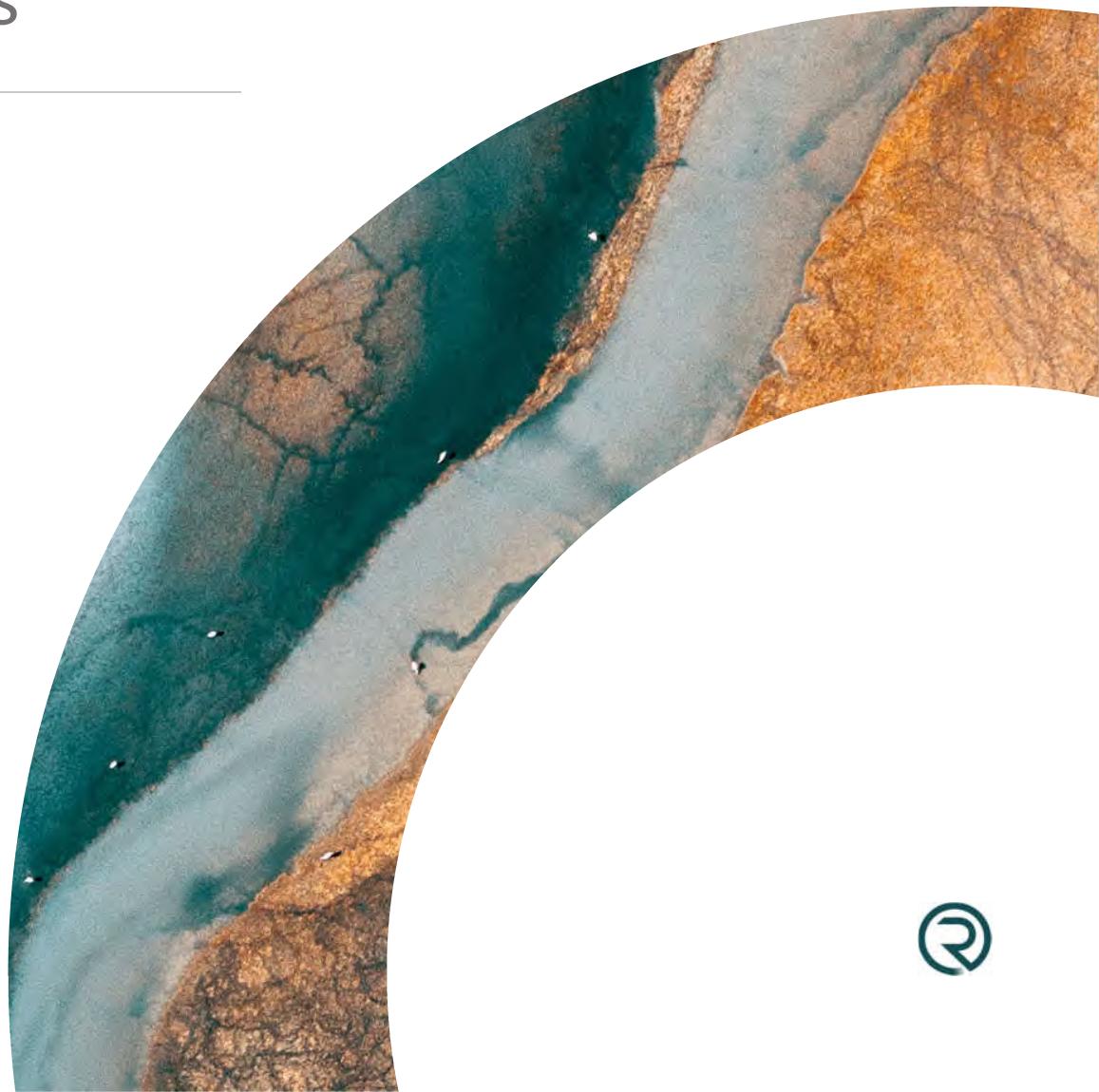
A photograph showing two construction workers in orange high-visibility vests and hard hats operating a yellow hydraulic drilling rig. They are working on a dark-colored brick building with large windows. A blue tarp is spread on the ground in front of the building. The background shows trees and a clear sky.

# C

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## Summary Results Tables

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**Table 1**  
Soil Analytical Results Summary

EQL	BTEX								Asbestos							
	Naphthalene (BTEX)		Benzene	Toluene	Ethylbenzene	Xylylene (m & p)	Xylylene (o)	Total Xylylene	Total BTEX	Asbestos (<2mm AF/FA)	Asbestos (ACM >7mm) Estimation	Total Asbestos	Asbestos fibres			
	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	g	%w/w	%w/w	g/kg	Detect			
PFAS NEMP 2020 Residential with minimal opportunities for soil access (HIL B)	1	0.2	0.5	0.5	0.5	0.5	0.5	0.5	0.2	-	-	-	-			
CCME 2002 Canadian Soil Quality Guidelines, Table 5 SQGH - Residential/Parkland, provisional guidelines																
NEPM 2013 Table 1A(1) HILs Res B Soil																
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour intrusion, Sand																
>0m, <1m	3	0.5	160	55			40									
>1m, <2m	NL	0.5	220	NL			60									
>2m, <4m	NL	0.5	310	NL			95									
>=4m	NL	0.5	540	NL			170									
NEPM 2013 Table 1B(5) Site-Specific EIL - Urban Res & Public Open Space																
NEPM 2013 Table 1B(6) ESLs for Urban Res, Coarse Soil		50	85	70			105									
NEPM 2013 Table 1B(7) Management Limits in Res / Parkland, Coarse Soil																
<b>Field ID</b>	<b>Depth</b>	<b>Date</b>	<b>Lab Report</b>													
BH102	0.5	21 May 2024	352254 / PFE1917	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH102	2	21 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH102 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-	-
BH103	1	22 May 2024	352254 / PFE1917	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH103	3	22 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH103	4	22 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH105	0.5	21 May 2024	352254 / PFE1917	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH105	3	21 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH106	0.5	22 May 2024	352254 / PFE1917	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH106	2	22 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH107	0.5	21 May 2024	352254 / PFE1917	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH107	3	21 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH107 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-	-
BH108	0.3	23 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH109	0.3	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH109	0.5	20 May 2024	PFE1917	-	-	-	-	-	-	-	-	-	-	-	-	-
BH109	1	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH110	0.25	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH110	1	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH111	0.5	23 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH111	2	23 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH112	0.3	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH112	1	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH113	0.5	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH113	2	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH114	0.5	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	ND	<0.001	<0.01	<0.1	ND
BH114	1	20 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
BH115	0.2	23 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	ND
BH115	1	23 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
DUP02 (BH102)	2	21 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
DUP03 (BH107)	3	21 May 2024	352254	<1	<0.2	<0.5	<1	<2	<1	<1	-	-	-	-	-	-
TRIP02 (BH102)	2	21 May 2024	ES2417505	<1	<0.2	<0.5	<0.5	<0.5	<0.5	<0.5	<0.2	-	-	-	-	-
TRIP03 (BH107)	3	21 May 2024	ES2417505	<1	<0.2	<0.5	<0.5	<0.5	<0.5	<0.2	-	-	-	-	-	-

ND = Non-detect at 0.1g/kg limit of reporting

<sup>^</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) HIL criteria

**Table 1**  
Soil Analytical Results Summary

ND = Non-detect at 0.1g/kg limit of reporting

<sup>a</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) HIL criteria.

Table 1  
Soil Analytical Results Summary

Naphthalene (BTEX)	Inorganics												Metals																						
	Exchangeable Calcium			pH 1:5 soil:water			Exchangeable Magnesium			Exchangeable Potassium			Exchangeable Sodium			Cation Exchange Capacity		Arsenic		Cadmium		Chromium (III+VI)		Copper		Iron		Lead		Mercury		Nickel		Zinc	
	mg/kg	meq/100g	-	meq/100g	meq/100g	meq/100g	meq/100g	meq/100g	meq/100g	meq/100g	meq/100g	meq/100g	meq/100g	meq/100g	-	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg							
EQL	1	0.1	-	0.1	0.1	0.1	1	-	4	0.4	1	1	10	1	0.1	1	1	-	32	<0.1	5	54	-	-	-	-	-	-	-						
PFAS NEMP 2020 Residential with minimal opportunities for soil access (HIL B)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-							
CCME 2002 Canadian Soil Quality Guidelines, Table 5 SQGHH - Residential/Parkland, provisional guidelines	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-							
NEPM 2013 Table 1A(1) HILs Res B Soil	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour intrusion, Sand	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
>0m, <1m	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
>1m, <2m	NL	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
>2m, <4m	NL	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
>=4m	NL	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
NEPM 2013 Table 1B(5) Site-Specific EIL - Urban Res & Public Open Space	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
NEPM 2013 Table 1B(6) ESLs for Urban Res, Coarse Soil	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
NEPM 2013 Table 1B(7) Management Limits in Res / Parkland, Coarse Soil	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
Field ID	Depth	Date	Lab Report																																
BH102	0.5	21 May 2024	352254 / PFE1917	<1	-	-	-	-	-	-	<4	<0.4	8	18	-	32	<0.1	5	54	-	-	-	-	-	-	-	-	-	-	-					
BH102	2	21 May 2024	352254	<1	0.8	4.4	1.3	0.2	0.8	3.1	6	<0.4	12	5	27,000	13	<0.1	<1	3	-	-	-	-	-	-	-	-	-	-	-					
BH102 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	5	<0.4	9	19	-	44	<0.1	6	51	-	-	-	-	-	-	-	-	-	-	-					
BH103	1	22 May 2024	352254 / PFE1917	<1	-	-	-	-	-	-	6	<0.4	6	21	-	19	<0.1	2	23	-	-	-	-	-	-	-	-	-	-	-					
BH103	3	22 May 2024	352254	<1	-	-	-	-	-	-	6	<0.4	16	8	-	17	<0.1	2	7	-	-	-	-	-	-	-	-	-	-	-					
BH103	4	22 May 2024	352254	<1	-	-	-	-	-	-	21	<0.4	6	3	-	6	<0.1	3	9	-	-	-	-	-	-	-	-	-	-	-					
BH105	0.5	21 May 2024	352254 / PFE1917	<1	-	-	-	-	-	-	5	<0.4	13	12	-	30	<0.1	5	39	-	-	-	-	-	-	-	-	-	-	-					
BH105	3	21 May 2024	352254	<1	-	-	-	-	-	-	9	<0.4	18	5	-	12	<0.1	1	11	-	-	-	-	-	-	-	-	-	-	-					
BH106	0.5	22 May 2024	352254 / PFE1917	<1	-	-	-	-	-	-	<4	<0.4	7	16	-	15	<0.1	6	30	-	-	-	-	-	-	-	-	-	-	-					
BH106	2	22 May 2024	352254	<1	1.1	4.9	1.2	0.2	0.3	2.9	<4	<0.4	9	3	16,000	10	<0.1	<1	2	-	-	-	-	-	-	-	-	-	-	-	-				
BH107	0.5	21 May 2024	352254 / PFE1917	<1	-	-	-	-	-	-	5	<0.4	9	17	-	19	<0.1	8	51	-	-	-	-	-	-	-	-	-	-	-	-				
BH107	3	21 May 2024	352254	<1	-	-	-	-	-	-	<4	<0.4	7	11	-	24	<0.1	3	45	-	-	-	-	-	-	-	-	-	-	-	-				
BH107 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	5	<0.4	8	19	-	18	<0.1	6	33	-	-	-	-	-	-	-	-	-	-	-	-				
BH108	0.3	23 May 2024	352254	<1	-	-	-	-	-	-	5	<0.4	8	41	35,000	19	<0.1	22	75	-	-	-	-	-	-	-	-	-	-	-	-	-			
BH109	0.3	20 May 2024	352254	<1	-	-	-	-	-	-	<4	<0.4	9	40	-	18	<0.1	16	59	-	-	-	-	-	-	-	-	-	-	-	-	-			
BH109	0.5	20 May 2024	PFE1917	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
BH109	1	20 May 2024	352254	<1	-	-	-	-	-	-	<4	<0.4	10	1	-	10	<0.1	2	13	-	-	-	-	-	-	-	-	-	-	-	-	-			
BH110	0.25	20 May 2024	352254	<1																															

Table 1  
Soil Analytical Results Summary

	Naphthalene (BTEX)	1,2,3,4,6,7,8-Heptachloroanthrene	1,2,3,4,6,7,8-Heptachlorodibenzofuran	1,2,3,4,6,7,8-Heptachlorodibenzofuran	1,2,3,4,7,8,9-Hexachloroxanthrene	1,2,3,4,7,8,9-Hexachloroxanthrene	1,2,3,6,7,8-Hexachlorobenzofuran	1,2,3,6,7,8-Hexachlorobenzofuran	1,2,3,7,8,9-Hexachlorobenzofuran	Dioxins and Furans	1,2,3,7,8,9-Hexachlorobenzofuran	1,2,3,7,8,9-Hexachlorobenzofuran	1,2,3,7,8-Tetrachloroxanthene	1,2,3,7,8-Tetrachloroxanthene	1,2,3,7,8-Pentachlorodibenzofuran	1,2,3,7,8-Pentachlorodibenzofuran	1,2,3,7,8-Pentachlorodibenzofuran	1,2,3,4,6,7,8,9-Octachloroanthene	1,2,3,4,6,7,8,9-Octachlorobenzofuran
mg/kg	1	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g		pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	
EQL	1																		
PFAS NEMP 2020 Residential with minimal opportunities for soil access (HIL B)																			
CCME 2002 Canadian Soil Quality Guidelines, Table 5 SQGHH - Residential/Parkland, provisional guidelines																			
NEPM 2013 Table 1A(1) HILs Res B Soil																			
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour intrusion, Sand																			
>=0m, <1m	3																		
>=1m, <2m	NL																		
>=2m, <4m	NL																		
>=4m	NL																		
NEPM 2013 Table 1B(5) Site-Specific EIL - Urban Res & Public Open Space																			
NEPM 2013 Table 1B(6) ESLs for Urban Res, Coarse Soil																			
NEPM 2013 Table 1B(7) Management Limits in Res / Parkland, Coarse Soil																			
Field ID	Depth	Date	Lab Report																
BH102	0.5	21 May 2024	352254 / PFE1917	<1	120	57	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	1,400	18
BH102	2	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH102 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH103	1	22 May 2024	352254 / PFE1917	<1	7.3	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<0.50	<2.0	<2.0	<2.0	<2.0	760	<5.0
BH103	3	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH103	4	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH105	0.5	21 May 2024	352254 / PFE1917	<1	21	3.9	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<0.50	<2.0	<2.0	<2.0	<2.0	2,300	<5.0
BH105	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH106	0.5	22 May 2024	352254 / PFE1917	<1	13	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<0.50	<2.0	<2.0	<2.0	<2.0	580	<5.0
BH106	2	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH107	0.5	21 May 2024	352254 / PFE1917	<1	12	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<0.50	<2.0	<2.0	<2.0	<2.0	1,200	<5.0
BH107	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH107 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH108	0.3	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH109	0.3	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH109	0.5	20 May 2024	PFE1917	-	6.1	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<2.0	<0.50	<2.0	<2.0	<2.0	<2.0	800	<5.0
BH109	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH110	0.25	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH110	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH111	0.5	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH111	2	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH112	0.3	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH112	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH113	0.5	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH113	2	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH114	0.5	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH114	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH115	0.2	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
BH115	1	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
DUP02 (BH102)	2	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
DUP03 (BH107)	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TRIP02 (BH102)	2	21 May 2024	ES2417505	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TRIP03 (BH107)	3	21 May 2024	ES2417505	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

ND = Non-detect at 0.1g/kg limit of reporting

<sup>a</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) HIL criteria

Naphthalene (B-TEX)	Dioxins I-TEQ 1xLOR														
	1234678-HxCDD I-TEQ3 (LOR)	1234678-HxCDF I-TEQ3 (LOR)	1234789-HxCDF I-TEQ3 (LOR)	1234789-HxCDD I-TEQ3 (LOR)	123478-HxCDF I-TEQ3 (LOR)	123678-HxCDD I-TEQ3 (LOR)	123678-HxCDF I-TEQ3 (LOR)	123789-HxCDD I-TEQ3 (LOR)	123789-HxCDF I-TEQ3 (LOR)	12378-PeCDD I-TEQ3 (LOR)	1234678-HxCDF I-TEQ3 (LOR)	2378-TCDI I-TEQ3 (LOR)	OCDD I-TEQ3 (LOR)	OCDF I-TEQ3 (LOR)	
mg/kg	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g	pg/g		
EQL	1														
PFAS NEMP 2020 Residential with minimal opportunities for soil access (HIL B)															
CCME 2002 Canadian Soil Quality Guidelines, Table 5 SQGHH - Residential/Parkland, provisional guidelines													4		
NEPM 2013 Table 1A(1) HILs Res B Soil															
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour intrusion, Sand															
>=0m, <1m	3														
>=1m, <2m	NL														
>=2m, <4m	NL														
>=4m	NL														
NEPM 2013 Table 1B(5) Site-Specific EIL - Urban Res & Public Open Space															
NEPM 2013 Table 1B(6) ESLs for Urban Res, Coarse Soil															
NEPM 2013 Table 1B(7) Management Limits in Res / Parkland, Coarse Soil															
<b>Field ID</b>	<b>Depth</b>	<b>Date</b>	<b>Lab Report</b>												
BH102	0.5	21 May 2024	352254 / PFE1917	<1	1.2	0.56	-	-	-	-	-	-	1.4	0.018	3.178
BH102	2	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH102 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-
BH103	1	22 May 2024	352254 / PFE1917	<1	0.073	-	-	-	-	-	-	-	0.76	-	0.833
BH103	3	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH103	4	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH105	0.5	21 May 2024	352254 / PFE1917	<1	0.21	0.039	-	-	-	-	-	-	2.3	-	2.549
BH105	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH106	0.5	22 May 2024	352254 / PFE1917	<1	0.13	-	-	-	-	-	-	-	0.58	-	0.71
BH106	2	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH107	0.5	21 May 2024	352254 / PFE1917	<1	0.12	-	-	-	-	-	-	-	1.2	-	1.32
BH107	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH107 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-
BH108	0.3	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH109	0.3	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH109	0.5	20 May 2024	PFE1917	-	0.061	-	-	-	-	-	-	-	0.8	-	0.861
BH109	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH110	0.25	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH110	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH111	0.5	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH111	2	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH112	0.3	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH112	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH113	0.5	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH113	2	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH114	0.5	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH114	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH115	0.2	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
BH115	1	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
DUP02 (BH102)	2	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
DUP03 (BH107)	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-
TRIP02 (BH102)	2	21 May 2024	ES2417505	<1	-	-	-	-	-	-	-	-	-	-	-
TRIP03 (BH107)	3	21 May 2024	ES2417505	<1	-	-	-	-	-	-	-	-	-	-	-

ND = Non-detect at 0.1g/kg limit of reporting

<sup>a</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) HIL criteria

Table 1  
Soil Analytical Results Summary

		Organochlorine Pesticides																						
		Naphthalene (BTEX)	4,4'-DDE	a-BHC	Aldrin	Dieldrin	b-BHC	Chlordane (cis)	Chlordane (trans)	d-BHC	DDD	DDT	DDT+DDE-DDO	Endosulfan I	Endosulfan II	Endosulfan sulphate	Endrin	Endrin aldehyde	g-BHC (Lindane)	Hepachlor	Hepachlor epoxide	Hexachlorobenzene	Methoxychlor	Mirex
mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	
EQL		1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	
PFAS NEMP 2020 Residential with minimal opportunities for soil access (HIL B)																								
CCME 2002 Canadian Soil Quality Guidelines, Table 5 SQGHH - Residential/Parkland, provisional guidelines																								
NEPM 2013 Table 1A(1) HILs Res B Soil																								
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour intrusion, Sand																								
>=0m, <1m																								
>=1m, <2m																								
>=2m, <4m																								
>=4m																								
NEPM 2013 Table 1B(5) Site-Specific EIL - Urban Res & Public Open Space																								
NEPM 2013 Table 1B(6) ESLs for Urban Res, Coarse Soil																								
NEPM 2013 Table 1B(7) Management Limits in Res / Parkland, Coarse Soil																								
Field ID	Depth	Date	Lab Report																					
BH102	0.5	21 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH102	2	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH102 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH103	1	22 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH103	3	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH103	4	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH105	0.5	21 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH105	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH106	0.5	22 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH106	2	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH107	0.5	21 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH107	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH107 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH108	0.3	23 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH109	0.3	20 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH109	0.5	20 May 2024	PFE1917	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH109	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH110	0.25	20 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH110	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH111	0.5	23 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH111	2	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH112	0.3	20 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH112	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH113	0.5	20 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	
BH113	2	20 May 2024	352254	<1	-</td																			

ND = Non-detect at 0.1g/kg limit of reporting

<sup>^</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) HIL criteria.

Table 1  
Soil Analytical Results Summary

Naphthalene (BTEX)	PCBs										Particle Size	Organic											
	Arochlor 1016	Arochlor 1221	Arochlor 1232	Arochlor 1242	Arochlor 1248	Arochlor 1254	Arochlor 1260	PCBs (Sum of total)	% Clay in soils <2um	TOC %			Perfluorobutane sulfonic acid (PFBS)	Perfluoropentane sulfonic acid (PFPS)	Perfluorohexane sulfonic acid (PFHxS)	Perfluoroheptane sulfonic acid (PFHpS)	Perfluoroctane sulfonic acid (PFOS)	Perfluorobutanoic acid (PFBA)	Perfluorohexanoic acid (PFHA)	Perfluoropentanoic acid (PFPeA)			
	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg			mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	
EQL	1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	100	0.0001	0.0001	0.0001	2	2								0.0002	
PFAS NEMP 2020 Residential with minimal opportunities for soil access (HIL B)																							
CCME 2002 Canadian Soil Quality Guidelines, Table 5 SGHH - Residential/Parkland, provisional guidelines																							
NEPM 2013 Table 1A(1) HILs Res B Soil									1														
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour intrusion, Sand																							
>0m, <1m	3																						
>=1m, <2m	NL																						
>=2m, <4m	NL																						
>=4m	NL																						
NEPM 2013 Table 1B(5) Site-Specific EIL - Urban Res & Public Open Space																							
NEPM 2013 Table 1B(6) ESLs for Urban Res, Coarse Soil																							
NEPM 2013 Table 1B(7) Management Limits in Res / Parkland, Coarse Soil																							
Field ID	Depth	Date	Lab Report	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH102	0.5	21 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH102	2	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	20	3,100	-	-	-	-	-	-	-	-	-
BH102 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH103	1	22 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	0.0002	<0.0002	<0.0002	<0.0001	<0.0002
BH103	3	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH103	4	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH105	0.5	21 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH105	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH106	0.5	22 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH106	2	22 May 2024	352254	<1	-	-	-	-	-	-	-	-	30	2,600	-	-	-	-	-	-	-	-	-
BH107	0.5	21 May 2024	352254 / PFE1917	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH107	3	21 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH107 - [TRIPPLICATE]	0.5	21 May 2024	352254	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH108	0.3	23 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH109	0.3	20 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH109	0.5	20 May 2024	PFE1917	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH109	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH110	0.25	20 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH110	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH111	0.5	23 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH111	2	23 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH112	0.3	20 May 2024	352254	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	-	-	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0002	<0.0002	<0.0001	<0.0002
BH112	1	20 May 2024	352254	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
BH113	0.5	20 May 2024	352254	<1</td																			

**Table 1**  
Soil Analytical Results Summary

ND = Non-detect at 0.1g/kg limit of reporting

<sup>a</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) HIL criteria.

**Table 2**  
 Soil Analytical Results Summary  
 Acid Sulfate Soils

	Acid Sulphate Soils - Acid Base Accounting				Acid Sulphate Soils - Acidity Trail		Acid Sulphate Soils - ANC		Acid Sulphate Soils - Liming Rate		Acid Sulphate Soils - pH	Acid Sulphate Soils - Potential Acidity		Acid Sulphate Soils - Retained Acidity		Acid Sulphate Soils - Sulfur Trail
	a-Net Acidity without ANC	s-Net Acidity without ANC	Net Acidity (acidity units)	Net Acidity (sulfur units)	Titratable Actual Acidity (sulfur units)	Titratable Actual Acidity	Acid Neutralising Capacity	Acid Neutralising Capacity (sulfur units)	Liming Rate	Liming Rate excluding ANC	pH <sub>KCl</sub>	Chromium Reducible Sulphur (acidity units)	Chromium Reducible Sulphur	HCl Extractable Sulfur	Net Acid Soluble Sulfur	KCl Extractable Sulfur
	moles H+/t	% S	mole H+/t	%S	%S	mole H+/t	%CaCO <sub>3</sub>	%S	kg CaCO <sub>3</sub> /t	kg CaCO <sub>3</sub> /t	-	mole H+/t	%w/w	%S	%S	%
EQL	5	0.005	5	0.005	0.01	5	0.05	0.05	0.75	0.75		3	0.005	0.005	0.005	0.005
National Acid Sulfate Soils Guidance (June 2018)	18	0.03										0.01				
ASSMAC (1998) Assessment Guidelines Table 4.4; >1,000 tonnes disturbed, coarse texture	18	0.03														

Field ID	Depth	Date	Lab Report	870	1.4	500	0.80	<0.01	<5	2.8	0.88	38	65	9.1	870	1.4	-	-	-
BH102	4	21 May 2024	352254	870	1.4	500	0.80	<0.01	<5	2.8	0.88	38	65	9.1	870	1.4	-	-	-
BH102	6	21 May 2024	352254	230	0.37	150	0.24	<0.01	<5	0.60	0.19	11	17	7.8	230	0.37	-	-	-
BH103	4	22 May 2024	352254	420	0.68	<5	<0.005	<0.01	<5	3.5	1.1	<0.75	32	8.9	420	0.68	-	-	-
BH103	6	22 May 2024	352254	30	0.047	30	0.050	0.03	21	-	-	2	2.2	4.0	<3	<0.005	0.044	0.012	0.038
BH105	4	21 May 2024	352254	340	0.55	340	0.55	<0.01	<5	-	-	26	26	5.6	340	0.54	-	-	-
BH106	3	22 May 2024	352254	87	0.14	87	0.14	0.12	74	-	-	7	6.6	3.8	10	0.02	0.007	0.009	<0.005
BH106	6	22 May 2024	352254	27	0.044	27	0.044	<0.01	<5	-	-	2	2.0	5.4	26	0.04	-	-	-
BH107	4	21 May 2024	352254	240	0.38	240	0.38	<0.01	<5	-	-	18	18	6.3	240	0.38	-	-	-
BH109	2	20 May 2024	352254	<5	0.0060	<5	0.0060	<0.01	<5	-	-	<0.75	<0.75	5.1	<3	<0.005	-	-	-
BH114	1	20 May 2024	352254	110	0.18	110	0.18	0.16	99	-	-	8.5	8.5	3.5	10	0.02	0.018	0.010	0.013
BH115	5.1	23 May 2024	352254	22	0.035	22	0.040	0.02	11	-	-	2	1.7	4.4	<3	<0.005	0.037	0.019	0.028
BH115	10.7	23 May 2024	352254	130	0.21	130	0.21	<0.01	<5	-	-	9.8	9.8	5.6	130	0.21	-	-	-

#### Statistics

Number of Results	12	12	12	12	12	12	3	3	12	12	12	12	12	4	4	4
Number of Detects	11	12	10	11	4	4	3	3	10	11	12	9	9	4	4	3
Minimum Concentration	<5	0.006	<5	<0.005	<0.01	<5	0.6	0.19	<0.75	<0.75	3.5	<3	<0.005	0.007	0.009	<0.005
Minimum Detect	22	0.006	22	0.006	0.02	11	0.6	0.19	2	1.7	3.5	10	0.02	0.007	0.009	0.013
Maximum Concentration	870	1.4	500	0.8	0.16	99	3.5	1.1	38	65	9.1	870	1.4	0.044	0.019	0.038
Maximum Detect	870	1.4	500	0.8	0.16	99	3.5	1.1	38	65	9.1	870	1.4	0.044	0.019	0.038
Average Concentration *	209	0.34	137	0.22	0.031	19	2.3	0.72	10	16	5.8	190	0.31	0.027	0.012	0.02
Median Concentration *	120	0.195	98.5	0.16	0.005	2.5	2.8	0.88	7.75	9.15	5.5	78	0.125	0.0275	0.011	0.0205
Standard Deviation *	247	0.4	154	0.25	0.052	33	1.5	0.47	12	19	1.9	260	0.42	0.017	0.0045	0.016
95% UCL (Student's-t) *	337.3	0.543	216.5	0.348	0.0579	35.64	4.851	1.524	16.46	25.37	6.78	324.9	0.522	0.0465	0.0178	0.0389
% of Detects	92	100	83	92	33	33	100	100	83	92	100	75	75	100	100	75
% of Non-Detects	8	0	17	8	67	67	0	0	17	8	0	25	25	0	0	25

\* A Non Detect Multiplier of 0.5 has been applied.

**Table 3**  
 Groundwater Analytical Results Summary

	BTEX								TRH					
	Naphthalene (BTEX) µg/L	Benzene µg/L	Toluene µg/L	Ethylbenzene µg/L	Xylene (m & p) µg/L	Xylene (o) µg/L	Xylene Total µg/L	Total BTEX µg/L	C6-C10 Fraction (F1) µg/L	>C10-C16 Fraction (F1 minus BTEX) µg/L	>C10-C16 Fraction (F2 minus Naphthalene) µg/L	>C16-C34 Fraction (F3) µg/L	>C34-C40 Fraction (F4) µg/L	>C10-C40 Fraction (sum) µg/L
EQL	1	1	1	1	2	1	2	1	10	10	50	50	100	100
ANZG Marine Water Toxicant DGVs LOSP 95% (July 2023)	70	700	180	80										
ANZG 2023 Dioxins in Freshwater Technical Brief 95% Species Protection														
NEPM 2013 Table 1A(4) Res HSL A & B GW for Vapour Intrusion, Sand														
>=2m, <4m	NL	800	NL	NL			NL		1,000		1,000			
PFAS NEMP 2020 Recreational Water														
PFAS NEMP 2020 Interim Marine 95%														
PFAS NEMP 2020 Interim Marine 99%														

Field ID	Date	Lab Report Number	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
EW1	31 May 2024	352854-A	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
MW1-B	31 May 2024	352854-A	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
MW2-B	31 May 2024	352854-A	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
MW3-B	30 May 2024	352732-A / PFF0035	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
MW4-B	31 May 2024	352854-A	<1	<1	<1	<1	<2	<1	-	-	50	50	<50	<50	<100	<100	<50
RMW01	30 May 2024	352732-A / PFF0035	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
RMW02	30 May 2024	352732-A	<1	<1	46	<1	<2	<1	-	-	93	47	<50	<50	<100	<100	<50
RMW03	30 May 2024	352732-A	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
RMW04	30 May 2024	352732-A / PFF0035	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
RMW05	31 May 2024	352854-A	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
DUP1 (MW1-B)	31 May 2024	352854-A	<1	<1	<1	<1	<2	<1	-	-	<10	<10	<50	<50	<100	<100	<50
DUP2 (EW1)	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
TRIP1 (MW1-B)	31 May 2024	ES2418204	<5	<1	<2	<2	<2	<2	<2	<1	<20	<20	<100	<100	<100	<100	<100
TRIP2 (EW1)	31 May 2024	ES2418204	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

<sup>^</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) ANZG criteria

**Table 3**  
 Groundwater Analytical Results Summary

	PAH																PAHs (Sum of total)		PAHs (Sum of positives)	
	Benzo(b+j+k)fluoranthene	Acenaphthene	Acenaphthylene	Anthracene	Benz(a)anthracene	Benzo(a) pyrene	Benzo(b+j)fluoranthene	Benzo(g,h,i)perylene	Benzo(k)fluoranthene	Chrysene	Dibenz(a,h)anthracen	Fluoranthene	Fluorene	Indeno[1,2,3-c,d]pyrene	Naphthalene	Phenanthrene	Pyrene	Benzo(a)pyrene TEQ	PAHs (Sum of total)	PAHs (Sum of positives)
	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L
EQL	0.2	0.1	0.1	0.1	0.1	0.1	1	0.1	1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.5	0.5	0.1
ANZG Marine Water Toxicant DGVs LOSP 95% (July 2023)				0.4		0.2						1.4			70	2				
ANZG 2023 Dioxins in Freshwater Technical Brief 95% Species Protection																				
NEPM 2013 Table 1A(4) Res HSL A & B GW for Vapour Intrusion, Sand															NL   NL   NL					
>=2m, <4m															NL					
PFAS NEMP 2020 Recreational Water																				
PFAS NEMP 2020 Interim Marine 95%																				
PFAS NEMP 2020 Interim Marine 99%																				

Field ID	Date	Lab Report Number																				
EW1	31 May 2024	352854-A	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
MW1-B	31 May 2024	352854-A	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
MW2-B	31 May 2024	352854-A	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
MW3-B	30 May 2024	352732-A / PFF0035	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
MW4-B	31 May 2024	352854-A	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
RMW01	30 May 2024	352732-A / PFF0035	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
RMW02	30 May 2024	352732-A	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	0.22
RMW03	30 May 2024	352732-A	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
RMW04	30 May 2024	352732-A / PFF0035	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
RMW05	31 May 2024	352854-A	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DUP1 (MW1-B)	31 May 2024	352854-A	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DUP2 (EW1)	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TRIP1 (MW1-B)	31 May 2024	ES2418204	-	<1.0	<1.0	<1.0	<1.0	<1.0	<0.5	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<0.5
TRIP2 (EW1)	31 May 2024	ES2418204	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

<sup>a</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) ANZG criteria

**Table 3**  
 Groundwater Analytical Results Summary

	Inorganics																		
	Hardness (filtered) µg/L	Alkalinity (Bicarbonate as CaCO <sub>3</sub> ) µg/L	Alkalinity (Carbonate as CaCO <sub>3</sub> ) µg/L	Alkalinity (Hydroxide) as CaCO <sub>3</sub> µg/L	Alkalinity (total) as CaCO <sub>3</sub> µg/L	Ammonia as N (filtered) µg/L	Chloride µg/L	Ionic Balance %	Kjeldahl Nitrogen Total µg/L	Nitrate (as N) (filtered) µg/L	Nitrite (as N) (filtered) µg/L	Organic Nitrogen as N µg/L	Nitrogen (Total Oxidised) µg/L	Nitrogen (Total) µg/L	Total Phosphorus (Organic Phosphate) µg/L	Reactive Phosphorus as P (Orthophosphate as P) (filtered) µg/L	Sodium (filtered) µg/L	Sulphate µg/L	Total Dissolved Solids (lab) µg/L
EQL	3,000	5,000	5,000	5,000	5,000	5	1,000		100	5	5	200	5	100	50	5	500	1,000	5,000
ANZG Marine Water Toxicant DGVs LOSP 95% (July 2023)						910													
ANZG 2023 Dioxins in Freshwater Technical Brief 95% Species Protection																			
NEPM 2013 Table 1A(4) Res HSL A & B GW for Vapour Intrusion, Sand																			
>=2m, <4m																			
PFAS NEMP 2020 Recreational Water																			
PFAS NEMP 2020 Interim Marine 95%																			
PFAS NEMP 2020 Interim Marine 99%																			

Field ID	Date	Lab Report Number	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
EW1	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
MW1-B	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
MW2-B	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
MW3-B	30 May 2024	352732-A / PFF0035	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
MW4-B	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
RMW01	30 May 2024	352732-A / PFF0035	2,500,000	<5,000	<5,000	<5,000	<5,000	210	6,500,000	-3.0	1,100	<5	<5	900	<5	1,100	<50	<5	3,200,000
RMW02	30 May 2024	352732-A	1,500,000	<5,000	<5,000	<5,000	<5,000	130	6,300,000	0	300	10	<5	<200	10	300	<50	<5	4,000,000
RMW03	30 May 2024	352732-A	2,000,000	<5,000	<5,000	<5,000	<5,000	150	8,000,000	0	300	20	7	<200	30	400	<50	<5	5,000,000
RMW04	30 May 2024	352732-A / PFF0035	750,000	140,000	<5,000	<5,000	140,000	260	5,000,000	-6.0	300	<5	<5	<200	8	300	<50	<5	3,100,000
RMW05	31 May 2024	352854-A	520,000	75,000	<5,000	<5,000	75,000	16	1,500,000	-3.0	200	310	21	<200	300	500	<50	<5	850,000
DUP1 (MW1-B)	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
DUP2 (EW1)	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TRIP1 (MW1-B)	31 May 2024	ES2418204	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TRIP2 (EW1)	31 May 2024	ES2418204	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

<sup>^</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) ANZG criteria

**Table 3**  
 Groundwater Analytical Results Summary

	Metals																			
	Aluminium (filtered) µg/L	Antimony (filtered) µg/L	Arsenic (filtered) µg/L	Barium (filtered) µg/L	Beryllium (filtered) µg/L	Boron (filtered) µg/L	Cadmium (filtered) µg/L	Calcium (filtered) µg/L	Chromium (III+VI) (filtered) µg/L	Cobalt (filtered) µg/L	Copper (filtered) µg/L	Iron (filtered) µg/L	Lead (filtered) µg/L	Lithium (filtered) µg/L	Magnesium (filtered) µg/L	Manganese (filtered) µg/L	Mercury (filtered) µg/L	Molybdenum (filtered) µg/L	Nickel (filtered) µg/L	Potassium (filtered) µg/L
EQL	10	1	1	1	0.5	20	0.1	500	1	1	1	10	1	1	500	5	0.05	1	1	500
ANZG Marine Water Toxicant DGVs LOSP 95% (July 2023)							5.5		4.4^	1	1.3		4.4				0.4		70	
ANZG 2023 Dioxins in Freshwater Technical Brief 95% Species Protection																				
NEPM 2013 Table 1A(4) Res HSL A & B GW for Vapour Intrusion, Sand																				
>=2m, <4m																				
PFAS NEMP 2020 Recreational Water																				
PFAS NEMP 2020 Interim Marine 95%																				
PFAS NEMP 2020 Interim Marine 99%																				

Field ID	Date	Lab Report Number	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
EW1	31 May 2024	352854-A	-	-	<1	-	-	-	<0.1	-	<1	-	<1	-	<1	-	<0.05	-	120	-		
MW1-B	31 May 2024	352854-A	-	-	2	-	-	-	<0.1	-	<1	-	<1	-	<1	-	<0.05	-	2	-		
MW2-B	31 May 2024	352854-A	-	-	2	-	-	-	<0.1	-	<1	-	<1	-	<1	-	<0.05	-	25	-		
MW3-B	30 May 2024	352732-A / PFF0035	-	-	<1	-	-	-	0.1	-	<1	-	1	-	<1	-	<0.05	-	<1	-		
MW4-B	31 May 2024	352854-A	-	-	<1	-	-	-	<0.1	-	<1	-	2	-	<1	-	<0.05	-	<1	-		
RMW01	30 May 2024	352732-A / PFF0035	9,200	<1	4	60	1	1,000	0.5	290,000	3	25	<1	120,000	3	4	430,000	480	<0.05	<1	30	96,000
RMW02	30 May 2024	352732-A	5,600	<1	4	25	5	1,600	0.9	91,000	1	73	12	66,000	44	21	310,000	2,500	<0.05	<1	61	120,000
RMW03	30 May 2024	352732-A	16,000	<1	9	96	6	1,400	1.1	130,000	3	92	120	50,000	19	24	400,000	2,600	<0.05	<1	130	120,000
RMW04	30 May 2024	352732-A / PFF0035	100	<1	3	46	<0.5	2,000	<0.1	71,000	<1	51	<1	31,000	<1	2	140,000	1,900	<0.05	<1	9	96,000
RMW05	31 May 2024	352854-A	60	<1	<1	50	3	60	<0.1	10,000	<1	91	<1	48,000	<1	36	120,000	4,300	<0.05	<1	94	7,800
DUP1 (MW1-B)	31 May 2024	352854-A	-	-	2	-	-	-	<0.1	-	<1	-	<1	-	<1	-	-	<0.05	-	2	-	
DUP2 (EW1)	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-		
TRIP1 (MW1-B)	31 May 2024	ES2418204	-	-	<1	-	-	-	<0.1	-	<1	-	<1	-	<1	-	-	<0.1	-	2	-	
TRIP2 (EW1)	31 May 2024	ES2418204	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			

<sup>^</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) ANZG criteria

**Table 3**  
 Groundwater Analytical Results Summary

	Selenium & Trace Elements														Dioxins & Furans																				
	Selenium (filtered)		Silicon		Silver (filtered)		Strontium (filtered)		Uranium		Vanadium (filtered)		Zinc (filtered)		2,3,7,8-TcDD		1,2,3,7,8-PeCDD		1,2,3,4,7,8-HxCDD		1,2,3,7,8,9-HxCDD		1,2,3,4,6,7,8-HpCDD		OCDD		2,3,7,8-TCDF		1,2,3,7,8-PeCDF		2,3,4,7,8-PeCDF		1,2,3,6,7,8-HxCDF		1,2,3,7,8-HxCDF
	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L	pg/L			
EQL	1	200	1	1	0.5	1	1	1	5	20	20	20	20	20	50	5	20	20	20	20	20	20	20	20	20	20	20	20	20	20	20				
ANZG Marine Water Toxicant DGVs LOSP 95% (July 2023)					1.4				100	8																									
ANZG 2023 Dioxins in Freshwater Technical Brief 95% Species Protection																																			
NEPM 2013 Table 1A(4) Res HSL A & B GW for Vapour Intrusion, Sand																																			
>=2m, <4m																																			
PFAS NEMP 2020 Recreational Water																																			
PFAS NEMP 2020 Interim Marine 95%																																			
PFAS NEMP 2020 Interim Marine 99%																																			
Field ID	Date	Lab Report Number	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
EW1	31 May 2024	352854-A	-	-	-	-	-	-	-	68	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
MW1-B	31 May 2024	352854-A	-	-	-	-	-	-	-	9	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
MW2-B	31 May 2024	352854-A	-	-	-	-	-	-	-	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
MW3-B	30 May 2024	352732-A / PFF0035	-	-	-	-	-	-	-	32	<5	<20	<20	<20	<20	<20	<20	<20	57	<5	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20				
MW4-B	31 May 2024	352854-A	-	-	-	-	-	-	-	7	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
RMW01	30 May 2024	352732-A / PFF0035	<1	15,000	<1	5,200	<0.5	5	490	<5	<20	<20	<20	<20	<20	<20	<20	<20	120	<5	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20				
RMW02	30 May 2024	352732-A	<1	12,000	<1	1,800	2.0	<1	420	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
RMW03	30 May 2024	352732-A	<1	12,000	<1	2,700	1.3	4	620	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
RMW04	30 May 2024	352732-A / PFF0035	<1	3,700	<1	1,000	<0.5	<1	32	<5	<20	<20	<20	<20	<20	<20	<20	<50	<5	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20	<20					
RMW05	31 May 2024	352854-A	<1	11,000	<1	88	<0.5	<1	400	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
DUP1 (MW1-B)	31 May 2024	352854-A	-	-	-	-	-	-	-	6	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-			
DUP2 (EW1)	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
TRIP1 (MW1-B)	31 May 2024	ES2418204	-	-	-	-	-	-	-	<5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				
TRIP2 (EW1)	31 May 2024	ES2418204	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-				

<sup>a</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) ANZG criteria

**Table 3**  
 Groundwater Analytical Results Summary

	2,3,4,6,7,8-HxCDF	1,2,3,4,6,7,8-HpCDF	1,2,3,4,7,8,9-HpCDF	OCDF	Dioxins and Furans TEQ (Detects)	PFAS																		
	pg/L	pg/L	pg/L	pg/L	pg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	μg/L	
EQL	20	20	20	50	-	0.02	0.0002	0.0002	0.1	0.02	0.02	0.02	0.0002	0.05	0.0004	0.0004	0.05	0.0002	0.0002	0.0002	0.0002	0.0002		
ANZG Marine Water Toxicant DGVs LOSP 95% (July 2023)																								
ANZG 2023 Dioxins in Freshwater Technical Brief 95% Species Protection					5																			
NEPM 2013 Table 1A(4) Res HSL A & B GW for Vapour Intrusion, Sand																								
>=2m, <4m																								
PFAS NEMP 2020 Recreational Water									2	2							10					2		
PFAS NEMP 2020 Interim Marine 95%										0.13							220							
PFAS NEMP 2020 Interim Marine 99%										0.00023							19							

Field ID	Date	Lab Report Number	-	-	-	-	-	-	<0.0002	<0.0002	-	-	-	-	<0.0002	-	<0.0004	<0.0004	-	<0.0002	<0.0002	<0.0002	
EW1	31 May 2024	352854-A	-	-	-	-	-	-	<0.0002	<0.0002	-	-	-	-	<0.0002	-	<0.0004	<0.0004	-	<0.0002	<0.0002	<0.0002	
MW1-B	31 May 2024	352854-A	-	-	-	-	-	-	0.0081	0.001	-	-	-	-	0.0023	-	0.0007	<0.0004	-	0.0094	0.012	0.0035	
MW2-B	31 May 2024	352854-A	-	-	-	-	-	-	0.001	0.002	-	-	-	-	0.0020	-	0.0007	<0.0004	-	0.0032	0.0060	0.0038	
MW3-B	30 May 2024	352732-A / PFF0035	<20	<20	<20	<50	0.017	-	0.002	0.010	-	-	-	-	0.002	-	<0.0004	<0.0004	-	0.012	0.014	0.012	
MW4-B	31 May 2024	352854-A	-	-	-	-	-	-	0.0039	0.013	-	-	-	-	0.018	-	<0.0004	<0.0004	-	0.017	0.035	0.031	
RMW01	30 May 2024	352732-A / PFF0035	<20	<20	<20	<50	0.036	-	0.068	0.0031	-	-	-	-	0.0029	-	<0.0004	<0.0004	-	0.071	0.074	0.0060	
RMW02	30 May 2024	352732-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
RMW03	30 May 2024	352732-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
RMW04	30 May 2024	352732-A / PFF0035	<20	<20	<20	<50	-	-	<0.0002	0.0003	-	-	-	-	0.0002	-	<0.0004	<0.0004	-	0.0003	0.0006	0.0006	
RMW05	31 May 2024	352854-A	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DUP1 (MW1-B)	31 May 2024	352854-A	-	-	-	-	-	-	-	0.0074	0.001	-	-	-	-	0.0021	-	0.0005	<0.0004	-	0.0085	0.011	0.0031
DUP2 (EW1)	31 May 2024	352854-A	-	-	-	-	-	-	<0.0002	0.0002	-	-	-	-	<0.0002	-	<0.0004	<0.0004	-	0.0002	0.0002	0.0002	
TRIP1 (MW1-B)	31 May 2024	ES2418204	-	-	-	-	-	-	0.03	<0.01	<0.01	<0.1	<0.02	0.04	<0.02	<0.01	<0.05	<0.05	<0.05	<0.01	-	-	
TRIP2 (EW1)	31 May 2024	ES2418204	-	-	-	-	-	-	<0.02	<0.01	<0.01	<0.1	<0.02	<0.02	<0.01	<0.05	<0.05	<0.05	<0.01	-	-	-	

<sup>a</sup> Chromium (III+VI) concentrations have been compared to Chromium (VI) ANZG criteria

	BTEX							TRH																									
	Naphthalene (BTEX)		Benzene	Toluene	Ethylbenzene	Xylene (m & p)	Xylene (o)	C6-C10 Fraction (F1)		C6-C10 (F1 minus BTEX)		>C10-C16 Fraction (F2)		>C10-C16 Fraction (F2 minus Naphthalene)		>C16-C34 Fraction (F3)		>C34-C40 Fraction (F4)		>C10-C40 Fraction (Sum)		Benz[b(j+k)]fluoranthene		Acenaphthene		Acenaphthylene		Anthracene		Benz(a)anthracene		Benz(a) pyrene	
	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg				
EQL	1	0.2	0.5	0.5	0.5	0.5	0.5	10	10	50	50	100	100	50	0.2	0.1	0.1	0.1	0.1	0.1	0.05												

Field ID	Depth	Date	Lab Report	Matrix																									
BH102	2	21 May 2024	352254	Soil	<1	<0.2	<0.5	<1	<2	<1	<1	<25	<25	<50	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DUP02	2	21 May 2024	352254	Soil	<1	<0.2	<0.5	<1	<2	<1	<1	<25	<25	<50	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
RPD					0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
BH102	2	21 May 2024	352254	Soil	<1	<0.2	<0.5	<1	<2	<1	<1	<25	<25	<50	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
TRIPO2		21 May 2024	ES2417505	Soil	<1	<0.2	<0.5	<0.5	<0.5	<0.5	<0.5	<10	<10	<50	<50	<100	<100	<50	-	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	
RPD					0	0	0	0	0	0	0	0	0	0	0	0	0	0	-	0	0	0	0	0	0	0	0		
BH107	3	21 May 2024	352254	Soil	<1	<0.2	<0.5	<1	<2	<1	<1	<25	<25	<50	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DUP03	3	21 May 2024	352254	Soil	<1	<0.2	<0.5	<1	<2	<1	<1	<25	<25	<50	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05		
RPD					0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		
BH107	3	21 May 2024	352254	Soil	<1	<0.2	<0.5	<1	<2	<1	<1	<25	<25	<50	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
TRIP03		21 May 2024	ES2417505	Soil	<1	<0.2	<0.5	<0.5	<0.5	<0.5	<0.5	<10	<10	<50	<50	<100	<100	<50	-	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	
RPD					0	0	0	0	0	0	0	0	0	0	0	0	0	0	-	0	0	0	0	0	0	0	0		

\*RPDs have only been considered where a concentration is greater than 1 times the EQL.

\*\*Elevated RPDs are highlighted as per QAQC Profile settings (Acceptable RPDs for each EQL multiplier range are: 100 (0 - 5 x EQL); 75 (5 - 10 x EQL); 30 (> 10 x EQL))

\*\*\*Interlab Duplicates are matched on a per compound basis as methods vary between laboratories. Any methods in the row header relate to those used in the primary laboratory

**Table 4**  
QA/QC Analytical Results Summary  
Soil RPD

	PAH												Metals								
	Benzol(g,h,i)perylene	Chrysene	Dibenz(a,h)anthracene	Fluoranthene	Fluorene	Indeno[1,2,3-c,d]pyrene	Naphthalene	Phenanthrene	Pyrene	Benzol(a)pyrene TEQ calc (Half)	Benzol(a)pyrene TEQ (LOR)	PAHs (Sum of positives)	Arsenic	Cadmium	Chromium (III+VI)	Copper	Iron	Lead	Mercury	Nickel	Zinc
	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg
EQL	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.5	0.5	0.05	4	0.4	1	1	10	1	0.1	1	1

Field ID	Depth	Date	Lab Report	Matrix	PAH												Metals							
BH102	2	21 May 2024	352254	Soil	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	6	<0.4	12	5	27,000	13	<0.1	<1	3
DUP02	2	21 May 2024	352254	Soil	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	5	<0.4	10	3	-	11	<0.1	<1	3
RPD					0	0	0	0	0	0	0	0	0	0	0	18	0	18	50	-	17	0	0	0
BH102	2	21 May 2024	352254	Soil	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	6	<0.4	12	5	27,000	13	<0.1	<1	3
TRIPO2		21 May 2024	ES2417505	Soil	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<5	<1	8	<5	-	18	<0.1	<2	<5
RPD					0	0	0	0	0	0	0	0	0	0	0	18	0	40	0	-	32	0	0	0
BH107	3	21 May 2024	352254	Soil	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<4	<0.4	7	11	-	24	<0.1	3	45
DUP03	3	21 May 2024	352254	Soil	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<4	<0.4	7	13	-	25	<0.1	3	42
RPD					0	0	0	0	0	0	0	0	0	0	0	0	0	0	17	-	4	0	0	7
BH107	3	21 May 2024	352254	Soil	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<4	<0.4	7	11	-	24	<0.1	3	45
TRIP03		21 May 2024	ES2417505	Soil	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<0.5	<5	<1	8	17	-	46	<0.1	6	58
RPD					0	0	0	0	0	0	0	0	0	0	0	0	0	13	43	-	63	0	67	25

\*RPDs have only been considered where a concentration is greater than 1 times the EQL.

\*\*Elevated RPDs are highlighted as per QAQC Profile settings (Acceptable RPDs for each EQL multiplier range are: 100 (0 - 5 x EQL); 75 (5 - 10 x EQL); 30 (> 10 x EQL))

\*\*\*Interlab Duplicates are matched on a per compound basis as methods vary between laboratories. Any methods in the row header relate to those used in the primary laboratory

	BTEX							TRH							Perfluoroalkane Carboxylic Acids		(n:2) Fluorotelomer Sulfonic Acids	
	Naphthalene (VOC)	Benzene	Toluene	Ethylbenzene	Xylene (m & p)	Xylene (o)	C6-C10 Fraction (F1)	C6-C10 (F1 minus BTEX)	>C10-C16 Fraction (F2)	>C10-C16 Fraction (F2 minus Naphthalene)	>C16-C34 Fraction (F3)	>C34-C40 Fraction (F4)	>C10-C40 Fraction (Sum)	Perfluorooctanoic acid (PFOA)	6:2 Fluorotelomer sulfonic acid (6:2 FTS)	8:2 Fluorotelomer sulfonic acid (8:2 FTS)		
	mg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L		
EQL	0.001	1	1	1	2	1	10	10	50	50	100	100	50	0.0002	0.0004	0.0004		

Field ID	Date	Lab Report	Matrix Type	Analytical Results (µg/L)															
EW1	31 May 2024	352854-A	Water	<0.001	<1	<1	<1	<2	<1	<10	<10	<50	<50	<100	<100	<50	<0.0002	<0.0004	<0.0004
DUP2	31 May 2024	352854-A	Water	-	-	-	-	-	-	-	-	-	-	-	-	-	<0.0002	<0.0004	<0.0004
RPD				-	-	-	-	-	-	-	-	-	-	-	-	0	0	0	
EW1	31 May 2024	352854-A	Water	<0.001	<1	<1	<1	<2	<1	<10	<10	<50	<50	<100	<100	<50	<0.0002	<0.0004	<0.0004
TRIP2	31 May 2024	ES2418204	Water	-	-	-	-	-	-	-	-	-	-	-	-	<0.01	<0.05	<0.05	
RPD				-	-	-	-	-	-	-	-	-	-	-	-	0	0	0	
MW1-B	31 May 2024	352854-A	Water	<0.001	<1	<1	<1	<2	<1	<10	<10	<50	<50	<100	<100	<50	0.0023	0.0007	<0.0004
DUP1	31 May 2024	352854-A	Water	<0.001	<1	<1	<1	<2	<1	<10	<10	<50	<50	<100	<100	<50	0.0021	0.0005	<0.0004
RPD				0	0	0	0	0	0	0	0	0	0	0	0	9	33	0	
MW1-B	31 May 2024	352854-A	Water	<0.001	<1	<1	<1	<2	<1	<10	<10	<50	<50	<100	<100	<50	0.0023	0.0007	<0.0004
TRIP1	31 May 2024	ES2418204	Water	<0.005	<1	<2	<2	<2	<2	<20	<20	<100	<100	<100	<100	<100	<0.01	<0.05	<0.05
RPD				0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	

\*RPDs have only been considered where a concentration is greater than 1 times the EQL.

\*\*Elevated RPDs are highlighted as per QAQC Profile settings (Acceptable RPDs for each EQL multiplier range are: 100 (0 - 5 x EQL); 75 (5 - 10 x EQL); 30 (> 10 x EQL))

\*\*\*Interlab Duplicates are matched on a per compound basis as methods vary between laboratories. Any methods in the row header relate to those used in the primary laboratory

	Perfluoroalkane Sulfonic Acids		PFAS			Metals												
	Perfluorohexane sulfonic acid (PFHxS) µg/L	Perfluorooctane sulfonic acid (PFOS) µg/L	Sum of PFHxS and PFOS µg/L	Sum of PFAS µg/L	Sum of PFOS + pFOA µg/L	Arsenic (filtered) mg/L	Cadmium (filtered) mg/L	Chromium (III+VI) (filtered) mg/L	Copper (filtered) mg/L	Lead (filtered) mg/L	Mercury (filtered) mg/L	Nickel (filtered) mg/L	Zinc (filtered) mg/L	Benzene (b+j+k)fluoranthene µg/L	Acenaphthene µg/L	Acenaphthylene µg/L	Anthracene µg/L	Benzo(a)anthracene µg/L
						µg/L	µg/L	µg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	µg/L	µg/L	µg/L	µg/L
EQL	0.0002	0.0002	0.0002	0.0002	0.0002	0.001	0.0001	0.001	0.001	0.001	0.00005	0.001	0.001	0.0002	0.1	0.1	0.1	0.1

Field ID	Date	Lab Report	Matrix Type																	
EW1	31 May 2024	352854-A	Water	<0.0002	<0.0002	<0.0002	<0.0002	<0.0002	<0.001	<0.0001	<0.001	<0.001	<0.001	<0.00005	<b>0.12</b>	<b>0.068</b>	<0.0002	<0.1	<0.1	<0.1
DUP2	31 May 2024	352854-A	Water	<0.0002	<b>0.0002</b>	<b>0.0002</b>	<b>0.0002</b>	<b>0.0002</b>	-	-	-	-	-	-	-	-	-	-	-	-
RPD				0	0	0	0	0	-	-	-	-	-	-	-	-	-	-	-	-
EW1	31 May 2024	352854-A	Water	<0.0002	<0.0002	<0.0002	<0.0002	<0.0002	<0.001	<0.0001	<0.001	<0.001	<0.001	<0.00005	<b>0.12</b>	<b>0.068</b>	<0.0002	<0.1	<0.1	<0.1
TRIP2	31 May 2024	ES2418204	Water	<0.01	<0.01	<0.01	-	-	-	-	-	-	-	-	-	-	-	-	-	-
RPD				0	0	0	-	-	-	-	-	-	-	-	-	-	-	-	-	-
MW1-B	31 May 2024	352854-A	Water	<b>0.0081</b>	<b>0.001</b>	<b>0.0094</b>	<b>0.012</b>	<b>0.0035</b>	<b>0.002</b>	<0.0001	<0.001	<0.001	<0.001	<0.00005	<b>0.002</b>	<b>0.009</b>	<0.0002	<0.1	<0.1	<0.1
DUP1	31 May 2024	352854-A	Water	<b>0.0074</b>	<b>0.001</b>	<b>0.0085</b>	<b>0.011</b>	<b>0.0031</b>	<b>0.002</b>	<0.0001	<0.001	<0.001	<0.001	<0.00005	<b>0.002</b>	<b>0.006</b>	<0.0002	<0.1	<0.1	<0.1
RPD				9	0	10	9	12	0	0	0	0	0	0	0	40	0	0	0	0
MW1-B	31 May 2024	352854-A	Water	<b>0.0081</b>	<b>0.001</b>	<b>0.0094</b>	<b>0.012</b>	<b>0.0035</b>	<b>0.002</b>	<0.0001	<0.001	<0.001	<0.001	<0.00005	<b>0.002</b>	<b>0.009</b>	<0.0002	<0.1	<0.1	<0.1
TRIP1	31 May 2024	ES2418204	Water	<0.01	<0.01	<0.01	-	-	<0.001	<0.0001	<0.001	<0.001	<0.001	<0.0001	<b>0.002</b>	<0.005	-	<1.0	<1.0	<1.0
RPD				0	0	0	-	-	67	0	0	0	0	0	0	57	-	0	0	0

\*RPDs have only been considered where a concentration is greater than 1 times the EQL.

\*\*Elevated RPDs are highlighted as per QAQC Profile settings (Acceptable RPDs for each EQL multiplier range are: 100 (0 - 5 x EQL); 75 (5 - 10 x EQL); 30 (> 10 x EQL))

\*\*\*Interlab Duplicates are matched on a per compound basis as methods vary between laboratories. Any methods in the row header relate to those used in the primary laboratory

	PAH												TPH				
	Benz[a] pyrene	Benz[e,h,i]perylene	Chrysene	Dibenz(a,h)anthracene	Fluoranthene	Fluorene	Indeno(1,2,3-c,d)pyrene	Naphthalene	Phenanthrene	Pyrene	Benz[a]pyrene TEQ	PAHs (Sum of positives)	C6-C9 Fraction	C10-C14 Fraction	C15-C28 Fraction	C29-C36 Fraction	C10-C36 Fraction (Sum)
	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	mg/L	mg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L
EQL	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.0005	0.0001	10	50	100	50	50	

Field ID	Date	Lab Report	Matrix Type															
EW1	31 May 2024	352854-A	Water	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.0005	<0.0001	<10	<50	<100	<100	<50
DUP2	31 May 2024	352854-A	Water	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
RPD				-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
EW1	31 May 2024	352854-A	Water	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.0005	<0.0001	<10	<50	<100	<100	<50
TRIP2	31 May 2024	ES2418204	Water	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
RPD				-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
MW1-B	31 May 2024	352854-A	Water	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.0005	<0.0001	<10	<50	<100	<100	<50
DUP1	31 May 2024	352854-A	Water	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.0005	<0.0001	<10	<50	<100	<100	<50
RPD				0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
MW1-B	31 May 2024	352854-A	Water	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.0005	<0.0001	<10	<50	<100	<100	<50
TRIP1	31 May 2024	ES2418204	Water	<0.5	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	<1.0	-	-	<20	<50	<100	<50	<50
RPD				0	0	0	0	0	0	0	0	-	-	0	0	0	0	0

\*RPDs have only been considered where a concentration is greater than 1 times the EQL.

\*\*Elevated RPDs are highlighted as per QAQC Profile settings (Acceptable RPDs for each EQL multiplier range are: 100 (0 - 5 x EQL); 75 (5 - 10 x EQL); 30 (> 10 x EQL))

\*\*\*Interlab Duplicates are matched on a per compound basis as methods vary between laboratories. Any methods in the row header relate to those used in the primary laboratory

	BTEX						TRH	
	Naphthalene (BTEX)	Benzene	Toluene	Ethylbenzene	Xylenes (m & p)	Xylenes (o)	Xylene Total	C6-C10 Fraction (F1)
	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg
EQL	1	0.2	0.5	1	2	1	1	25

Field ID	Date	Lab Report Number	Matrix Type	<1	<0.2	<0.5	<1	<2	<1	<1	<25	<25
Trip Blank	23 May 2024	352254	Soil									

	BTEX						TRH	
	Naphthalene (BTEX)	Benzene	Toluene	Ethylbenzene	Xylene (m & p)	Xylene (o)	Xylene Total	C6-C10 Fraction (F1)
	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg
EQL	1	0.2	0.5	1	2	1	1	25

Field ID	Date	Lab Report Number	Matrix Type	<1	<0.2	<0.5	<1	<2	<1	<1	<25	<25
Trip Blank	23 May 2024	352254	Soil									

**Table 8**  
 QA/QC Results Summary Tables

**Trip Spikes**

Lab Report Number	Matrix Type	Analysis Batch	Field ID	Sampled Date/Time	Chem Name	Spike Recovery %	Method Name	Lab Sample ID
352254	Soil	2024-05-29	Trip Spike	23/05/2024	Ethylbenzene	88	Org-023 - BTEX and C6-C10 alkanes in soil & water	352254-41
352254	Soil	2024-05-29	Trip Spike	23/05/2024	Xylene (m & p)	80	Org-023 - BTEX and C6-C10 alkanes in soil & water	352254-41
352254	Soil	2024-05-29	Trip Spike	23/05/2024	Toluene	87	Org-023 - BTEX and C6-C10 alkanes in soil & water	352254-41
352254	Soil	2024-05-29	Trip Spike	23/05/2024	Benzene	84	Org-023 - BTEX and C6-C10 alkanes in soil & water	352254-41
352254	Soil	2024-05-29	Trip Spike	23/05/2024	Xylene (o)	88	Org-023 - BTEX and C6-C10 alkanes in soil & water	352254-41

**Trip Spike Recoveries.** Where no lab LCL and UCL is available, user defined limits between 70% and 130% have been adopted for non-compliance.

Inorganics	Metals										BTEX								TRH																												
	Moisture Content %	Arsenic		Cadmium		Chromium (III+VI)		Copper		Lead		Mercury		Nickel		Zinc		Naphthalene (VOC)		Benzene		Toluene		Ethylbenzene		Xylene (m & p)		Xylene (o)		Xylene Total		Total BTEX		C6-C10 Fraction (F1)		C6-C10 (F1 minus BTEX)		>C10-C16 Fraction (F2)		>C10-C16 (F2 minus Naphthalene)		>C16-C34 Fraction (F3)		>C34-C40 Fraction (F4)		>C10-C40 Fraction (F5)	
		mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg									
EQL	0.1	4	0.3	1	1	1	0.1	1	1	1	1	1	1	1	1	1	1	3   3   3   3	0.5   0.5   0.5   0.5	360   220   330   540	55	40   60   95   170	230	260   370   630	45   70   110   200	110   240   440	100	100	50	100	50	100	100	50	100	50	100	100	50	100	50						
NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand																																															
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand																																															
NEPM 2013 Table 1A(1) HILs Res B Soil		500	150		30,000	1,200	120	1,200	60,000																																						
NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil		3,000	900		240,000	1,500	730	6,000	400,000																																						
ASSMAC 1998 >1000t disturbed, medium texture																																															

Field ID	Date	8.9	<4	<0.4	24	6	10	<0.1	4	20	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH1.1-0-2.0	06 Sep 2022	18	<4	<0.4	14	4	14	<0.1	2	6	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH4-1-0-2.0	06 Sep 2022	19	8	<0.4	11	4	10	<0.1	2	16	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH4-5-0-6.0	06 Sep 2022	29	6	<0.4	17	110	1,000	<0.1	4	190	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH5-1-0-2.0	06 Sep 2022	15	<4	<0.4	16	2	12	<0.1	2	6	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH6-0-0-1	06 Sep 2022	25	4	<0.4	21	9	17	<0.1	5	9	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH6-1-0-2.0	06 Sep 2022	11	<4	<0.4	8	4	10	<0.1	<1	3	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH7-0-0-0.1	06 Sep 2022	15	7	<0.4	23	67	82	<0.1	19	83	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH7-1-0-2.0	06 Sep 2022	10	<4	<0.4	23	28	15	<0.1	4	27	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BR1	06 Sep 2022	26	<4	<0.4	25	5	17	<0.1	9	45	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH1_0-1	06 Sep 2022	12	<4	<0.4	18	5	16	<0.1	8	40	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	<100	<100	<50	<25	<25	<50	<50	<100	<100	<50
DS1.BH2_0-0-1	28 Sep 2022	15	<4	<0.4	2	8	3	<0.1	<1	2	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25	<25	<50	<50	190	190	<100	<100	190	190	<100	<100	190	
DS1.BH2_1-9-2.0	28 Sep 2022	20	<4	<0.4	8	2	14	<0.1	1	7	<1	<0.2	<0.5	<1	<2	<1	<1	<1	<25													

	TPH						PAH																			Benzof[b+g]fluoranthene		Acenaphthene		Acenaphthylenne		Anthracene		Benz[a]anthracene		Benz[a]pyrene		Benz[b]fluoranthene		Benz[e]pyrene		Chrysene		Diphen[a,h,i]anthracene		Fluoranthene		Fluorene		Indeno[1,2,3-c,d]pyrene		Naphthalene		Phenanthrene		Pyrene		PAHs (Sum of positives)	
	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg																																															
	25	50	100	100	50	0.2	0.1	0.1	0.1	0.1	0.05	0.3	0.1	0.3	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.05																							
EQL																																																											
NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand																																																											
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand																																																											
NEPM 2013 Table 1A(1) HILs Res B Soil																																																											
NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil																																																											
ASSMAC 1998 >1000t disturbed, medium texture																																																											

Field ID	Date	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05
DS1.BH1.1-0-2.0	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05
DS1.BH4-1.0-2.0	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BH4-5.0-6.0	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BH5-1.0-2.0	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BH5-6.0-7.0	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BH6-0-0.1	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BH6-1.0-2.0	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BH7-0.0-0.1	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BH7-1.0-2.0	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BR1	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BH1_0-1	06 Sep 2022	<25	<50	<100	<100	<50	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.05	
DS1.BH2_0-0.1	28 Sep 2022	<25	<50	100	<100	100	<0.2	<0.1	<0.1	<0.1	<0.05	-	<0.1	-	<0.1	<0.1															

	Chlorinated Hydrocarbons																								
	1,1,1,2-tetrachloroethane	1,1,1-trichloroethane	1,1,2,2-tetrachloroethane	1,1,2-trichloroethane	1,1-dichloroethane	1,1-dichloropropene	1,2,3-trichloropropene	1,2-dibromo-3-chloropropane	1,2-dichloroethane	1,2-dichloropropene	1,3-dichloropropene	Bromochloromethane	Bromodichloromethane	Bromoform	Carbon tetrachloride	Chloroethane	Chlorofrom	Chloromethane	cis-1,2-dichloroethene	dibromomethane	hexachlorobutadiene	Trichloroethylene	tetrachloroethylene	trans-1,2-dichloroethene	
	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg
EQL	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand																									
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand																									
NEPM 2013 Table 1A(1) HILs Res B Soil																									
NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil																									
ASSMAC 1998 >1000t disturbed, medium texture																									

Field ID	Date																									
DS1.BH1-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH4-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH4-5.0-6.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH5-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH5-6.0-7.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH6-0-0.1	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH6-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH7-0.0-0.1	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH7-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BR1	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH1_0-1	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH2_0-0.1	28 Sep 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	
DS1.BH2_1.9-2.0	28 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BR	28 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.SR	28 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1_BH3_0-0.1	28 Sep 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	
DS1_BH3_1.9-2.0	28 Sep 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

#### Environmental Standards

2013, NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand

2013, NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand

2013, NEPM 2013 Table 1A(1) HILs Res B Soil

2013, NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil

	Halogenated Benzenes												Halogenated Hydrocarbons				Perfluoroalkane Carboxylic Acids									
	trans-1,3-dichloropropene	Vinyl chloride	1,2,3-trichlorobenzene	1,2,4-trichlorobenzene	1,2-dichlorobenzene	1,3-dichlorobenzene	1,4-dichlorobenzene	2-chlorotoluene	4-chlorotoluene	Bromobenzene	Chlorobenzene	Hexachlorobenzene	1,2-dibromoethane	Bromomethane	Dichlorodifluoromethane	Trichlorofluoromethane	Perfluorobutanoic acid (PFBA)	Perfluorohexanoic acid (PFHA)	Perfluoropentanoic acid (PFPeA)	Perfluorohexanoic acid (PFHpA)	Perfluoroctanoic acid (PFOA)	Perfluorodecanoic acid (PFDoA)	Perfluorononanoic acid (PFNA)	Perfluorotetradecanoic acid (PFTeDA)	Perfluorotridecanoic acid (PFTeDA)	Perfluoroundecanoic acid (PFUnDA)
	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg
EQL	1	1	1	1	1	1	1	1	1	1	1	0.1	1	1	1	1	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005	0.005
NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand																										
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand																										
NEPM 2013 Table 1A(1) HILs Res B Soil													15													
NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil													80													
ASSMAC 1998 >1000t disturbed, medium texture																										

Field ID	Date																									
DS1.BH1-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH4-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH4-5.0-6.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH5-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH5-6.0-7.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH6-0-0.1	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH6-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH7-0.0-0.1	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH7-1.0-2.0	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BR1	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH1_0-1	06 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH2_0-0.1	28 Sep 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.1	<1	<1	<1	<1	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	<0.005	
DS1.BH2_1.9-2.0	28 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BR	28 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.SR	28 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.10	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1_BH3_0-0.1	28 Sep 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.1	<1	<1	<1	<1	-	-	<0.005	-	-	-	-	-	-	
DS1_BH3_1.9-2.0	28 Sep 2022	-	-	-	-	-	-	-	-	-	-	<0.1	-	-	-	-	-	-	-	<0.005	-	-	-	-	-	

#### Environmental Standards

2013, NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand

2013, NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand

2013, NEPM 2013 Table 1A(1) HILs Res B Soil

2013, NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil



## **Environmental Standards**

2013, NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand

2013, NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand

#### 2013- NEPM 2013 Table 1A(1) Hils Bes B Soil

2013, NERM 2013 Table 1A(1) HILs Res B Soil

Field ID	Date	Organochlorine Pesticides																																							
		tert-butylbenzene		4,4-DDE		a-BHC		Aldrin		b-BHC		Chlordane (cis)		Chlordane (trans)		DDD		DDT+DDE+DDD		Dieldrin		Endosulfan I		Endosulfan II		Endosulfan sulphate		Endrin		Endrin aldehyde		Endrin ketone		g-BHC (Lindane)		Heptachlor		Heptachlor epoxide		Methoxychlor	
		mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg								
EQL		1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1	0.1								
NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand																																									
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand																																									
NEPM 2013 Table 1A(1) HILs Res B Soil																																									
NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil																																									
ASSMAC 1998 >1000t disturbed, medium texture																																									

Field ID	Date	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH1-1.0-2.0	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH4-1.0-2.0	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH4-5.0-6.0	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH5-1.0-2.0	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH5-6.0-7.0	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH6-0.0-1	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH6-1.0-2.0	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH7-0.0-1	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH7-1.0-2.0	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BR1	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH1_0-1	06 Sep 2022	-	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH2_0-0.1	28 Sep 2022	<1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1
DS1.BH2_1.9-2.0	28 Sep 2022	-</td																												

## **Environmental Standards**

2013, NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand

2013, NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand

2013, NEPM 2013 Table 1A(1) HILs Res B Soil

2013, NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil

	Pesticides		Solvents	Acid Sulphate Soils - Acid Base Accounting			Acid Sulphate Soils - Acidity Trial		Acid Sulphate Soils - Liming Rate		Acid Sulphate Soils - pH	Acid Sulphate Soils - Potential Acidity	Acid Sulphate Soils - Retained Acidity	Acid Sulphate Soils - KCl Extractable Sulfur	Acid Sulphate Soils		
	DEF	Parathion	Cyclohexane	a-Net Acidity without ANCE	Net Acidity (acidity units)	Net Acidity (sulfur units)	Titratable Actual Acidity (sulfur units)	Titratable Actual Acidity	Liming Rate	Liming Rate excluding ANC	pH/KCl	Chromium Reducible Sulfur (acidity units)	HCl Extractable Sulfur	Net Acid Soluble Sulfur	KCl Extractable Sulfur	% Net Acidity without ANCE	
		mg/kg	mg/kg	mg/kg	moles H+/t	moles H+/t	%S	%S	mole H+/t	kg CaCO <sub>3</sub> /t	kg CaCO <sub>3</sub> /t	-	mole H+/t	%w/w	%S	%	% S
EQL		0.1	0.1	1	5	5	0.005	0.01	5	0.75	0.75	-	3	0.005	0.005	0.005	0.005
NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand																	
NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand																	
NEPM 2013 Table 1A(1) HILs Res B Soil																	
NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil																	
ASSMAC 1998 >1000t disturbed, medium texture							18	0.03						0.03			

Field ID	Date	-	<0.1	-	28	28	0.046	<0.01	<5	2	2.1	5.1	24	0.04	-	-	0.046	
DS1.BH1-1.0-2.0	06 Sep 2022	-	<0.1	-	28	28	0.046	<0.01	<5	2	2.1	5.1	24	0.04	-	-	0.046	
DS1.BH4-1.0-2.0	06 Sep 2022	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH4-5.0-6.0	06 Sep 2022	-	<0.1	-	11	11	0.017	0.01	8	0.8	0.80	5.7	<3	0.005	-	-	0.017	
DS1.BH5-1.0-2.0	06 Sep 2022	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH5-6.0-7.0	06 Sep 2022	-	<0.1	-	13	13	0.021	<0.01	<5	1	0.97	6.2	11	0.02	-	-	0.021	
DS1.BH6-0-0.1	06 Sep 2022	-	<0.1	-	120	120	0.19	0.18	110	9.0	9.0	4.3	6	0.009	0.015	<0.005	0.018	0.19
DS1.BH6-1.0-2.0	06 Sep 2022	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH7-0.0-0.1	06 Sep 2022	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH7-1.0-2.0	06 Sep 2022	-	<0.1	-	28	28	0.044	0.04	23	2	2.1	4.8	5	0.008	-	-	0.044	
DS1.BR1	06 Sep 2022	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH1_0-1	06 Sep 2022	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH2_0-0.1	28 Sep 2022	-	<0.1	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.BH2_1.9-2.0	28 Sep 2022	-	<0.1	-	64	64	0.10	0.07	42	5	4.8	4.5	22	0.04	-	-	0.10	
DS1.BR	28 Sep 2022	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1.SR	28 Sep 2022	<0.10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1_BH3_0-0.1	28 Sep 2022	-	<0.1	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	
DS1_BH3_1.9-2.0	28 Sep 2022	-	<0.1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

**Environmental Standards**

2013, NEPM 2013 Table 1A(3) Comm/Ind D Soil HSL for Vapour Intrusion, Sand

2013, NEPM 2013 Table 1A(3) Res A/B Soil HSL for Vapour Intrusion, Sand

2013, NEPM 2013 Table 1A(1) HILs Res B Soil

2013, NEPM 2013 Table 1A(1) HILs Comm/Ind D Soil

	Metals										Herbicides	Perfluoroalkane Carboxylic Acids	(n:2) Fluorotelomer Sulfonic Acids		Perfluoroalkane Sulfonic Acids		PFAS			Dioxins and Furans		
	Arsenic (filtered)		Cadmium (filtered)		Chromium (III+VI) (filtered)		Copper (filtered)		Lead (filtered)				(n:2) Fluorotelomer Sulfonic Acids		Perfluoroalkane Sulfonic Acids		PFAS					
	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L			mg/L	μg/L	mg/L	μg/L	mg/L	μg/L	mg/L	μg/L		
EQL	0.001	0.0001	0.001	0.001	0.001	0.00005	0.001	0.001	10	0.01	0.01	0.02	0.01	0.01	0.01	0.01	0.01	0.01	0.01	5		
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)		0.0002		0.0014	0.0034	0.0006	0.011	0.008														
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)		0.0055		0.0013	0.0044	0.0004	0.07	0.015														
PFAS NEMP 2020 Freshwater 99%										19						0.00023						
PFAS NEMP 2020 Interim Marine 99%										19						0.00023						
PFAS NEMP 2020 Recreational Water										10						2	2	2				
NEPM 2013 Table 1C GILs, Fresh Waters		0.0002		0.0014	0.0034	0.00006	0.011	0.008														
NEPM 2013 Table 1C GILs, Marine Waters		0.0007		0.0013	0.0044	0.0001	0.007	0.015														

Field ID	Date	<0.001	<0.0001	<0.001	<0.001	<0.001	<0.001	<0.00005	0.004	0.025	<10	<0.01	<0.01	<0.02	<0.01	0.02	0.02	0.02	0
Dup01	12 Oct 2022	<0.001	<0.0001	<0.001	<0.001	<0.001	<0.001	<0.00005	0.004	0.028	<10	<0.01	<0.01	<0.02	<0.01	0.02	0.02	0.02	0
Dup01A	12 Oct 2022	<0.001	<0.0001	<0.001	<0.001	<0.001	<0.001	<0.00005	0.004	0.028	<10	<0.01	<0.01	<0.02	<0.01	0.02	0.02	0.02	0
MW1	12 Oct 2022	<0.001	<0.0001	<0.001	<0.001	<0.001	<0.001	<0.00005	0.008	0.044	<10	<0.01	<0.01	<0.02	0.02	<0.01	0.02	0.02	<0.01
MW2	12 Oct 2022	0.003	<0.0001	<0.001	0.001	<0.001	<0.001	<0.00005	0.024	0.034	<10	<0.01	0.04	<0.02	<0.01	<0.01	0.04	<0.01	0
MW3	12 Oct 2022	<0.001	<0.0001	<0.001	<0.001	<0.001	<0.001	<0.00005	0.004	0.026	<10	<0.01	<0.01	<0.02	<0.01	0.02	0.02	0.02	0
MW4	12 Oct 2022	0.001	<0.0001	<0.001	0.004	<0.001	<0.001	<0.00005	<0.001	0.019	<10	<0.01	0.01	<0.02	<0.01	<0.01	0.01	<0.01	0
Rinsate	12 Oct 2022	<0.001	<0.0001	<0.001	<0.001	<0.001	<0.001	<0.00005	<0.001	<0.001	<10	<0.01	<0.01	<0.02	<0.01	<0.01	<0.01	<0.01	0
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

note to table on PFAS LOR

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

2,3,7,8-e  
Tetrachlorooxanthene

	Naphthalene (VOC)	BTEX												TRH												3&4-Methylphenol (m&p-cresol)						
		Benzene			Toluene			Ethylbenzene			Xylene (m & p)			Xylene (o)			C6-C10 Fraction (F1)			C6-C10 (F1 minus BTEX)			>C10-C16 Fraction (F2)			>C16-C34 Fraction (F3)			>C34-C40 Fraction (F4)			
		mg/L	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%	µg/L	%		
EQL		0.001	1	1	1	1	1	1	2	2	1	1	10	10	50	50	100	100	100	100	50	50	4									
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)		0.016	950		180		80				350																					
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)		0.07	700		180		80																									
PFAS NEMP 2020 Freshwater 99%																																
PFAS NEMP 2020 Interim Marine 99%																																
PFAS NEMP 2020 Recreational Water																																
NEPM 2013 Table 1C GILs, Fresh Waters		0.016	950								350																					
NEPM 2013 Table 1C GILs, Marine Waters		0.05	500																													

Field ID	Date	BTEX												TRH														
Dup01	12 Oct 2022	<0.001	<1	-	<1	-	<1	-	<2	-	<1	-	40	40	93	93	550	220	860	<4								
Dup01A	12 Oct 2022	<0.001	<1	-	<1	-	<1	-	<2	-	<1	-	37	37	74	74	530	240	840	<4								
MW1	12 Oct 2022	<0.001	<1	-	<1	-	<1	-	<2	-	<1	-	<10	<10	85	85	<100	<100	80	<4								
MW2	12 Oct 2022	<0.001	<1	-	<1	-	<1	-	<2	-	3	-	50	47	250	250	110	<100	360	<4								
MW3	12 Oct 2022	<0.001	<1	-	<1	-	<1	-	<2	-	<1	-	35	35	72	72	600	250	930	<4								
MW4	12 Oct 2022	<0.001	<1	-	<1	-	<1	-	<2	-	<1	-	27	27	<50	<50	<100	<100	<50	<4								
Rinsate	12 Oct 2022	<0.001	<1	-	<1	-	<1	-	<2	-	<1	-	<10	<10	<50	<50	<100	<100	<50	<4								
TB	12 Oct 2022	<0.001	<1	-	<1	-	<1	-	<2	-	<1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
TS	12 Oct 2022	-	-	118	-	114	-	113	-	111	-	110	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
note to table on PFAS LOR																												

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2022)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

	Phenols															Amino Aliphatics		Amino Aromati											
	2,3,4,6-Tetrachlorophenol		2,4,5-Trichlorophenol		2,4,6-Trichlorophenol		2,4-Dichlorophenol		2,4-Dimethylphenol		2,6-Dichlorophenol		2-Chlorophenol		2-Methylphenol		4,6-Dinitro-2-methylphenol		4-chloro-3-methylphenol		4-Nitrophenol		Pentachlorophenol		Phenol				
	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	mg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L		
EQL	2	2	2	2	2	0.02	2	2	2	2	20	10	20	10	2	5	5	5	5	5	5	5	5	5	5	5			
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)	20		20	160		0.045		490								10	320												
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)																22	400												
PFAS NEMP 2020 Freshwater 99%																													
PFAS NEMP 2020 Interim Marine 99%																													
PFAS NEMP 2020 Recreational Water																													
NEPM 2013 Table 1C GILs, Fresh Waters	10		3	120		0.045		340								3.6	320												
NEPM 2013 Table 1C GILs, Marine Waters																11	400												

Field ID	Date	<2	<2	<2	<2	<2	<0.02	<2	<2	<2	<2	<20	<10	<20	<10	<2	<5	<5	<5	<5	<5	<5	<5	<5	<5	<5
Dup01	12 Oct 2022	<2	<2	<2	<2	<2	<0.02	<2	<2	<2	<2	<20	<10	<20	<10	<2	<5	<5	<5	<5	<5	<5	<5	<5	<5	<5
Dup01A	12 Oct 2022	<2	<2	<2	<2	<2	<0.02	<2	<2	<2	<2	<20	<10	<20	<10	<2	<5	<5	<5	<5	<5	<5	<5	<5	<5	<5
MW1	12 Oct 2022	<2	<2	<2	<2	<2	<0.02	<2	<2	<2	<2	<20	<10	<20	<10	<2	<5	<5	<5	<5	<5	<5	<5	<5	<5	<5
MW2	12 Oct 2022	<2	<2	<2	<2	<2	<0.02	<2	<2	<2	<2	<20	<10	<20	<10	<2	<5	<5	<5	<5	<5	<5	<5	<5	<5	<5
MW3	12 Oct 2022	<2	<2	<2	<2	<2	<0.02	<2	<2	<2	<2	<20	<10	<20	<10	<2	<5	<5	<5	<5	<5	<5	<5	<5	<5	<5
MW4	12 Oct 2022	<2	<2	<2	<2	<2	<0.02	<2	<2	<2	<2	<20	<10	<20	<10	<2	<5	<5	<5	<5	<5	<5	<5	<5	<5	<5
Rinsate	12 Oct 2022	<2	<2	<2	<2	<2	<0.02	<2	<2	<2	<2	<20	<10	<20	<10	<2	<5	<5	<5	<5	<5	<5	<5	<5	<5	<5
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

note to table on PFAS LOR

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2020)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

CS	Diphenylamine	Anilines																																	
		2-nitroaniline		3-nitroaniline		4-chloroaniline		4-nitroaniline		2-methyl-5-nitroaniline		Aniline		1,1,1,2-tetrachloroethane		1,1,1,1-trichloroethane		1,1,2,2-tetrachloroethane		1,1,2-trichloroethane		1,1-dichloroethane		1,1-dichloropropene		1,2,3-trichloropropene		1,2-dibromo-3-chloropropane		1,2-dichloroethane		1,2-dichloropropane		1,3-dichloropropane	
		µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L				
EQL		5	5	5	5	5	5	5	5	250		1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1					
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)												270	400	6,500		700												1,900	900	1,100					
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)												270	400	1,900		700												1,900	900	1,100					
PFAS NEMP 2020 Freshwater 99%																																			
PFAS NEMP 2020 Interim Marine 99%																																			
PFAS NEMP 2020 Recreational Water																																			
NEPM 2013 Table 1C GILs, Fresh Waters												8								6,500															
NEPM 2013 Table 1C GILs, Marine Waters																				1,900															

Field ID	Date																										
Dup01	12 Oct 2022	<5	<5	<5	<5	<5	<5	<5	<5	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1
Dup01A	12 Oct 2022	<5	<5	<5	<5	<5	<5	<5	<5	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1
MW1	12 Oct 2022	<5	<5	<5	<5	<5	<5	<5	<5	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1
MW2	12 Oct 2022	<5	<5	<5	<5	<5	<5	<5	<5	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1
MW3	12 Oct 2022	<5	<5	<5	<5	<5	<5	<5	<5	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1
MW4	12 Oct 2022	<5	<5	<5	<5	<5	<5	<5	<5	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1
Rinsate	12 Oct 2022	<5	<5	<5	<5	<5	<5	<5	<5	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

note to table on PFAS LOR

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2022)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

	Chlorinated Hydrocarbons																	
	2,2-dichloropropane		Bromochloromethane		Bromodichloromethane		Bromoform		Carbon tetrachloride		Chlorodibromomethane		Chloroethane		Chloroform		Chloromethane	
	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L
EQL	1	1	1	1	1	1	10	1	10	1	1	1	1	5	2	1	1	1
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)					240			770						360	330	70		
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)					240			770						330	70			
PFAS NEMP 2020 Freshwater 99%																		
PFAS NEMP 2020 Interim Marine 99%																		
PFAS NEMP 2020 Recreational Water																		
NEPM 2013 Table 1C GILs, Fresh Waters														290				
NEPM 2013 Table 1C GILs, Marine Waters																		

Field ID	Date	<1	<1	<1	<1	<1	<1	<10	24	<10	<1	<1	<1	<1	<5	<2	<1	<1	<1
Dup01	12 Oct 2022	<1	<1	<1	<1	<1	<1	<10	24	<10	<1	<1	<1	<1	<5	<2	<1	<1	<1
Dup01A	12 Oct 2022	<1	<1	<1	<1	<1	<1	<10	27	<10	<1	<1	<1	<1	<5	<2	<1	<1	<1
MW1	12 Oct 2022	<1	<1	<1	<1	<1	<1	<10	2	<10	<1	<1	<1	<1	<5	<2	<1	<1	<1
MW2	12 Oct 2022	<1	<1	<1	<1	<1	<1	<10	3	<10	<1	<1	<1	<1	<5	<2	<1	<1	<1
MW3	12 Oct 2022	<1	<1	<1	<1	<1	<1	<10	21	<10	<1	<1	<1	<1	<5	<2	<1	<1	<1
MW4	12 Oct 2022	<1	<1	<1	<1	<1	<1	<10	3	<10	<1	<1	<1	<1	<5	<2	9	<1	<1
Rinsate	12 Oct 2022	<1	<1	<1	<1	<1	<1	<10	<1	<10	<1	<1	<1	<1	<5	<2	<1	<1	<1
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

note to table on PFAS LOR

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

	Vinyl chloride	Explosives			Halogenated Benzenes												Halogenated Hydrocarbons																		
		1,3-Dinitrobenzene		2,6-dinitrotoluene		Nitrobenzene		1,2,3-trichlorobenzene		1,2,4,5-tetrachlorobenzene		1,2-dichlorobenzene		1,3-dichlorobenzene		1,4-dichlorobenzene		2-chlorotoluene		4-chlorotoluene		Bromobenzene		Chlorobenzene		Hexachlorobenzene		Pentachlorobenzene		1,2-dibromoethane		Bromomethane		Dichlorodifluoromethane	
		µg/L	mg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L				
EQL		10	0.005	5	5	1	2	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	2	2	1	10	10						
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)		100				550	10	7	170	160	260	60													55	0.1	2								
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)		100						5	80																55	0.1	2								
PFAS NEMP 2020 Freshwater 99%																																			
PFAS NEMP 2020 Interim Marine 99%																																			
PFAS NEMP 2020 Recreational Water																																			
NEPM 2013 Table 1C GILs, Fresh Waters								3		85	160	260	60																						
NEPM 2013 Table 1C GILs, Marine Waters									20																										

Field ID	Date	<10	<0.005	<5	<5	<1	<2	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<10	<10
Dup01	12 Oct 2022	<10	<0.005	<5	<5	<1	<2	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<10	<10
Dup01A	12 Oct 2022	<10	<0.005	<5	<5	<1	<2	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<10	<10	
MW1	12 Oct 2022	<10	<0.005	<5	<5	<1	<2	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<10	<10	
MW2	12 Oct 2022	<10	<0.005	<5	<5	<1	<2	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<10	<10	
MW3	12 Oct 2022	<10	<0.005	<5	<5	<1	<2	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<10	<10	
MW4	12 Oct 2022	<10	<0.005	<5	<5	<1	<2	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<10	<10	
Rinsate	12 Oct 2022	<10	<0.005	<5	<5	<1	<2	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<10	<10	
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-		
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-		

note to table on PFAS LOR

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2020)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

	Trichlorofluoromethane	MAH												Nitroaromatics																										
		1,2,4-trimethylbenzene		1,3,5-trimethylbenzene		Isopropylbenzene		n-butylbenzene		n-propylbenzene		p-isopropyltoluene			sec-butylbenzene		Styrene		tert-butylbenzene				Pentachloronitrobenzene		4,4-DDE		a-BHC		Aldrin		b-BHC		Chlordane (cis)		Chlordane (trans)		d-BHC		DDD	
		µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L							
EQL		10	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	5	2	2	2	2	2	2	2	2	2	2	2	2									
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)						30																																		
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)						30																																		
PFAS NEMP 2020 Freshwater 99%																																								
PFAS NEMP 2020 Interim Marine 99%																																								
PFAS NEMP 2020 Recreational Water																																								
NEPM 2013 Table 1C GILs, Fresh Waters																																								
NEPM 2013 Table 1C GILs, Marine Waters																																								

Field ID	Date																											
Dup01	12 Oct 2022	<10	1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
Dup01A	12 Oct 2022	<10	1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
MW1	12 Oct 2022	<10	8	4	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
MW2	12 Oct 2022	<10	18	10	1	<1	1	<1	1	<1	<1	<1	<1	<1	<1	<1	<1	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
MW3	12 Oct 2022	<10	1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
MW4	12 Oct 2022	<10	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
Rinsate	12 Oct 2022	<10	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

note to table on PFAS LOR

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2020)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

	Organochlorine Pesticides																																				
	DDT		Dieldrin		Endosulfan I		Endosulfan II		Endosulfan sulphate		Endrin		Endrin aldehyde		$\gamma$ -BHC (Lindane)		Heptachlor		Heptachlor epoxide		Methoxychlor		Azinophos methyl		Bromophos-ethyl		Chlorpyrifos-methyl		Coumaphos		Diazinon		Dichlorvos		Dimethoate		
	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L		µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	mg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L					
EQL	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	0.002	2	2	2	2	2	2								
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)	0.01								0.02						0.2	0.09											0.01			0.15							
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)									0.008																												
PFAS NEMP 2020 Freshwater 99%																																					
PFAS NEMP 2020 Interim Marine 99%																																					
PFAS NEMP 2020 Recreational Water																																					
NEPM 2013 Table 1C GILs, Fresh Waters	0.006								0.01						0.2	0.01											0.01			0.15							
NEPM 2013 Table 1C GILs, Marine Waters									0.004																												

Field ID	Date	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
Dup01	12 Oct 2022	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
Dup01A	12 Oct 2022	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
MW1	12 Oct 2022	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
MW2	12 Oct 2022	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
MW3	12 Oct 2022	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
MW4	12 Oct 2022	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
Rinsate	12 Oct 2022	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

note to table on PFAS LOR

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2022)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

	Organophosphorous Pesticides												Other						Benzene		Benzo(b+j+k)fluoranthene		2-chloronaphthalene		2-methylnaphthalene		3-methylcholanthrene		7,12-dimethylbenz(a)anthracene											
	Disulfoton		Ethion		Ethyl methanesulfonate		Fentrothion		Fenthion		Malathion		Methidathion		Methyl parathion		Mevinphos (Phosdrin)		Phorate		Ronnel		Safrole		Acetophenone		Phosalone		Benzene		Benzo(b+j+k)fluoranthene		2-chloronaphthalene		2-methylnaphthalene		3-methylcholanthrene		7,12-dimethylbenz(a)anthracene	
	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L	mg/L							
EQL	2	2	5	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	5	5	0.002	0.002	2	2	2	2	2	2										
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)				0.2			0.05																																	
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)																																								
PFAS NEMP 2020 Freshwater 99%																																								
PFAS NEMP 2020 Interim Marine 99%																																								
PFAS NEMP 2020 Recreational Water																																								
NEPM 2013 Table 1C GILs, Fresh Waters				0.2			0.05																																	
NEPM 2013 Table 1C GILs, Marine Waters																																								

Field ID	Date																												
Dup01	12 Oct 2022	<2	<2	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<5	<5	<0.002	<0.002	<2	<2	<2	<2	
Dup01A	12 Oct 2022	<2	<2	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<5	<5	<0.002	<0.002	<2	<2	<2	<2	
MW1	12 Oct 2022	<2	<2	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<5	<5	<0.002	<0.002	<2	<2	<2	<2	
MW2	12 Oct 2022	<2	<2	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<5	<5	<0.002	<0.002	<2	<2	<2	<2	
MW3	12 Oct 2022	<2	<2	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<5	<5	<0.002	<0.002	<2	<2	<2	<2	
MW4	12 Oct 2022	<2	<2	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<5	<5	<0.002	<0.002	<2	<2	<2	<2	
Rinsate	12 Oct 2022	<2	<2	<5	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<2	<5	<5	<0.002	<0.002	<2	<2	<2	<2	
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
note to table on PFAS LOR																													

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2020)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

	PAH																		
	Acenaphthene	Acenaphthylene	Anthracene	Benzo(a)anthracene	Benzo(a) pyrene	Benzo(g,h,i)perylene	Chrysene	Dibenz(a,h)anthracene	Fluoranthene	Fluorene	Indeno(1,2,3-c,d)pyrene	Naphthalene	Phenanthrene	Pyrene	Benzo(a)pyrene TEQ	PAHs (Sum of positives)	Arochlor 1016	Arochlor 1221	Arochlor 1232
	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	mg/L	µg/L	µg/L	µg/L	µg/L
EQL	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0.005	0.001	2	2	2
ANZG (2018) Freshwater 95% LOSP Toxicant DGVs (March 2021)			0.4		0.2				1.4			16	2						
ANZG (2018) Marine Water 95% LOSP Toxicant DGVs (March 2021)			0.4		0.2				1.4			70	2						
PFAS NEMP 2020 Freshwater 99%																			
PFAS NEMP 2020 Interim Marine 99%																			
PFAS NEMP 2020 Recreational Water																			
NEPM 2013 Table 1C GILs, Fresh Waters												16							
NEPM 2013 Table 1C GILs, Marine Waters												50							

Field ID	Date	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.005	0	<2	<2	<2
Dup01	12 Oct 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.005	0	<2	<2	<2
Dup01A	12 Oct 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.005	0	<2	<2	<2
MW1	12 Oct 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.005	0	<2	<2	<2
MW2	12 Oct 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.005	0	<2	<2	<2
MW3	12 Oct 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.005	0	<2	<2	<2
MW4	12 Oct 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.005	0	<2	<2	<2
Rinsate	12 Oct 2022	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<1	<0.005	0	<2	<2	<2
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

note to table on PFAS LOR

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGVs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGVs (March 2020)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

	PCBs				Pesticides				Phthalates							Solvents																						
	Arochlor 1242		Arochlor 1248		Arochlor 1254		Arochlor 1260		Carbazole		Fenamiphos		Parathion		Bis(2-ethylhexyl) phthalate		Butyl benzyl phthalate		Diethyl phthalate		Dimethyl phthalate		Di-n-butyl phthalate		Di-n-octyl phthalate		Cyclohexane		Isophorone		2-(acetylamo) fluorene		4-(dimethylamino) azobenzene		4-bromophenyl phenyl ether		4-chlorophenyl phenyl ether	
	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	mg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L	µg/L							
EQL	2	2	2	2	5	2	2	50	10	10	10	10	10	10	10	0.001	5	2	5	5	5	5	0.001	5	2	5	5	5	5	5								
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)	0.6		0.03					0.004			1,000	3,700	26																									
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)																																						
PFAS NEMP 2020 Freshwater 99%																																						
PFAS NEMP 2020 Interim Marine 99%																																						
PFAS NEMP 2020 Recreational Water																																						
NEPM 2013 Table 1C GILs, Fresh Waters	0.3		0.01					0.004			1,000	3,700	10																									
NEPM 2013 Table 1C GILs, Marine Waters																																						

Field ID	Date																											
Dup01	12 Oct 2022	<2	<2	<2	<2	<5	<2	<2	<50	<10	<10	<10	<10	<10	<10	<10	<0.001	<5	<2	<5	<5	<5						
Dup01A	12 Oct 2022	<2	<2	<2	<2	<5	<2	<2	<50	<10	<10	<10	<10	<10	<10	<10	<0.001	<5	<2	<5	<5	<5						
MW1	12 Oct 2022	<2	<2	<2	<2	<5	<2	<2	<50	<10	<10	<10	<10	<10	<10	<10	<0.001	<5	<2	<5	<5	<5						
MW2	12 Oct 2022	<2	<2	<2	<2	<5	<2	<2	<50	<10	<10	<10	<10	<10	<10	<10	<0.001	<5	<2	<5	<5	<5						
MW3	12 Oct 2022	<2	<2	<2	<2	<5	<2	<2	<50	<10	<10	<10	<10	<10	<10	<10	<0.001	<5	<2	<5	<5	<5						
MW4	12 Oct 2022	<2	<2	<2	<2	<5	<2	<2	<50	<10	<10	<10	<10	<10	<10	<10	<0.001	<5	<2	<5	<5	<5						
Rinsate	12 Oct 2022	<2	<2	<2	<2	<5	<2	<2	<50	<10	<10	<10	<10	<10	<10	<10	<0.001	<5	<2	<5	<5	<5						
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-						
note to table on PFAS LOR																												

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2020)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

	SVOCs												TPH						VOCs
	Azobenzene µg/L	Benzyl alcohol µg/L	Bis(2-chloroethoxy) methane µg/L	Bis(2-chloroethyl)ether µg/L	Bis(2-chloroisopropyl) ether µg/L	Dibenzofuran µg/L	Hexachloropropene µg/L	Isosafrole µg/L	Methapyriline µg/L	N-nitrosomorpholine µg/L	N-nitrosopiperidine µg/L	Phenacetin µg/L	C6-C9 Fraction µg/L	C10-C14 Fraction µg/L	C15-C28 Fraction µg/L	C29-C36 Fraction (sum) µg/L	C10-C36 Fraction µg/L		
EQL	5	5	5	5	5	5	2	5	10	5	5	5	10	50	100	100	50	2	
ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)																		80	
ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)																		80	
PFAS NEMP 2020 Freshwater 99%																			
PFAS NEMP 2020 Interim Marine 99%																			
PFAS NEMP 2020 Recreational Water																			
NEPM 2013 Table 1C GILs, Fresh Waters																			
NEPM 2013 Table 1C GILs, Marine Waters																			

Field ID	Date																		
Dup01	12 Oct 2022	<5	<5	<5	<5	<5	<5	<2	<5	<10	<5	<5	<5	22	100	240	450	790	<2
Dup01A	12 Oct 2022	<5	<5	<5	<5	<5	<5	<2	<5	<10	<5	<5	<5	18	81	230	400	710	<2
MW1	12 Oct 2022	<5	<5	<5	<5	<5	<5	<2	<5	<10	<5	<5	<5	<10	95	<100	<100	100	<2
MW2	12 Oct 2022	<5	<5	<5	<5	<5	<5	<2	<5	<10	<5	<5	<5	23	340	140	<100	480	<2
MW3	12 Oct 2022	<5	<5	<5	<5	<5	<5	<2	<5	<10	<5	<5	<5	16	79	260	460	810	<2
MW4	12 Oct 2022	<5	<5	<5	<5	<5	<5	<2	<5	<10	<5	<5	<5	17	<50	<100	<100	<50	<2
Rinsate	12 Oct 2022	<5	<5	<5	<5	<5	<5	<2	<5	<10	<5	<5	<5	<10	<50	<100	<100	<50	<2
TB	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
TS	12 Oct 2022	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

note to table on PFAS LOR

\* A Non Detect Multiplier of 0.5 has been applied.

#### Environmental Standards

ANZG, March 2021, ANZG (2018) Freshwater 95% LOSP Toxicant DGvs (March 2021)

ANZG, March 2021, ANZG (2018) Marine Water 95% LOSP Toxicant DGvs (March 2021)

HEPA, January 2020, PFAS NEMP 2020 Freshwater 99%

HEPA, January 2020, PFAS NEMP 2020 Interim Marine 99%

HEPA, January 2020, PFAS NEMP 2020 Recreational Water

2013, NEPM 2013 Table 1C GILs, Fresh Waters

2013, NEPM 2013 Table 1C GILs, Marine Waters

PFAS Detections. PFHxS, PFOS, PFOA, and sum of PFHxS and PFOS are have regulatory guidelines for TCC.

*NT* Not Tested

PFAS Detections, PFHxS, PEOS, PFOA, and sum of PFHxS and PEOS are have regulatory guidelines for ASI

NT Not Tested

PFAS Detections. PFHxS, PFOS, PFOA, and sum of PFHxS and PFOS are have regulatory guidelines for ASLE

NT Not Tested

**Refer to PFAS NEMP 2.0 for further information**

Notes:  
1 - Sum of PCCD/F + PCB TEQ results to the nearest integer  
2 - Site specific HLL derived by Jacobs

Table A. Soil Analytical Results

Method_Type	ChemName	Units	EQL	NEPM 2013 HIL A Soil	NEPM 2013 HSL A & B - Soil Vapour Intrusion, 0-1m	CRC Care HSL A - Direct Contact	NEPM 2013 ESLs - Urban residential and public open space	NEPM 2013 Management Limits - Residential, parkland & open space (Coarse)	NEPM 2013 EILs - Urban residential and public open space	Field_ID	BH08_0.1-0.3	BH08_0.9-1.1	BH08_2.0	BH09_0.3-0.5	BH09_3.3-3.5	BH10_0.1-0.2	BH11_0.9-1.0	BH12_1.1-1.3	BH12_5.3-5.5	BH13_0.55-0.75	BH13_2.5-2.7	BH14_1.3-1.5	BH14_1.95-2.15	BH15_0.8-1.1	BH16_0.3-0.5	
										LocCode	BH08	BH08	BH08	BH09	BH09	BH10	BH11	BH12	BH12	BH13	BH13	BH14	BH14	BH15	BH16	
										Sample_Depth_Range	0.1-0.3	0.9-1.1	2	0.3-0.5	3.3-3.5	0.1-0.2	0.9-1	1.1-1.3	5.3-5.5	0.55-0.75	2.5-2.7	1.3-1.5	1.95-2.15	0.8-1.1	0.3-0.5	
										Sampled_Date-Time	18/08/2016	18/08/2016	17/08/2016	16/08/2016	16/08/2016	16/08/2016	16/08/2016	16/08/2016	16/08/2016	16/08/2016	16/08/2016	19/08/2016	19/08/2016	19/08/2016	19/08/2016	
Matrix_Description																										
Soil																										
Soil properties	Moisture Content (dried @ 103°C)	%	1							-	-	6.2	-	30	8.2	29	-	34	-	30	-	27	15	11		
	Cation Exchange Capacity	meq/100g	1							-	-	-	-	-	-	-	-	-	-	-	-	5.2	-			
	% Clay	%	1							-	-	-	-	-	-	-	-	-	-	-	-	46	-			
	pH (Lab)	pH_Units	0.1							-	-	-	-	-	-	-	-	-	-	-	-	4.2	-			
Dioxins & Furans	WHO (2005)-PCDD/F TEQ (upper-bound)	ng/kg dw	-							-	65.3	29.9	-	6.05	-	-	-	8.61	-	-	-	-	-	-		
	WHO (2005)-PCB TEQ (upper-bound)	ng/kg dw	-							-	0.366	0.215	-	0.18	-	-	-	0.637	-	-	-	-	-	-		
	WHO (2005)-Sum of PCDD/F + PCB TEQ (upper-bound) <sup>1</sup>	ng/kg dw	-		120 <sup>2</sup>					-	66	30	-	6	-	-	-	9	-	-	-	-	-	-		
Asbestos	Approximate Sample Mass	g	-							185	-	-	238	-	203	112	154	-	176	-	254	-	109	178		
	Asbestos from ACM in Soil	%w/w	-							0	-	-	0	-	0	0	0	-	0	-	0	0	0	0		
	Asbestos from FA & AF in Soil	%w/w	-							0	-	-	0	-	0	0	0	-	0	-	0	0	0	0		
	Mass ACM	g	-							0	-	-	0	-	0	0	0	-	0	-	0	0	0	0		
	Mass AF	g	-							0	-	-	0	-	0	0	0	-	0	-	0	0	0	0		
	Mass Asbestos in ACM	g	-							0	-	-	0	-	0	0	0	-	0	-	0	0	0	0		
	Mass Asbestos in AF	g	-							0	-	-	0	-	0	0	0	-	0	-	0	0	0	0		
	Mass Asbestos in FA	g	-							0	-	-	0	-	0	0	0	-	0	-	0	0	0	0		
	Mass Asbestos in FA & AF	g	-							0	-	-	0	-	0	0	0	-	0	-	0	0	0	0		
	Mass FA	g	-							0	-	-	0	-	0	0	0	-	0	-	0	0	0	0		
Heavy Metals	Arsenic	mg/kg	2	100						100	-	-	3.6	-	10	5.7	9.5	-	13	-	21	-	5.2	10	5.9	
	Cadmium	mg/kg	0.4	20						3	-	-	<0.4	-	<0.4	<0.4	<0.4	-	<0.4	-	<0.4	-	<0.4	<0.4	<0.4	
	Chromium (III+VI)	mg/kg	5	100						410	-	-	10	-	<5	13	17	-	9.5	-	7.1	-	9.4	15	9.4	
	Copper	mg/kg	5	6000						65	-	-	<5	-	<5	<5	<5	-	<5	-	<5	-	<5	19		
	Lead	mg/kg	5	300						1110	-	-	7.5	-	5.5	10	64	-	5.6	-	<5	-	11	10	35	
	Mercury	mg/kg	0.05	40						1	-	-	<0.05	-	<0.05	0.08	0.07	-	<0.05	-	<0.05	-	<0.05	<0.05	<0.05	
	Nickel	mg/kg	5	400						35	-	-	<5	-	<5	<5	<5	-	<5	-	<5	-	<5	<5	8.8	
	Zinc	mg/kg	5	7400						75	-	-	17	-	10	13	88	-	5.9	-	7.1	-	5	49		
PAHs in soil	Benz[a]anthracene	mg/kg	0.5							-	-	-	<0.5	-	<0.5	<0.5	<0.5	-	<0.5	-	<0.5	-	<0.5	<0.5	<0.5	
	Acenaphthene	mg/kg	0.5							-	-	-	<0.5	-	<0.5	<0.5	<0.5	-	<0.5	-	<0.5	-	<0.5	<0.5	<0.5	
	Acenaphthylene	mg/kg	0.5							-	-	-	<0.5	-	<0.5	<0.5	<0.5	-	<0.5	-	<0.5	-	<0.5	<0.5	<0.5	
	Anthracene	mg/kg	0.5							-	-	-	<0.5	-	<0.5	<0.5	<0.5	-	<0.5	-	<0.5	-	<0.5	<0.5	<0.5	
	Benz[a]pyrene	mg/kg	0.5							0.7	-	-	<0.5	-	<0.5	<0.5	<0.5	-	<0.5	-	<0.5	-	<0.5	<0.5	<0.5	
	Benz[a]pyrene TEQ (lower bound)*	ng/kg	0.5							-	-	-	<0.5	-	<0.5	<0.5	<0.5	-	<0.5	-	<0.5	-	<0.5	<0.5	<0.5	
	Benz[a]pyrene TEQ (medium bound)*	ng/kg	0.5							3	-	-	0.6	-	0.6	0.6	0.6	-	0.6</							

		Field_ID	MW01	MW03	MW04	MW07	MW09	MW12	MW13	MW14
	LocCode	BH01/MW01	BH03/MW03	MW04/MW04	BH09/MW07	BH09/MW09	BH12/MW12	BH13/MW13	BH14/MW14	
	WellCode	MW01	MW03	MW04	MW07	MW09	MW12	MW13	MW14	
	Sampled_Date-Time	24/08/2016	24/08/2016	24/08/2016	25/08/2016	25/08/2016	25/08/2016	25/08/2016	25/08/2016	
	NEPM 2013 HSL A & B, Clay, 2-4m	NEPM 2013 GILs - Marine Waters								
Method_Type	ChemName	Units	EQL							
Heavy Metals	Arsenic (Filtered)	µg/L	1		2	1	1	3	<1	<1
	Cadmium (Filtered)	µg/L	0.2		0.7	0.3	<0.2	<0.2	<0.2	<0.2
	Chromium (III+VI) (Filtered)	µg/L	1		<1	<1	<1	<1	<1	<1
	Copper (Filtered)	µg/L	1		1.3	100	2	4	<1	16
	Lead (Filtered)	µg/L	1		4.4	9	5	<1	2	2
	Mercury (Filtered)	µg/L	0.1		0.1	<0.1	<0.1	<0.1	<0.1	<0.1
	Nickel (Filtered)	µg/L	1		7	66	10	10	11	74
SVOC	Zinc (Filtered)	µg/L	1		15	280	14	11	41	7
	2,4,5-trichlorophenol	µg/L	10		<10	<10	<10	<10	<10	<10
	2,4,6-trichlorophenol	µg/L	10		<10	<10	<10	<10	<10	<10
	2,4-dichlorophenol	µg/L	3		<3	<3	<3	<3	<3	<3
	2,4-dimethylphenol	µg/L	3		<3	<3	<3	<3	<3	<3
	2,4-dinitrophenol	µg/L	30		<30	<30	<30	<30	<30	<30
	2,6-dichlorophenol	µg/L	3		<3	<3	<3	<3	<3	<3
	2-chlorophenol	µg/L	3		<3	<3	<3	<3	<3	<3
	2-methylphenol	µg/L	3		<3	<3	<3	<3	<3	<3
	2-nitrophenol	µg/L	10		<10	<10	<10	<10	<10	<10
	3-&4-methylphenol	µg/L	6		<6	<6	<6	<6	<6	<6
	4,6-Dinitro-2-methylphenol	µg/L	30		<30	<30	<30	<30	<30	<30
	4,6-Dinitro-o-cyclohexyl phenol	µg/L	100		<100	<100	<100	<100	<100	<100
	4-chloro-3-methylphenol	µg/L	10		<10	<10	<10	<10	<10	<10
	4-nitrophenol	µg/L	30		<30	<30	<30	<30	<30	<30
	Dinoseb	µg/L	100		<100	<100	<100	<100	<100	<100
	Pentachlorophenol	µg/L	10		11	<10	<10	<10	<10	<10
	Phenol	µg/L	3		400	<3	<3	<3	<3	<3
	tetrachlorophenols	µg/L	30		<30	<30	<30	<30	<30	<30
	Phenols (Total Halogenated)	µg/L	10		<10	<10	<10	<10	<10	<10
	Phenols (Total Non Halogenated)	µg/L	100		<100	<100	<100	<100	<100	<100
TRH in water NEPM 2013	C6-C10	µg/L	20		<20	<20	<20	<20	<20	<20
	C6-C10 less BTEX (F1)	µg/L	20	NL	<20	<20	<20	<20	<20	<20
	C10-C16	µg/L	50		<50	<50	<50	<50	<50	<50
	C10-C16 less Naphthalene (F2)	µg/L	50	NL	<50	<50	<50	<50	<50	<50
	C16-C34	µg/L	100		<100	<100	<100	<100	<100	<100
	C34-C40	µg/L	100		<100	<100	<100	<100	<100	<100
TRH in water NEPM 1999	C6 - C9	µg/L	20		<20	<20	<20	<20	<20	<20
	C10 - C14	µg/L	50		<50	<50	<50	<50	<50	<50
	C15 - C28	µg/L	100		<100	<100	<100	<100	100	<100
	C29-C36	µg/L	100		<100	<100	<100	<100	<100	<100
	+C10 - C36 (Sum of total)	µg/L	100		<100	<100	<100	<100	100	<100
BTEXN in water NEPM 2013	Benzene	µg/L	1	5000	500	<1	<1	<1	<1	<1
	Toluene	µg/L	1	NL	<1	<1	<1	<1	<1	<1
	Ethylbenzene	µg/L	1	NL	<1	<1	<1	<1	<1	<1
	Xylene (m & p)	µg/L	2		<2	<2	<2	<2	<2	<2
	Xylene (o)	µg/L	1		<1	<1	<1	<1	<1	<1
	Xylene Total	µg/L	3	NL	<3	<3	<3	<3	<3	<3
	Naphthalene	µg/L	10	NL	50	<10	<10	<10	<10	<10

Table B. Groundwater Analytical Results

 Rhodes East Priority Investigation Area  
 October 2016

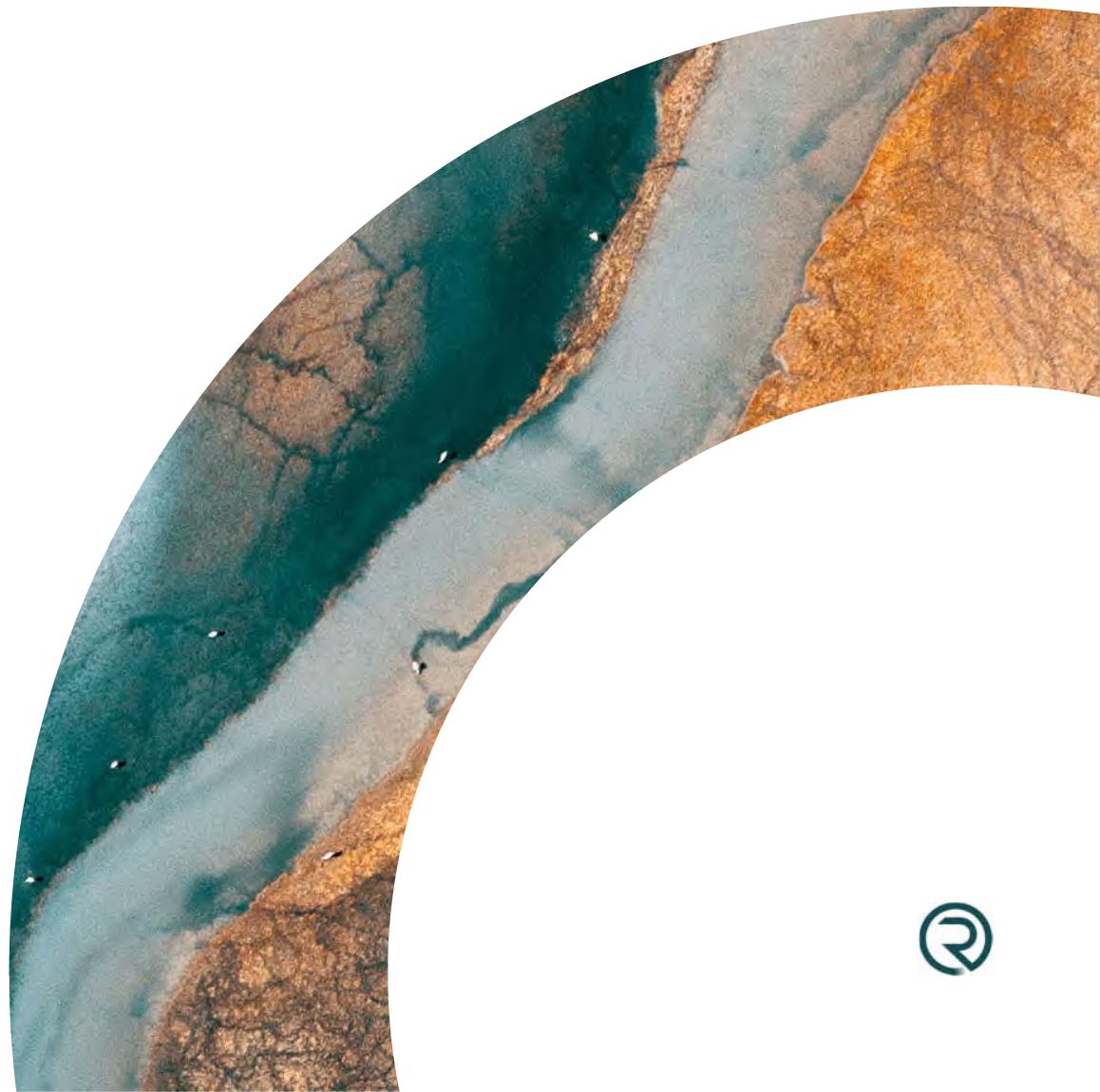
		Field_ID	MW01	MW03	MW04	MW07	MW09	MW12	MW13	MW14
	LocCode	BH01/MW01	BH03/MW03	MW04/MW04	BH09/MW07	BH09/MW09	BH12/MW12	BH13/MW13	BH14/MW14	
	WellCode	MW01	MW03	MW04	MW07	MW09	MW12	MW13	MW14	
	Sampled_Date-Time	24/08/2016	24/08/2016	24/08/2016	25/08/2016	25/08/2016	25/08/2016	25/08/2016	25/08/2016	
NEPM 2013 HSL A & B, Clay, 2-4m		NEPM 2013 GILs - Marine Waters								
Method_Type	ChemName	Units	EQL							
VOC	1,1,1,2-tetrachloroethane	µg/L	1		<1	<1	<1	<1	<1	<1
	1,1,1-trichloroethane	µg/L	1		<1	<1	<1	<1	<1	<1
	1,1,2,2-tetrachloroethane	µg/L	1		<1	<1	<1	<1	<1	<1
	1,1,2-trichloroethane	µg/L	1	1900	<1	<1	<1	<1	<1	<1
	1,1-dichloroethane	µg/L	1		<1	<1	<1	<1	<1	<1
	1,1-dichloroethene	µg/L	1		<1	<1	<1	<1	<1	<1
	1,2,3-trichloropropane	µg/L	1		<1	<1	<1	<1	<1	<1
	1,2,4-trimethylbenzene	µg/L	1		<1	<1	<1	<1	<1	<1
	1,2-dibromoethane	µg/L	1		<1	<1	<1	<1	<1	<1
	1,2-dichlorobenzene	µg/L	1		<1	<1	<1	<1	<1	<1
	1,2-dichloroethane	µg/L	1		<1	<1	<1	<1	<1	<1
	1,2-dichloropropane	µg/L	1		<1	<1	<1	<1	<1	<1
	1,3,5-trimethylbenzene	µg/L	1		<1	<1	<1	<1	<1	<1
	1,3-dichlorobenzene	µg/L	1		<1	<1	<1	<1	<1	<1
	1,3-dichloropropane	µg/L	1		<1	<1	<1	<1	<1	<1
	1,4-dichlorobenzene	µg/L	1		<1	<1	<1	<1	<1	<1
	Methyl Ethyl Ketone	µg/L	1		<1	<1	<1	<1	<1	<1
	4-chlorotoluene	µg/L	1		<1	<1	<1	<1	<1	<1
	4-Methyl-2-pentanone	µg/L	1		<1	<1	<1	<1	<1	<1
	Acetone	µg/L	1		6	5	<1	<1	11	47
	Allyl chloride	µg/L	1		<1	<1	<1	<1	<1	<1
	Bromobenzene	µg/L	1		<1	<1	<1	<1	<1	<1
	Bromochloromethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Bromodichloromethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Bromoform	µg/L	1		<1	<1	<1	<1	<1	<1
	Bromomethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Carbon disulfide	µg/L	1		<1	<1	<1	<1	<1	<1
	Carbon tetrachloride	µg/L	1		<1	<1	<1	<1	<1	<1
	Chlorobenzene	µg/L	1		<1	<1	<1	<1	<1	<1
	Chlorodibromomethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Chloroethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Chloroform	µg/L	5		<5	<5	<5	<5	<5	<5
	Chloromethane	µg/L	1		<1	<1	<1	<1	<1	<1
	cis-1,2-dichloroethene	µg/L	1		<1	<1	<1	<1	<1	<1
	cis-1,3-dichloropropene	µg/L	1		<1	<1	<1	<1	<1	<1
	Dibromomethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Dichlorodifluoromethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Dichloromethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Iodomethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Isopropylbenzene	µg/L	1		<1	<1	<1	<1	<1	<1
	Styrene	µg/L	1		<1	<1	<1	<1	<1	<1
	Trichloroethene	µg/L	1		<1	<1	<1	<1	<1	<1
	Tetrachloroethene	µg/L	1		<1	<1	<1	<1	<1	<1
	trans-1,2-dichloroethene	µg/L	1		<1	<1	<1	<1	<1	<1
	trans-1,3-dichloropropene	µg/L	1		<1	<1	<1	<1	<1	<1
	Trichlorofluoromethane	µg/L	1		<1	<1	<1	<1	<1	<1
	Vinyl chloride	µg/L	1		<1	<1	<1	<1	<1	<1

# D

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## Borehole Logs

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Phone: (02) 9521 6567

Boring No.: BH105

Sheet : 1 OF 1

Easting : 0.00 Drill Supplier : Stratacore Drilling Pty Ltd Job Number : 22148  
Northing : 0.00 Driller Company : Stratacore Drilling Pty Ltd Client : Billbergia Pty Ltd  
Elevation : Not Surveyed Logged By : Renee Ashton Project : Detailed Site Investigation  
Total Depth : 6.3 m Date : 21/05/2024 Location : 25-27 Leeds Street, Rhodes NSW

Drilling Method	Depth (m)	Samples	PID (ppm)	Soil Origin	Graphic Log	Moisture	Material Description	Comments	Water
		Discrete	PID (ppm)						
				Non-Soil			Concrete		
		0.3	1.3	Fill		SLM	Fill. Gravelly SAND SP: loose, dark brown, coarse grained, coarse sized gravel, slightly moist, anthropogenic material including brick and concrete present..	Anthropogenic material including brick and concrete present.	
		0.5	0.3				Fill. Silty SAND SM: loose, brown, medium grained, with medium sized gravel, slightly moist, anthropogenic material including brick and concrete present..	Anthropogenic material including brick and concrete present.	
	1	1.0	2.8				As above, but brown red, trace low plasticity clay.	Anthropogenic material including brick and concrete present.	
	2	2.0	0.3	Natural			Natural. SAND SP: medium dense, orange brown, fine grained, slightly moist.	Weathered sandstone inclusions, crushed sandstone possibly used as fill?	
	3	3.0	0.2			M	Natural. Sandy CLAY CL-Cl: soft, low to medium plasticity, brown orange, fine to medium grained sand, inorganic, moist, minor sandstone inclusions.	Minor sandstone inclusions	
	4	4.0	0.2			W	Natural. Sandy CLAY CH: soft, high plasticity, grey, fine grained sand, organic, wet.		
	5	5.0	0.2			M	Natural. Sandy CLAY CI: soft, medium plasticity, pale grey, fine grained sand, inorganic, moist.		
	6	6.0	0.2	Rock		SLM	Rock. SANDSTONE: highly weathered, low strength, pale grey, fine grained, indistinct, slightly moist.		
							BH105 refusal at 6.3m (Refusal on rock at 6.3 m)		

Reditus Consulting Pty Ltd							Boring No.: BH106		
Lvl 1, Suite 1/29-33 Waratah St, Kirrawee NSW 2232, Australia Phone: (02) 9521 6567									
Easting : 0.00 Northing : 0.00 Elevation : Not Surveyed Total Depth : 7 m			Drill Supplier : Stratacore Drilling Pty Ltd Driller Company : Stratacore Drilling Pty Ltd Logged By : Renee Ashton Date : 22/05/2024			Job Number : 22148 Client : Bilbergia Pty Ltd Project : Detailed Site Investigation Location : 25-27 Leeds Street, Rhodes NSW		Sheet : 1 OF 1	
Drilling Method	Depth (m)	Samples	PID (ppm)	Soil Origin	Graphic Log	Moisture	Material Description	Comments	Water
		Discrete	Trip	Dup	PID (ppm)				
							Concrete		
	0.3				0.2	Non-Soil			
	0.5				1.9	Fill	SLM	Fill. Silty to gravelly SAND SM: loose, dark brown black, coarse grained, coarse sized gravel, slightly moist.	
	1							Fill. Silty SAND SM: loose, brown, medium to coarse grained, with medium sized gravel, slightly moist, slight decrease in gravel and increase in sand at 0.9 m.	
	2				1.9	Natural	M	Natural. Sandy CLAY CI: firm, medium plasticity, pale grey brown mottled, fine grained sand, inorganic, moist.	
	3				0.2		W	Natural. Sandy CLAY CH: soft, high plasticity, dark grey, fine grained sand, organic, wet, carbonate materials (shells) present.	Carbonate materials (shells) present.
	4				0.2	Fill	M	Fill. Sandy CLAY CH: high plasticity, stiff to very stiff, dark brown orange, fine grained sand, inorganic, moist.	
	5				0.4	Natural		Natural. Clayey SAND SC: dense, medium plasticity clay, brown red, medium grained, moist.	Trace sandstone inclusions
	6				0.2			Natural. Sandy CLAY CL: firm, low plasticity, red grey mottled, medium grained sand, inorganic, moist.	
						Rock		Rock. SANDSTONE: highly weathered, low strength, red grey orange, medium grained, indistinct, moist. <b>BH106 refusal at 7m (Refusal on rock at 7.0 m)</b>	



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Boring No.: BH107

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Easting	: 0.00	Drill Supplier	: Stratacore Drilling Pty Ltd	Job Number	: 22148	Sheet	: 1 OF 1
Northing	: 0.00	Driller Company	: Stratacore Drilling Pty Ltd	Client	: Bilbergia Pty Ltd		
Elevation	: Not Surveyed	Logged By	: Renee Ashton	Project	: Detailed Site Investigation		
Total Depth	: 0.4 m	Date	: 23/05/2024	Location	: 25-27 Leeds Street, Rhodes NSW		
Drilling Method	Depth (m)	Samples	PID (ppm)	Soil Origin	Graphic Log	Moisture	Material Description
		Discrete	PID (ppm)	Non-Soil			Concrete
	0.3		0.2	Fill		SLM	Fill. Gravelly SAND SP: loose, brown, medium grained, medium sized gravel, slightly moist, tree roots and anthropogenic inclusions.
							BH108 refusal at 0.4m (Refusal on concrete at 0.4m. Did not proceed due to potential service.)
	1						
	2						
	3						
	4						
	5						
	6						



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**Boring No.: BH109**



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**Boring No.: BH110**

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Lvl 1, Suite 1/29-33 Waratah St, Kirrawee NSW 2232, Australia				Boring No.: BH111		
Phone: (02) 9521 6567						
Easting : 0.00	Drill Supplier : Stratacore Drilling Pty Ltd			Job Number : 22148		Sheet : 1 OF 1
Northing : 0.00	Driller Company : Stratacore Drilling Pty Ltd			Client : Bilbergia Pty Ltd		
Elevation : Not Surveyed	Logged By : Renee Ashton			Project : Detailed Site Investigation		
Total Depth : 4 m	Date : 23/05/2024			Location : 25-27 Leeds Street, Rhodes NSW		
Drilling Method	Depth (m)	Samples	PID (ppm)	Soil Origin	Graphic Log	Moisture
		Discrete	PID (ppm)			
				Non-Soil		
	0.3		0.7			
	0.5		3.7	Fill		SLM
	1		4.1			
	1.0					
						Concrete
						Fill. Silty SAND SM: loose, dark brown, medium grained, with medium sized gravel, slightly moist, hydrocarbon odour at 0.5 m.
						Fill. Clayey SAND SC: low plasticity clay, medium dense, dark brown brown, medium to coarse grained, slightly moist, increase in clay content from 0.8-1.5 m. hydrocarbon odour at 1.0 m.
						Trace clay inclusions
	2		1.4	Rock		
	2.0					
						Rock. SANDSTONE: moderately weathered, low strength, red brown, fine grained, indistinct, slightly moist, trace clay inclusions.
						Residual clay inclusions. Trace iron staining
	3		0.8			
	3.0					
						Extremely weathered, rock. SAND SST: very loose, yellow brown, fine grained, dry.
	4		0.6	D		
	4.0					BH111 refusal at 4m (Refusal at 4.0 m)
	5					
	6					



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Boring No.: BH112

Easting : 0.00 Drill Supplier : Stratacore Drilling Pty Ltd Job Number : 22148 Sheet : 1 OF 1  
Northing : 0.00 Driller Company : Stratacore Drilling Pty Ltd Client : Billbergia Pty Ltd  
Elevation : Not Surveyed Logged By : Renee Ashton Project : Detailed Site Investigation  
Total Depth : 1 m Date : 20/05/2024 Location : 25-27 Leeds Street, Rhodes NSW



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**Boring No.: BH113**



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Phone: (02) 9521 6567

Boring No.: BH114

<b>Sheet</b>	<b>: 1 OF 1</b>
<b>Easting</b>	<b>: 0.00</b>
<b>Northing</b>	<b>: 0.00</b>
<b>Elevation</b>	<b>: Not Surveyed</b>
<b>Total Depth</b>	<b>: 1.6 m</b>
<b>Drill Supplier</b>	<b>: Stratacore Drilling Pty Ltd</b>
<b>Driller Company</b>	<b>: Stratacore Drilling Pty Ltd</b>
<b>Logged By</b>	<b>: Renee Ashton</b>
<b>Date</b>	<b>: 20/05/2024</b>
<b>Job Number</b>	<b>: 22148</b>
<b>Client</b>	<b>: Billbergia Pty Ltd</b>
<b>Project</b>	<b>: Detailed Site Investigation</b>
<b>Location</b>	<b>: 25-27 Leeds Street, Rhodes NSW</b>

Reditus Consulting Pty Ltd							Boring No.: BH101/RMW01			
Lvl 1, Suite 1/29-33 Waratah St, Kirrawee NSW 2232, Australia Phone: (02) 9521 6567										
Easting : 0.00 Northing : 0.00 Elevation : Not Surveyed Total Depth : 6.8 m			Drill Supplier : Stratacore Drilling Pty Ltd Driller Company : Stratacore Drilling Pty Ltd Logged By : Renee Ashton Date : 21/05/2024			Job Number : 22148 Client : Bilbergia Pty Ltd Project : Detailed Site Investigation Location : 25-27 Leeds Street, Rhodes NSW		Sheet : 1 OF 1		
Drilling Method	Depth (m)	Samples	PbD (ppm)	Soil Origin	Graphic Log	Moisture	Material Description	Comments	Well Diagram	Water
				Non-Soil			Concrete			
				Fill		SL M	Fill. Gravelly SAND SP: very loose, dark brown, fine grained, fine sized gravel, slightly moist.			
				Reworked Natural			Reworked natural. Silty SAND SM: loose, pale yellow white, fine grained, slightly moist.			Backfill
	1			Natural		M	Natural. Clayey SAND SC: medium dense, low plasticity clay, pale brown, fine to medium grained, moist.			50mm PVC Solid
	2						Natural. Sandy CLAY CI: soft, medium plasticity, pale brown grey yellow, fine to medium grained sand, inorganic, moist.			Bentonite
	3					W	Natural. Sandy CLAY ML: soft, non-plastic, grey, fine to medium grained sand, organic, wet, carbonate materials (shells) present..	Carbonate materials (shells) present.		Washed 1-2mm graded sand
	4						Natural. Sandy CLAY CI-CH: firm, medium to high plasticity, orangey red, fine grained sand, organic, moist.			50mm PVC Slotted
	5									
	6									
							BH101/RMW01 Terminated at 6.8m (Target Depth)			



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Boring No.: BH102/RMW02

Easting	: 0.00	Drill Supplier	: Stratacore Drilling Pty Ltd	Job Number	: 22148
Northing	: 0.00	Driller Company	: Stratacore Drilling Pty Ltd	Client	: Billbergia Pty Ltd
Elevation	: Not Surveyed	Logged By	: Renee Ashton	Project	: Detailed Site Investigation
Total Depth	: 10.2 m	Date	: 21/05/2024	Location	: 25-27 Leeds Street, Rhodes NSW

Sheet : 1 OF 2

Drilling Method	Samples		PID (ppm)	Soil Origin	Graphic Log	Moisture	Material Description	Comments	Well Diagram	Water
	Depth (m)	Discrete								
Sonic	0.3	0.3	0.3	Non-Soil		M	Concrete			
	0.5	0.3					Fill. Gravelly SAND SW: loose, dark brown, medium to coarse grained, medium to coarse sized gravel, moist. Fill. Silty SAND SM: loose, pale brown, medium grained, with fine sized gravel, moist.			
	1.0	0.6					Natural. Clayey SAND SC: medium dense, low plasticity clay, reddish brown, medium grained, moist.			
	2.0	0.7					Natural. Clayey SAND SC: medium dense, low plasticity clay, brown pale brown, medium grained, moist.			
	3.0	0.7					Natural. Sandy CLAY CI-CH: soft, medium to high plasticity, mottled brown grey orange, medium grained sand, inorganic, moist.			
	4.0	0.3					Natural. Sandy CLAY ML: soft, non-plastic, grey, fine to medium grained sand, organic, wet, carbonate materials (shells) present..	Carbonate materials (shells) present.		
	5.0	0.3					Natural. Sandy CLAY ML: soft, non-plastic, orangey red, fine grained sand, organic, wet, carbonate materials (shells) present..	Carbonate materials (shells) present.		
	6.0	0.2					As above, but CI-CH: firm, medium to high plasticity, inorganic, moist.			
	7.0	0.2					Natural. SAND SP: loose, grey pale yellow, medium grained, moist.			



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Boring No.: BH102/RMW02

Easting	: 0.00	Drill Supplier	: Stratacore Drilling Pty Ltd	Job Number	: 22148
Northing	: 0.00	Driller Company	: Stratacore Drilling Pty Ltd	Client	: Billbergia Pty Ltd
Elevation	: Not Surveyed	Logged By	: Renee Ashton	Project	: Detailed Site Investigation
Total Depth	: 10.2 m	Date	: 21/05/2024	Location	: 25-27 Leeds Street, Rhodes NSW

Sheet : 2 OF 2

Drilling Method	Samples		Material Description	Comments	Well Diagram	Water
	Depth (m)	Discrete				
	7.2		Natural. SAND SP: loose, grey pale yellow, medium grained, moist.			
	7.3		Rock, SANDSTONE: slightly weathered, very low strength, pale grey, fine grained, distinct, wet.			
	8					
	9		Rock. SANDSTONE: slightly weathered, very low strength, yellow brown, fine grained, bedding fabric, distinct, wet.			
	10		Rock. SANDSTONE: slightly weathered, medium strength, pale grey and orange, fine grained, distinct, moist.			
	11		BH102/RMW02 Terminated at 10.2m (Target Depth)			
	12					
	13					



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Boring No.: BH103/RMW03

Easting	: 0.00	Drill Supplier	: Stratacore Drilling Pty Ltd	Job Number	: 22148
Northing	: 0.00	Driller Company	: Stratacore Drilling Pty Ltd	Client	: Billbergia Pty Ltd
Elevation	: Not Surveyed	Logged By	: Renee Ashton	Project	: Detailed Site Investigation
Total Depth	: 10 m	Date	: 22/05/2024	Location	: 25-27 Leeds Street, Rhodes NSW

Sheet : 1 OF 2



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**Boring No.: BH103/RMW03**

Easting : 0.00      Drill Supplier : Stratacore Drilling Pty Ltd      Job Number : 22148 Northing : 0.00      Driller Company : Stratacore Drilling Pty Ltd      Client : Bilbergia Pty Ltd Elevation : Not Surveyed      Logged By : Renee Ashton      Project : Detailed Site Investigation Total Depth : 10 m      Date : 22/05/2024      Location : 25-27 Leeds Street, Rhodes NSW							Sheet : 2 OF 2			
Drilling Method	Depth (m)	Samples		Soil Origin	Graphic Log	Moisture	Material Description	Comments	Well Diagram	Water
		Discrete	PID (ppm)							
	7.0		0.2				Rock, SANDSTONE: highly weathered, very low strength, pale grey, medium to coarse grained, indistinct, slightly moist.			
	8									
	9									
	10						BH103/RMW03 Terminated at 10m (Target Depth)			
	11									
	12									
	13									



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Phone: (02) 9521 6567

Boring No.: BH104/RMW04

Easting	: 0.00	Drill Supplier	: Stratacore Drilling Pty Ltd	Job Number	: 22148
Northing	: 0.00	Driller Company	: Stratacore Drilling Pty Ltd	Client	: Billbergia Pty Ltd
Elevation	: Not Surveyed	Logged By	: Renee Ashton	Project	: Detailed Site Investigation
Total Depth	: 7 m	Date	: 22/05/2024	Location	: 25-27 Leeds Street, Rhodes NSW

Sheet : 1 OF 1

Drilling Method	Depth (m)	Samples	PID (ppm)	Soil Origin	Graphic Log	Moisture	Material Description	Comments	Well Diagram	Water
				Non-Soil			Concrete			
1				Fill		SL M	Fill. Silty SAND SM: loose, dark brown, coarse grained, trace coarse sized gravel, slightly moist. Fill. Silty SAND SM: loose, brown, medium to coarse grained, with medium to coarse sized gravel, trace low plasticity clay, slightly moist.			-Backfill
2				Natural		M	Natural. Sandy CLAY CL: firm, low plasticity, red pale grey, medium grained sand, inorganic, moist.			50mm PVC Solid
3						W	Natural. Sandy CLAY ML: soft, non-plastic, dark grey, medium grained sand, organic, wet, carbonate materials (shells) present..	Carbonate materials (shells) present.		-Bentonite
4						M	Natural. Sandy CLAY CI-CH: firm to stiff, medium to high plasticity, red grey mottled, fine grained sand, organic, moist, carbonate materials (shells) present..	Carbonate materials (shells) present.		-Washed 1-2mm graded sand
5							As above, but CL-CI: stiff to very stiff, low to medium plasticity.			50mm PVC Slotted
6							Natural. Clayey SAND SC: medium dense, low plasticity clay, red white pale grey mottled, medium grained, moist.			
							BH104/RMW04 refusal at 7m (Refusal on rock at 7.0m)			

**Reditus Consulting Pty Ltd**

Lvl 1, Suite 1/29-33 Waratah St, Kirrawee NSW 2232, Australia

Phone: (02) 9521 6567

**Boring No.: BH115/RMW05**

Easting	: 0.00	Drill Supplier	: Stratacore Drilling Pty Ltd	Job Number	: 22148	Sheet	: 1 OF 2
Northing	: 0.00	Driller Company	: Stratacore Drilling Pty Ltd	Client	: Bilbergia Pty Ltd		
Elevation	: Not Surveyed	Logged By	: Renee Ashton	Project	: Detailed Site Investigation		
Total Depth	: 12.1 m	Date	: 23/05/2024	Location	: 25-27 Leeds Street, Rhodes NSW		
Drilling Method	Depth (m)	Samples		Material Description	Comments	Well Diagram	Water
		Discrete	PbD (ppm)	Soil Origin	Graphic Log	Moisture	
				Non-Soil			Concrete
		0.2	0.5	Fill		SL M	Fill. Silty SAND SM: loose, dark brown, medium to coarse grained, with medium to coarse sized gravel, slightly moist.
		0.5	0.3				Fill. Silty SAND SM: loose, red brown, medium grained, slightly moist, potentially reworked natural.
	1	1.0	0.6	Rock			Rock. SANDSTONE: moderately weathered, low strength, red brown, fine grained, indistinct.
	2	2.0	0.6				As above, but medium grained, pale brown grain inclusions.
	3	3.0	0.8				As above, but highly weathered, red pale grey mottled.
	4	4.0	0.3				Rock. SANDSTONE: highly weathered, low strength, grey light brown, fine grained, indistinct.
	5	5.1					Rock. SANDSTONE: moderately weathered, medium strength, red orange brown, fine grained, bedding fabric, distinct, sub-vertical fracture, cross-bedded, iron staining. Rock. SANDSTONE: slightly weathered, high strength, orange brown pale grey, fine grained, bedding fabric, distinct, cross-bedded, iron staining, evidence of clay bands, some sub-horizontal fractures, but mostly competent rock. heavy iron staining and clay bands at 4.9 m, followed by pale grey orange fine grained sandstone, cross-bedded with some sub-horizontal fractures, competent rock except for fractures and clay layers at 5.7 m & 5.9 m..
	6						Rock. SANDSTONE: fresh weathered, high strength, grey, fine grained, bedding fabric, distinct, competent, minor sub-horizontal fractures.



REDITUS

**Reditus Consulting Pty Ltd**

Lvl 1, Suite 1/29-33 Waratah St, Kirrawee NSW 2232, Australia  
Phone: (02) 9521 6567

Boring No.: BH115/RMW05

Easting : 0.00 Drill Supplier : Stratacore Drilling Pty Ltd Job Number : 22148  
Northing : 0.00 Driller Company : Stratacore Drilling Pty Ltd Client : Billbergia Pty Ltd  
Elevation : Not Surveyed Logged By : Renee Ashton Project : Detailed Site Investigation  
Total Depth : 12.1 m Date : 23/05/2024 Location : 25-27 Leeds Street, Rhodes NSW

Sheet : 2 OF 2

E

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## Field Sheets

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# RREDITUS Groundwater Monitoring Field Sheet

Job Information	
Date: 30/5/24	Time: arrive 9:00 depart
Project Name: Groundwater Monitoring Event	Project Number: 22148
Site Location: 25-27 Leeds St, Rhodes NSW	Operator: KS
Well ID: RMW01	Weather: Sunny, still.

Equipment	
Water quality equipment description: HANNA YSI Pro	
Interface probe number: Heron H.Oil	
Purging equipment: (please circle)	Bailer type: Plastic Teflon
	Pump type: Peristaltic Submersible Micro-purge Amazon Other:

Well Gauging and Purge Volume Calculations								
Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm
Conversion Factor (volume in factor L/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3
Total Well Depth (-) Water level (=) Water Column								
6.80 m (-) 2.245 m (=) 4.56 m								
Water Column (x) Conversion Factor (=) Litres per 1 Well Volume								
4.56 m (x) 1.96 (=) 8.93 L								
<b>Volume of water in well / V = Pr x r x h</b> V = volume in litres P = 3.14159 r = radius in cm h = height of water column in cm								

Water Quality Parameters								
Beginning purge time: 10:26							Ending purge time:	
Litres	Time	pH	Temp C°	Cond mS/cm	DO ppm	Redox mV	Depth to water mbtoc	Comments
1	10:30	6.23	20.9	18.27	2.61	-174.8	2.30	turbid orange, clearing up
3	10:44	5.88	20.8	18.14	3.37	-18.9	2.31	clear, organic odour.
5	10:59	4.99	21.1	17.17	3.78	-16.6	2.34	"
6	11:05	4.92	21.0	17.70	3.75	8.2	2.34	"
8	11:10	4.79	21.1	17.59	3.73	-46.2	2.36	"
10	11:20	4.76	21.1	17.51	3.75	-149.4	2.37	"
11	11:25	4.74	21.1	17.51	3.71	-128.3	2.38	"
12	11:30	4.73	21.1	17.58	3.78	-123.5	2.38	"
								sampled
<b>Stabilisation Criteria</b>	+/- 0.05	+/- 10%	+/- 3%	+/- 10%	+/- 10%			<b>Example Comments:</b> clear / slightly cloudy / turbid / very turbid / no odour / slight odour / odour / strong odour
	Total Well Volume	*pH, temp, cond readings not necessary if well is purged dry & ready						
	Actual amount of water prior to sampling							
		Did field parameters stabilise? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA						
		Was the well dry purged? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA						

Field QC Checks								
Was pre-cleaning sampling equipment used for these samples?	<input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA							
Was pre-cleaning sampling equipment properly protected from contamination?	<input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA							
Was documentation of equipment conducted?	<input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA							
Were air bubbles present in vials at time of collection?	<input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA							
Was sample for metals field filtered prior to preservations?	<input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA							
Duplicate sample collected?	<input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA							
	Sample containers:							
	Duplicate sample ID							



# **REDITUS** Groundwater Monitoring Field Sheet

Job Information			
Date: <u>30/5/24</u>	Time: arrive <u>9:00</u>	depart	
Project Name: Groundwater Monitoring Event	Project Number:	22148	
Site Location: 25-27 Leeds St, Rhodes NSW	Operator:	KS	
Well ID: <del>RW02</del> RMW02	Weather:	Sunny / still	

## Equipment

Water quality equipment description: ~~HANNA~~ YSI Pro

Interface probe number: Heron H.Oil

Purging equipment:  
(please circle) Bailer type: Plastic Teflon  
Pump type: Peristaltic Submerge

## Well Gauging and Purge Volume Calculations

Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm	Volume of water in well / V = $\pi r^2 h$
Conversion Factor (volume in factor l/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3	V = volume in litres $R = 3.14159$

Total Well Depth (-) Water level (=) Water Column

Total Well Depth (-) Water level (-) Water Column  
10.00 m (-) 2.38 m (=) 7.62 m

**Volume of water in well / V**  
**= Pr<sup>2</sup> x r x h**  
**V = volume in litres**  
**P = 3.14159**  
**r = radius in cm**  
**h = height of water column in cm**

Water Column (x) Conversion Factor (=) Litres per 1 Well Volume  
7.62 m (x) 1.96 (=) 14.94 L

## Water Quality Parameters

## Field QC Checks

Was pre-cleaning sampling equipment used for these samples?

Was pre-cleaning sampling equipment properly protected from contamination?

Was documentation of equipment conducted?

Were air bubbles present in vials at time of collection?

Was a sample from each field filtered prior to preservation?

Do I have enough samples collected?

obilise?  Y  N  NA

\*pH, temp, cond readings not necessary if well is purged dry

### Did field parameters stabilise?

Was the well dry purged?

N

#### Sample containers:

Y	N	NA

Duplicate sample ID

# RREDITUS Groundwater Monitoring Field Sheet

Job Information	
Date: 30/6/24	Time: arrive 9:00 depart
Project Name: Groundwater Monitoring Event	Project Number: 22148
Site Location: 25-27 Leeds St, Rhodes NSW	Operator: KS
Well ID: R MW03	Weather: Partly cloudy

Equipment	
Water quality equipment description: HANNA YSI Pro	
Interface probe number: Heron H.Oil	
Purging equipment: Bailer type: Plastic	Teflon
(please circle)	
Pump type: Peristaltic	Submersible Micro-purge Amazon Other:

Well Gauging and Purge Volume Calculations									
Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm	
Conversion Factor (volume in factor L/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3	
Total Well Depth (-) Water level (=) Water Column	4.85 m	(-) 2.73 m	(=) _____ m	$V = \pi r^2 h$ $V = \text{volume in litres}$ $P = 3.14159$ $r = \text{radius in cm}$ $h = \text{height of water column in cm}$					
Water Column (x) Conversion Factor (=) Litres per 1 Well Volume	_____ m	(x) _____	(=) _____ L						

Water Quality Parameters								
Beginning purge time: 2:17							Ending purge time:	
Litres	Time	pH	Temp C°	Cond mS/cm	DO ppm	Redox mV	Depth to water mbtoc	Comments
1	2:21	4.28	21.7	23.88	4.08	152.1	3.11	clear, purge slowed.
2	2:36	4.31	21.7	22.23	6.09	205.7	4.04	purge slowed further.
5	2:41	4.36	21.7	21.92	5.83	209.3	4.31	clear
7	2:51	4.36	21.6	21.82	5.68	211.7	4.67	"
8	2:56	4.35	21.6	21.82	5.73	212.1	4.69	"
								Sampled
Stabilisation Criteria								Example Comments: clear / slightly cloudy / turbid / very turbid / no odour / slight odour / odour / strong odour
Total Well Volume Actual amount of water prior to sampling								*pH, temp, cond readings not necessary if well is purged dry
Did field parameters stabilise? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA								Was the well dry purged? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA

Field QC Checks							
Was pre-cleaning sampling equipment used for these samples?	<input checked="" type="checkbox"/> Y	N	NA	Sample containers:			
Was pre-cleaning sampling equipment properly protected from contamination?	<input checked="" type="checkbox"/> Y	N	NA				
Was documentation of equipment conducted?	<input checked="" type="checkbox"/> Y	N	NA				
Were air bubbles present in vials at time of collection?	<input checked="" type="checkbox"/> Y	<input checked="" type="checkbox"/> N	NA				
Was sample for metals field filtered prior to preservations?	<input checked="" type="checkbox"/> Y	N	NA				
Duplicate sample collected?	<input checked="" type="checkbox"/> Y	<input checked="" type="checkbox"/> N		Duplicate sample ID			



# REDITUS Groundwater Monitoring Field Sheet

Job Information		
Date: 30/5/24	Time: arrive	depart
Project Name: Groundwater Monitoring Event		Project Number: 22148
Site Location: 25-27 Leeds St, Rhodes NSW		Operator: KS
Well ID: RMW04	Weather: Sunny, still	

Equipment								
Water quality equipment description: HANNA YS1 Pro								
Interface probe number: Heron H.Oil								
Purging equipment: (please circle)	Bailer type:	Plastic	Teflon					
	Pump type:	Peristaltic		Submersible	Micro-purge	Amazon		Other:

Well Gauging and Purge Volume Calculations								
Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm
Conversion Factor (volume in factor L/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3
Total Well Depth (-) Water level (=) Water Column								
6.78 m	(-) 3.34 m	=						
Water Column (x) Conversion Factor (=) Litres per 1 Well Volume								
	m (x)			(=)		L		

Water Quality Parameters								
Beginning purge time: 12:26							Ending purge time:	
Litres	Time	pH	Temp C°	Cond mS/cm	DO ppm	Redox mV	Depth to water mbtoc	Comments
1	12:36	5.36	21.8	13.20	3.85	-0.6	3.80	clear
2	12:42	5.33	21.8	13.06	4.22	14.0	4.00	clear, organic odour
3	12:46	5.34	21.9	12.84	6.56	23.2	4.15	"
6	12:58	5.70	22.0	12.47	1.60	35.2	4.50	pump slowed
8	1:05	5.68	22.0	12.51	107.85	39.2	4.70	Air bubbles in line impacting DO, sufficient column available for peristaltic pump.
10	1:14	5.67	21.3	12.52	5.04	42.8	4.90	column available for peristaltic pump.
11	1:19	5.63	21.7	12.44	4.98	45.0	5.00	Likely due to subsurface conditions.
12	1:24	5.61	21.9	12.44	1.91	47.0	5.10	clear, organic odour.
13	1:28	5.58	22.1	12.53	5.02	48.3	5.15	"
								suspected
Stabilisation Criteria	+/- 0.05	+/- 10%	+/- 3%	+/- 10%	+/- 10%			Example Comments: clear / slightly cloudy / turbid / very turbid / no odour / slight odour / odour / strong odour
15	Total Well Volume	pH, temp, cond readings not necessary if well is purged dry						
	Actual amount of water prior to sampling	Did field parameters stabilise? <input checked="" type="checkbox"/> N NA						
		Was the well dry purged? <input checked="" type="checkbox"/> N NA						

Field QC Checks								
Was pre-cleaning sampling equipment used for these samples?						<input checked="" type="checkbox"/> Y	N	NA
Was pre-cleaning sampling equipment properly protected from contamination?						<input checked="" type="checkbox"/> Y	N	NA
Was documentation of equipment conducted?						<input checked="" type="checkbox"/> Q	N	NA
Were air bubbles present in vials at time of collection?						<input checked="" type="checkbox"/> Y	<input checked="" type="checkbox"/> N	NA
Was sample for metals field filtered prior to preservations?						<input checked="" type="checkbox"/> Y	N	NA
Duplicate sample collected?						<input checked="" type="checkbox"/> Y	<input checked="" type="checkbox"/> N	Duplicate sample ID

# RREDITUS Groundwater Monitoring Field Sheet

Job Information	
Date: 31/5/24	Time: arrive 7:30 depart
Project Name: Groundwater Monitoring Event	Project Number: 22148
Site Location: 25-27 Leeds St, Rhodes NSW	Operator: KS
Well ID: RMW05	Weather: cloudy, gusts of wind.

Equipment	
Water quality equipment description: HANNA VS 1 Pro	
Interface probe number: Heron H.Oil	
Purging equipment: (please circle)	Bailer type: Plastic Teflon
	Pump type: Peristaltic Submersible Micro-purge Amazon Other:

Well Gauging and Purge Volume Calculations								
Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm
Conversion Factor (volume in factor L/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3
Total Well Depth (-) Water level (=) Water Column								
<u>12.00</u> m (-) <u>4.92</u> m (=) _____ m								
Water Column (x) Conversion Factor (=) Litres per 1 Well Volume								
<u>7.08</u> m (x) _____ (=) _____ L								
Volume of water in well / V = $\pi r^2 h$								
V = volume in litres								
P = 3.14159								
r = radius in cm								
h = height of water column in cm								

Water Quality Parameters								
Beginning purge time: 3:02					Ending purge time:			
Litres	Time	pH	Temp C°	Cond mS/cm	DO ppm	Redox mV	Depth to water mbtoc	Comments
1	3:07	5.76	21.4	3.956	2.03	7.14	5.03	clear.
2	3:17	5.72	21.4	4.251	2.62	45.9	5.10	"
3	3:27	5.72	21.4	4.510	3.47	49.3	5.13	"
4	3:37	5.68	21.3	4.689	4.05	51.1	5.13	"
5	3:47	5.67	21.3	4.739	4.36	52.3	5.14	"
6	3:57	5.66	21.2	4.783	4.07	53.5	5.14	"
7	4:07	5.64	21.2	4.794	4.95	54.3	5.14	"
8	4:07	5.63	21.1	4.807	4.38	54.9	5.14	"
9	4:17	5.62	21.2	4.816	4.55	55.9	5.14	"
								Sampled
Stabilisation Criteria		+/- 0.05	+/- 10%	+/- 3%	+/- 10%	+/- 10%		Example Comments: clear / slightly cloudy / turbid / very turbid / no odour / slight odour / odour / strong odour
Total Well Volume		Actual amount of water prior to sampling						*pH, temp, cond readings not necessary if well is purged dry
								Did field parameters stabilise? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA
								Was the well dry purged? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA

Field QC Checks							
Was pre-cleaning sampling equipment used for these samples?					<input checked="" type="checkbox"/> Y	N	NA
Was pre-cleaning sampling equipment properly protected from contamination?					<input checked="" type="checkbox"/> Y	N	NA
Was documentation of equipment conducted?					<input checked="" type="checkbox"/> Y	N	NA
Were air bubbles present in vials at time of collection?					<input type="checkbox"/> Y	<input checked="" type="checkbox"/> N	NA
Was sample for metals field filtered prior to preservations?					<input checked="" type="checkbox"/> Y	N	NA
Duplicate sample collected?					<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N	Duplicate sample ID



**REDITUS** Groundwater Monitoring Field Sheet

Job Information			
Date: <u>31/5/24</u>	Time: arrive	7:30	depart
Project Name: Groundwater Monitoring Event	Project Number:	22148	
Site Location: 25-27 Leeds St, Rhodes NSW	Operator:	KS	
Well ID: FW1	Weather:	Cloudy	

Equipment							
Water quality equipment description: <b>HANNA</b>							
Interface probe number: <b>Heron H.Oil</b>							
Purging equipment: (please circle)	Bailer type:	Plastic	Teflon				
	Pump type:	Peristaltic	Submersible	Micro-purge	Amazon	Other:	

Well Gauging and Purge Volume Calculations									Volume of water in well / V = $\pi r^2 h$
Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm	V = volume in litres $P = 3.14159$ $r = \text{radius in cm}$ $h = \text{height of water column in cm}$
Conversion Factor (volume in factor L/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3	V = volume in litres $P = 3.14159$ $r = \text{radius in cm}$ $h = \text{height of water column in cm}$
Total Well Depth (-) Water level (=) Water Column									
8.50	m	(-)	3.73	m	(=)	4.77	m		
Water Column (x) Conversion Factor (=) Litres per 1 Well Volume									
4.77	m	(x)	1.96		(=)	9.35	L		

Field QC Checks			
Was pre-cleaning sampling equipment used for these samples?	<input checked="" type="radio"/>	N	NA
Was pre-cleaning sampling equipment properly protected from contamination?	<input checked="" type="radio"/>	N	NA
Was documentation of equipment conducted?	<input checked="" type="radio"/>	N	NA
Were air bubbles present in vials at time of collection?	<input checked="" type="radio"/>	N	NA
Was sample for metals field filtered prior to preservations?	<input checked="" type="radio"/>	N	NA
Duplicate sample collected?	<input checked="" type="radio"/>	N	
Sample containers:			DUP2/TRIP2 PFAS ONLY

# RREDITUS Groundwater Monitoring Field Sheet

Job Information		
Date: 31/5/24	Time: arrive	depart
Project Name: Groundwater Monitoring Event	Project Number: 22148	
Site Location: 25-27 Leeds St, Rhodes NSW	Operator: KS	
Well ID: MW1-B	Weather: Cloudy	

Equipment		
Water quality equipment description: HANNA YSI Pro		
Interface probe number: Heron H.Oil		
Purging equipment: (please circle)	Bailer type: Plastic	Teflon
	Pump type: Peristaltic	Submersible Micro-purge Amazon Other:

Well Gauging and Purge Volume Calculations									
Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm	Volume of water in well / V = $\pi r^2 h$
Conversion Factor (volume in factor L/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3	V = volume in litres P = 3.14159 r = radius in cm h = height of water column in cm
Total Well Depth (-) Water level (=) Water Column	6.15 m	(-) 1.84 m	(=) 4.31 m						
	Water Column (x) Conversion Factor (=) Litres per 1 Well Volume	4.31 m (x) 1.96	(=) 8.45 L						

Water Quality Parameters									
Beginning purge time: 9:54					Ending purge time:				
Litres	Time	pH	Temp C°	Cond mS/cm	DO ppm	Redox mV	Depth to water mbtoc	Comments	
1	9:59	6.16	21.8	7.35	0.68	-13.0	1.85	yellow, organic odour	
3	10:09	6.20	22.2	7.51	2.12	16.3	1.95	"	
5	10:19	6.23	22.3	7.62	2.95	-35.9	1.95	"	
7	10:29	6.25	22.2	7.70	3.19	-13.6	2.40	purged slowed	
8	10:34	6.26	22.2	7.71	3.26	-130.9	2.65	yellow, organic odour	
9	10:39	6.26	22.2	7.71	3.29	-125.6	2.20	"	
10	10:44	6.26	22.2	7.73	3.30	-118.4	2.90	"	
								sampled	
<b>Stabilisation Criteria</b>	+/- 0.05	+/- 10%	+/- 3%	+/- 10%	+/- 10%			<b>Example Comments:</b> clear / slightly cloudy / turbid / very turbid / no odour / slight odour / odour / strong odour	
11	<b>Total Well Volume</b> Actual amount of water prior to sampling						*pH, temp, cond readings not necessary if well is purged dry		
							Did field parameters stabilise? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA	Was the well dry purged? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA	

Field QC Checks							
Was pre-cleaning sampling equipment used for these samples?	<input checked="" type="checkbox"/> Y	N	NA	Sample containers:			
Was pre-cleaning sampling equipment properly protected from contamination?	<input checked="" type="checkbox"/> Y	N	NA				
Was documentation of equipment conducted?	<input checked="" type="checkbox"/> Y	N	NA				
Were air bubbles present in vials at time of collection?	<input checked="" type="checkbox"/> Y	<input checked="" type="checkbox"/> N	NA				
Was sample for metals field filtered prior to preservations?	<input checked="" type="checkbox"/> Y	N	NA				
Duplicate sample collected?	<input checked="" type="checkbox"/> Y	N		Duplicate sample ID DUPI / TRIP1			

# R EDITUS Groundwater Monitoring Field Sheet

Job Information		
Date: 31/5/24	Time: arrive	depart
Project Name: Groundwater Monitoring Event	Project Number: 22148	
Site Location: 25-27 Leeds St, Rhodes NSW	Operator: KS	
Well ID: MW2 - 13	Weather: Cloudy	

Equipment		
Water quality equipment description: HANNA VSI Pro		
Interface probe number: Heron H.Oil		
Purging equipment: (please circle)	Bailer type: Plastic	Teflon
	Pump type: Peristaltic	Submersible Micro-purge Amazon Other:

Well Gauging and Purge Volume Calculations									
Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm	Volume of water in well / V = $\pi r^2 h$
Conversion Factor (volume in factor L/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3	V = volume in litres $P = 3.14159$ $r = \text{radius in cm}$ $h = \text{height of water column in cm}$
Total Well Depth (-) Water level (=) Water Column	11.00	1.955	9.045						
Water Column (x) Conversion Factor (=) Litres per 1 Well Volume	9.045	1.96	(=) 17.73	L					

Water Quality Parameters								
Beginning purge time: 11:34						Ending purge time:		
Litres	Time	pH	Temp C°	Cond mS/cm	DO ppm	Redox mV	Depth to water mbtoc	Comments
2	11:46	5.93	20.7	12.11	0.00	-123.7	2.60	Well inspected w/ endoscope, SCERN
3	11:51	5.94	20.7	12.07	0.00	-126.0	2.64	from 2m bgl. H <sub>2</sub> S odour
5	12:01	5.97	20.8	11.97	4.37	-139.4	3.05	on dipper, opaque, grey precipitate.
6	12:06	6.00	20.8	11.98	4.13	-142.7	3.26	Cleaning w/ purge.
7	12:11	6.02	20.8	11.82	4.13	-144.2	3.37	"
8	12:21	6.06	20.8	11.69	4.03	-148.2	3.48	"
9	12:26	6.16	20.8	11.34	3.92	-150.2	3.60	"
10	12:31	6.22	20.8	11.03	3.80	-147.2	3.62	Spot sheen, strong organic / H <sub>2</sub> S odour.
11	12:36	6.24	20.8	10.94	3.72	-145.9	3.70	"
12	12:41	6.25	20.8	10.78	3.65	-147.6	3.70	"
13	12:46	6.25	20.8	10.76	3.64	-147.6	3.70	" Sampled
Stabilisation Criteria	+/- 0.05	+/- 10%	+/- 3%	+/- 10%	+/- 10%			Example Comments: clear / slightly cloudy / turbid / very turbid / no odour / slight odour / odour / strong odour
	Total Well Volume						*pH, temp, cond readings not necessary if well is purged dry	
	Actual amount of water prior to sampling						Did field parameters stabilise? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA	
							Was the well dry purged? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA	

Field QC Checks							
Was pre-cleaning sampling equipment used for these samples?	<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA	Sample containers:			
Was pre-cleaning sampling equipment properly protected from contamination?	<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA				
Was documentation of equipment conducted?	<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA				
Were air bubbles present in vials at time of collection?	<input type="checkbox"/> X	<input type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA			
Was sample for metals field filtered prior to preservations?	<input type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA				
Duplicate sample collected?	<input type="checkbox"/> Y	<input checked="" type="checkbox"/> N	<input type="checkbox"/> NA	Duplicate sample ID			



# REDITUS Groundwater Monitoring Field Sheet

Job Information		
Date: 30/5/24	Time: arrive 9:00	depart
Project Name: Groundwater Monitoring Event	Project Number: 22148	
Site Location: 25-27 Leeds St, Rhodes NSW	Operator: KS	
Well ID: MW 3-13	Weather: P. Cloudy	

Equipment		
Water quality equipment description: HANNA YSI Pro		
Interface probe number: Heron H.Oil		
Purging equipment: (please circle)	Bailer type: Plastic	Teflon
	Pump type: Peristaltic	Submersible Micro-purge Amazon Other:

Well Gauging and Purge Volume Calculations									
Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm	Volume of water in well / V = $\pi r^2 h$
Conversion Factor (volume in factor L/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3	V = volume in litres $\pi = 3.14159$ $r = \text{radius in cm}$ $h = \text{height of water column in cm}$
Total Well Depth (-) Water level (=) Water Column	5.93 m	2.153 m	(=)						
	Water Column (x) Conversion Factor (=) Litres per 1 Well Volume _____ m (x) _____ (=) _____ L								

Water Quality Parameters								
Beginning purge time: 3:36							Ending purge time:	
Litres	Time	pH	Temp C°	Cond mS/cm	DO ppm	Redox mV	Depth to water mbtoc	Comments
12	3:46	6.80	21.4	38.72	3.04	-28.3	2.21	clear, organic odour
3	3:51	6.83	21.6	38.84	2.67	50.1	2.30	"
5	4:03	6.73	21.4	38.75	3.28	69.1	2.30	"
6	4:09	6.73	21.3	38.17	3.27	72.6	2.36	"
8	4:18	6.73	21.3	38.14	3.58	84.2	2.33	"
9	4:23	6.73	21.3	38.14	3.42	87.7	2.34	"
10	4:33	6.74	21.3	38.10	3.91	91.6	2.34	sampled
Stabilisation Criteria	+/- 0.05	+/- 10%	+/- 3%	+/- 10%	+/- 10%			Example Comments: clear / slightly cloudy / turbid / very turbid / no odour / slight odour / odour / strong odour
	Total Well Volume Actual amount of water prior to sampling							*pH, temp, cond readings not necessary if well is purged dry
	Did field parameters stabilise? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA							Was the well dry purged? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA

Field QC Checks							
Was pre-cleaning sampling equipment used for these samples?				<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA	Sample containers:
Was pre-cleaning sampling equipment properly protected from contamination?				<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA	
Was documentation of equipment conducted?				<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA	
Were air bubbles present in vials at time of collection?				<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA	
Was sample for metals field filtered prior to preservations?				<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N	<input type="checkbox"/> NA	
Duplicate sample collected?				<input checked="" type="checkbox"/> Y	<input type="checkbox"/> N		Duplicate sample ID



# REDITUS Groundwater Monitoring Field Sheet

Job Information		
Date: <u>3/5/24</u>	Time: arrive <u>7:30</u>	depart _____
Project Name: <b>Groundwater Monitoring Event</b>	Project Number: <b>22148</b>	
Site Location: <b>25-27 Leeds St, Rhodes NSW</b>	Operator: <b>KS</b>	
Well ID: <b>MW4-13</b>	Weather: <b>Cloudy</b>	

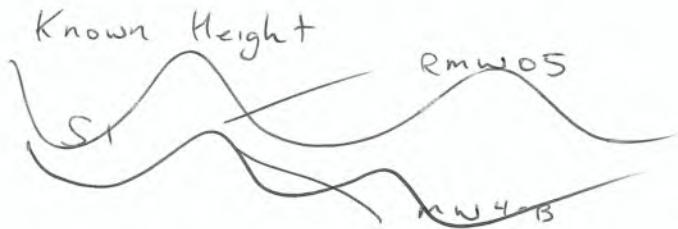
Equipment		
Water quality equipment description: <b>HANNA</b>		
Interface probe number: <b>Heron H.Oil</b>		
Purging equipment: (please circle)	Bailer type: <b>Plastic</b>	Teflon
	Pump type: <b>Peristaltic</b>	Submersible Micro-purge Amazon Other:

Well Gauging and Purge Volume Calculations									
Casing Diameter	25mm	50mm	100mm	125mm	150mm	200mm	250mm	300mm	Volume of water in well / V = $\pi r^2 h$ V = volume in litres $\pi = 3.14159$ r = radius in cm h = height of water column in cm
Conversion Factor (volume in factor L/m)	0.98	1.96	7.85	31.4	49.1	70.7	125.7	196.3	
Total Well Depth (-) Water level (=) Water Column <u>5.70</u> m (-) <u>3.785</u> m (=) <u>1.92</u> m									
Water Column (x) Conversion Factor (=) Litres per 1 Well Volume <u>1.92</u> m (x) <u>1.96</u> (=) <u>3.76</u> L									

Water Quality Parameters									
Beginning purge time: <u>8:12</u>						Ending purge time:			
Litres	Time	pH	Temp C°	Cond mS/cm	DO ppm	Redox mV	Depth to water mbtoc	Comments	
1	8:17	4.67	21.2	0.725	4.09	9.2	3.91	yellow hue, & no turbidity	
2	8:22	4.31	21.3	0.587	4.27	8.14	4.01	Becoming clearer	
8	8:52	4.72	21.1	0.570	4.93	95.7	4.27	<del>yellow</del> yellow tint, opaque.	
9	8:57	4.68	21.4	0.579	4.79	93.5	4.35	"	
10	9:02	4.67	21.4	0.586	2.80	93.0	4.39	"	
11	9:07	4.66	21.4	0.593	2.89	92.8	4.39	"	
12	9:12	4.63	21.4	0.597	3.13	92.7	4.39	"	
13	9:17	4.63	21.4	0.603	3.14	91.7	4.39	Sampled	
<b>Stabilisation Criteria</b>	+/- 0.05	+/- 10%	+/- 3%	+/- 10%	+/- 10%			<b>Example Comments:</b> clear / slightly cloudy / turbid / very turbid / no odour / slight odour / odour / strong odour	
<b>14</b>	<b>Total Well Volume</b> Actual amount of water prior to sampling						*pH, temp, cond readings not necessary if well is purged dry		
Did field parameters stabilise? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA								Was the well dry purged? <input checked="" type="checkbox"/> Y <input type="checkbox"/> N <input type="checkbox"/> NA	

Field QC Checks							
Was pre-cleaning sampling equipment used for these samples?	<input checked="" type="checkbox"/> Y	N	NA	Sample containers:			
Was pre-cleaning sampling equipment properly protected from contamination?	<input checked="" type="checkbox"/> Y	N	NA				
Was documentation of equipment conducted?	<input checked="" type="checkbox"/> Y	N	NA				
Were air bubbles present in vials at time of collection?	<input checked="" type="checkbox"/> Y	<input checked="" type="checkbox"/> N	NA				
Was sample for metals field filtered prior to preservations?	<input checked="" type="checkbox"/> Y	N	NA				
Duplicate sample collected?	<input checked="" type="checkbox"/> Y	<input checked="" type="checkbox"/> N		Duplicate sample ID			

Laser level Survey 22148



Known Height

SI → Laser level back to ↑ → RMW05 ↑ 153.0  
 SI 83.25 cm

7.622 mAHD

MW4-B ↓ 260.5

Additional walls still to survey during slug testing

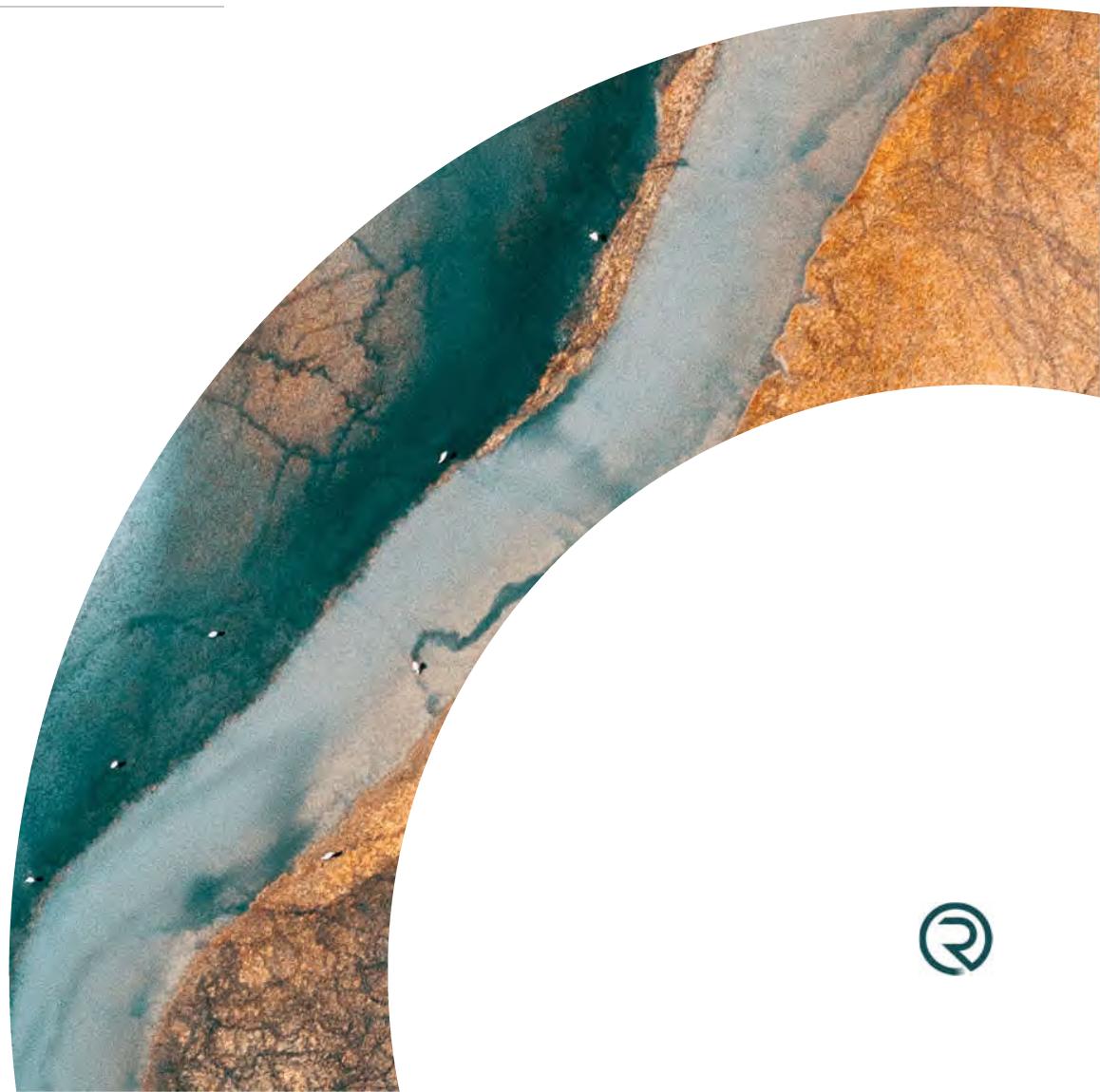
Diver Deploy Log	SWL Time	SWL
RMW01	5:08 pm	2.2 45
RMW02	5:06 pm	2.32
RMW03	5:04 pm	3.84
RMW04	5:02 pm	4.65
RMW05 31/5 4:37 pm	5.04	→ after sampling and develop on same day.

# F

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## Laboratory Reports

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# CHAIN OF CUSTODY FORM - Client

[Copyright and Confidential]

Client: Reditus Consulting	Client Project Name/Number/Site etc (ie report title): Detailed Site Investigation: 25-27 Leeds St, Rhodes NSW
Contact Person: Toby Scrivener	PO No.: 22148
Project Mgr: Toby Scrivener	Envirolab Quoto No.:
Sampler: Renée Ashton	Date results required:
Address: Unit 1A, 29-33 Waratah St, Kirrawee NSW 2232	Or choose: standard / same day / 1 day / 2 day / 3 day Note: Inform lab in advance if urgent turnaround is required - surcharges apply
Phone: (02) 9521 6567	Additional report format: esdat / equis /
Mob: 0477 882 907	Lab Comments:
Email: tobyscrivener@reditus.com.au renaeashon@reditus.com.au denise@reditus.com.au	

## ENVIROLAB GROUP

National phone number 1300 424 344

**Sydney Lab - Envirolab Services**  
12 Ashley St, Chatswood, NSW 2067  
02 9910 6200 | [sydney@envirolab.com.au](mailto:sydney@envirolab.com.au)

**Perth Lab - MPL Laboratories**  
16-18 Haydon Crk, Myaree, WA 6154  
08 9317 2505 | [lab@mpl.com.au](mailto:lab@mpl.com.au)

**Melbourne Lab - Envirolab Services**  
25 Research Drive, Croydon South, VIC 3136  
03 9763 2500 | [melbourne@envirolab.com.au](mailto:melbourne@envirolab.com.au)

**Adelaide Office - Envirolab Services**  
7a The Parade, Norwood, SA 5067  
08 7087 6800 | [adelaide@envirolab.com.au](mailto:adelaide@envirolab.com.au)

**Brisbane Office - Envirolab Services**  
205, 10-20 Depot St, Banyo, QLD 4014  
07 3266 9532 | [brisbane@envirolab.com.au](mailto:brisbane@envirolab.com.au)

**Darwin Office - Envirolab Services**  
Unit 7, 17 Willes Rd, Berrimah, NT 0820

Sample Information					Tests Required								Comments		
Envirolab Sample ID	Client Sample ID or Information	Depth	Data sampled	Type of sample	Combo Ea	Dioxins & Furans	PFAS	Asbestos (%w/w)	Combo 3	TRH, BTEX -PAH, Metals	TRH, STEX	NEPM ELS	Chromium Reducible Sulfur (CrS)	Hold	
1	BH102	0.5	21/05/2024	S	X	X	X	X							
2	BH102	2	21/05/2024	S					X			X			
3	BH102	4	21/05/2024	S									X		
4	BH102	6	21/05/2024	S									X		
5	BH103	1	22/05/2024	S	X	X	X	X							
6	BH103	3	22/05/2024	S					X						
7	BH103	4	22/05/2024	S					X				X		
8	BH103	6	22/05/2024	S									X		
9	BH103	7	22/05/2024	S								X			
10	BH105	0.5	21/05/2024	S	X	X	X	X							
11	BH105	3	21/05/2024	S					X						
12	BH105	4	21/05/2024	S								X			
13	BH106	0.5	22/05/2024	S	X	X	X	X							
14	BH106	2	22/05/2024	S					X		X				
15	BH106	3	22/05/2024	S								X			
16	BH106	6	22/05/2024	S								X			
17	BH107	0.5	21/05/2024	S	X	X	X	X							
18	BH107	3	21/05/2024	S					X						
19	BH107	4	21/05/2024	S								X			
20	BH108	0.3	23/05/2024	S	X										
21	BH109	0.3	20/05/2024	S	X	X	X	X							
22	BH109	1	20/05/2024	S					X						
23	BH109	2	20/05/2024	S								X			
24	BH110	0.25	20/05/2024	S	X		X	X							
25	BH110	1	20/05/2024	S					X						
26	BH111	0.5	23/05/2024	S	X		X	X							
27	BH111	2	23/05/2024	S					X						
28	BH112	0.3	20/05/2024	S	X		X	X							
29	BH112	1	20/05/2024	S					X						
30	BH113	0.5	20/05/2024	S	X			X				X			
31	BH113	2	20/05/2024	S					X						
32	BH114	0.5	20/05/2024	S	X		X	X							
33	BH114	1	20/05/2024	S					X			X			
34	BH115	0.2	23/05/2024	S	X		X	X							
35	BH115	1	23/05/2024	S					X						
36	BH115	5.1	23/05/2024	S								X			
37	BH115	10.7	23/05/2024	S								X			
38	DUP02	2	21/05/2024	S					X						
39	TRIP02	2	21/05/2024	S						X					Pls fwd to ALS
40	DUP03	3	21/05/2024	S					X						Pls fwd to ALS
41	TRIP03	3	21/05/2024	S					X						
42	Trip Blank			S								X			
43	Trip Spike			S								X			

Please tick the box if observed settled sediment present in water samples is to be included in the extraction and/or analysis

Relinquished by (Company): Reditus	Received by (Company): <b>PLS STD</b>	Lab Use Only
Print Name: Renée Ashton	Print Name: <b>Shann Tolc</b>	Cooling: Ice / Ice pack / None
Date & Time: 24/05/2024	Date & Time: <b>24/05/24 1620</b>	Temperature: <b>10</b>
Signature:		Security seal intact? Broken? None
		TAT Req = SAME day / 1 / 2 / 3 / 4 / STD

## CERTIFICATE OF ANALYSIS 352254

### **Client Details**

<b>Client</b>	Reditus Consulting
<b>Attention</b>	Toby Scrivener
<b>Address</b>	Shop 1, 29-33 Waratah St, KIRRRAWEE, NSW, 2232

### **Sample Details**

<b>Your Reference</b>	<u>Detailed Site Investigation:25-27 Leeds St, Rhodes</u>
<b>Number of Samples</b>	41 Soil
<b>Date samples received</b>	24/05/2024
<b>Date completed instructions received</b>	28/05/2024

### **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.  
Samples were analysed as received from the client. Results relate specifically to the samples as received.  
Results are reported on a dry weight basis for solids and on an as received basis for other matrices.  
**Please refer to the last page of this report for any comments relating to the results.**

### **Report Details**

<b>Date results requested by</b>	13/06/2024
<b>Date of Issue</b>	13/06/2024
NATA Accreditation Number 2901. This document shall not be reproduced except in full.	
Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	

#### **Asbestos Approved By**

Analysed by Asbestos Approved Analyst: Sneha Shakya  
Authorised by Asbestos Approved Signatory: Stuart Chen

#### **Authorised By**

Nancy Zhang, Laboratory Manager

#### **Results Approved By**

Diego Bigolin, Inorganics Supervisor  
Dragana Tomas, Senior Chemist  
Giovanni Agosti, Group Technical Manager  
Jenny He, Senior Chemist  
Liam Timmins, Organics Supervisor  
Lucy Zhu, Asbestos Supervisor  
Sean McAlary, Chemist (FAS)  
Stuart Chen, Asbestos Approved Identifier/Report coordinator  
Timothy Toll, Senior Chemist

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

vTRH(C6-C10)/BTEXN in Soil						
Our Reference	UNITS	352254-1	352254-2	352254-5	352254-6	352254-7
Your Reference		BH102	BH102	BH103	BH103	BH103
Depth		0.5	2	1	3	4
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	22/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	<25	<25	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	<25	<25	<25
vTRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	<25	<25	<25
Benzene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Toluene	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Ethylbenzene	mg/kg	<1	<1	<1	<1	<1
m+p-xylene	mg/kg	<2	<2	<2	<2	<2
o-Xylene	mg/kg	<1	<1	<1	<1	<1
Naphthalene	mg/kg	<1	<1	<1	<1	<1
Total +ve Xylenes	mg/kg	<1	<1	<1	<1	<1
Surrogate aaa-Trifluorotoluene	%	88	85	85	103	83

vTRH(C6-C10)/BTEXN in Soil						
Our Reference	UNITS	352254-10	352254-11	352254-13	352254-14	352254-17
Your Reference		BH105	BH105	BH106	BH106	BH107
Depth		0.5	3	0.5	2	0.5
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	<25	<25	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	<25	<25	<25
vTRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	<25	<25	<25
Benzene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Toluene	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Ethylbenzene	mg/kg	<1	<1	<1	<1	<1
m+p-xylene	mg/kg	<2	<2	<2	<2	<2
o-Xylene	mg/kg	<1	<1	<1	<1	<1
Naphthalene	mg/kg	<1	<1	<1	<1	<1
Total +ve Xylenes	mg/kg	<1	<1	<1	<1	<1
Surrogate aaa-Trifluorotoluene	%	75	77	109	81	81

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

vTRH(C6-C10)/BTEXN in Soil						
Our Reference	UNITS	352254-18	352254-20	352254-21	352254-22	352254-24
Your Reference		BH107	BH108	BH109	BH109	BH110
Depth		3	0.3	0.3	1	0.25
Date Sampled		21/05/2024	23/05/2024	20/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	<25	<25	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	<25	<25	<25
vTRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	<25	<25	<25
Benzene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Toluene	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Ethylbenzene	mg/kg	<1	<1	<1	<1	<1
m+p-xylene	mg/kg	<2	<2	<2	<2	<2
o-Xylene	mg/kg	<1	<1	<1	<1	<1
Naphthalene	mg/kg	<1	<1	<1	<1	<1
Total +ve Xylenes	mg/kg	<1	<1	<1	<1	<1
Surrogate aaa-Trifluorotoluene	%	85	92	74	82	74

vTRH(C6-C10)/BTEXN in Soil						
Our Reference	UNITS	352254-25	352254-26	352254-27	352254-28	352254-29
Your Reference		BH110	BH111	BH111	BH112	BH112
Depth		1	0.5	2	0.3	1
Date Sampled		20/05/2024	23/05/2024	23/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	<25	<25	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	<25	<25	<25
vTRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	<25	<25	<25
Benzene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Toluene	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Ethylbenzene	mg/kg	<1	<1	<1	<1	<1
m+p-xylene	mg/kg	<2	<2	<2	<2	<2
o-Xylene	mg/kg	<1	<1	<1	<1	<1
Naphthalene	mg/kg	<1	<1	<1	<1	<1
Total +ve Xylenes	mg/kg	<1	<1	<1	<1	<1
Surrogate aaa-Trifluorotoluene	%	85	80	79	73	104

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

vTRH(C6-C10)/BTEXN in Soil						
Our Reference	UNITS	352254-30	352254-31	352254-32	352254-33	352254-34
Your Reference		BH113	BH113	BH114	BH114	BH115
Depth		0.5	2	0.5	1	0.2
Date Sampled		20/05/2024	20/05/2024	20/05/2024	20/05/2024	23/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	<25	<25	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	<25	<25	<25
vTRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	<25	<25	<25
Benzene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Toluene	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Ethylbenzene	mg/kg	<1	<1	<1	<1	<1
m+p-xylene	mg/kg	<2	<2	<2	<2	<2
o-Xylene	mg/kg	<1	<1	<1	<1	<1
Naphthalene	mg/kg	<1	<1	<1	<1	<1
Total +ve Xylenes	mg/kg	<1	<1	<1	<1	<1
Surrogate aaa-Trifluorotoluene	%	70	93	71	68	88

vTRH(C6-C10)/BTEXN in Soil						
Our Reference	UNITS	352254-35	352254-38	352254-39	352254-40	352254-41
Your Reference		BH115	DUP02	DUP03	Trip Blank	Trip Spike
Depth		1	2	3	-	-
Date Sampled		23/05/2024	21/05/2024	21/05/2024	23/05/2024	23/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	<25	<25	[NA]
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	<25	<25	[NA]
vTRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	<25	<25	[NA]
Benzene	mg/kg	<0.2	<0.2	<0.2	<0.2	84%
Toluene	mg/kg	<0.5	<0.5	<0.5	<0.5	87%
Ethylbenzene	mg/kg	<1	<1	<1	<1	88%
m+p-xylene	mg/kg	<2	<2	<2	<2	80%
o-Xylene	mg/kg	<1	<1	<1	<1	88%
Naphthalene	mg/kg	<1	<1	<1	<1	[NA]
Total +ve Xylenes	mg/kg	<1	<1	<1	<1	[NA]
Surrogate aaa-Trifluorotoluene	%	79	91	76	76	89

svTRH (C10-C40) in Soil						
Our Reference	UNITS	352254-1	352254-2	352254-5	352254-6	352254-7
Your Reference		BH102	BH102	BH103	BH103	BH103
Depth		0.5	2	1	3	4
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	22/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	29/05/2024
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	<100	<100	<100	190
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (C10-C36)	mg/kg	<50	<50	<50	<50	190
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> -C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	<100	<100	<100	<100	230
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (>C10-C40)	mg/kg	<50	<50	<50	<50	230
Surrogate o-Terphenyl	%	96	89	90	90	90

svTRH (C10-C40) in Soil						
Our Reference	UNITS	352254-10	352254-11	352254-13	352254-14	352254-17
Your Reference		BH105	BH105	BH106	BH106	BH107
Depth		0.5	3	0.5	2	0.5
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	<100	<100	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (C10-C36)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> -C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	<100	<100	<100	<100	<100
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (>C10-C40)	mg/kg	<50	<50	<50	<50	<50
Surrogate o-Terphenyl	%	89	90	88	89	87

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

svTRH (C10-C40) in Soil						
Our Reference	UNITS	352254-18	352254-20	352254-21	352254-22	352254-24
Your Reference		BH107	BH108	BH109	BH109	BH110
Depth		3	0.3	0.3	1	0.25
Date Sampled		21/05/2024	23/05/2024	20/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	<100	<100	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (C10-C36)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub>	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>16</sub> - C <sub>34</sub>	mg/kg	<100	<100	<100	<100	<100
TRH >C <sub>34</sub> - C <sub>40</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (>C10-C40)	mg/kg	<50	<50	<50	<50	<50
Surrogate o-Terphenyl	%	89	89	87	91	86

svTRH (C10-C40) in Soil						
Our Reference	UNITS	352254-25	352254-26	352254-27	352254-28	352254-29
Your Reference		BH110	BH111	BH111	BH112	BH112
Depth		1	0.5	2	0.3	1
Date Sampled		20/05/2024	23/05/2024	23/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	<100	<100	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (C10-C36)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub>	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>16</sub> - C <sub>34</sub>	mg/kg	<100	<100	<100	<100	<100
TRH >C <sub>34</sub> - C <sub>40</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (>C10-C40)	mg/kg	<50	<50	<50	<50	<50
Surrogate o-Terphenyl	%	86	87	87	87	86

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

svTRH (C10-C40) in Soil						
Our Reference	UNITS	352254-30	352254-31	352254-32	352254-33	352254-34
Your Reference		BH113	BH113	BH114	BH114	BH115
Depth		0.5	2	0.5	1	0.2
Date Sampled		20/05/2024	20/05/2024	20/05/2024	20/05/2024	23/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	<100	<100	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (C10-C36)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub>	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>16</sub> - C <sub>34</sub>	mg/kg	<100	<100	<100	<100	<100
TRH >C <sub>34</sub> - C <sub>40</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (>C10-C40)	mg/kg	<50	<50	<50	<50	<50
Surrogate o-Terphenyl	%	83	85	86	85	85

svTRH (C10-C40) in Soil				
Our Reference	UNITS	352254-35	352254-38	352254-39
Your Reference		BH115	DUP02	DUP03
Depth		1	2	3
Date Sampled		23/05/2024	21/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100	<100	<100
Total +ve TRH (C10-C36)	mg/kg	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub>	mg/kg	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	<50	<50
TRH >C <sub>16</sub> - C <sub>34</sub>	mg/kg	<100	<100	<100
TRH >C <sub>34</sub> - C <sub>40</sub>	mg/kg	<100	<100	<100
Total +ve TRH (>C10-C40)	mg/kg	<50	<50	<50
Surrogate o-Terphenyl	%	84	84	84

PAHs in Soil						
Our Reference	UNITS	352254-1	352254-2	352254-5	352254-6	352254-7
Your Reference		BH102	BH102	BH103	BH103	BH103
Depth		0.5	2	1	3	4
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	22/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	03/06/2024	28/05/2024	03/06/2024	03/06/2024
Naphthalene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluorene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phenanthrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluoranthene	mg/kg	0.1	<0.1	<0.1	<0.1	<0.1
Pyrene	mg/kg	0.1	<0.1	<0.1	<0.1	<0.1
Benzo(a)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chrysene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(b,j+k)fluoranthene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Benzo(a)pyrene	mg/kg	0.06	<0.05	<0.05	<0.05	<0.05
Indeno(1,2,3-c,d)pyrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dibenzo(a,h)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(g,h,i)perylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve PAH's	mg/kg	0.3	<0.05	<0.05	<0.05	<0.05
Benzo(a)pyrene TEQ calc (zero)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(half)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Surrogate p-Terphenyl-d14	%	112	124	103	125	135

PAHs in Soil						
Our Reference	UNITS	352254-10	352254-11	352254-13	352254-14	352254-17
Your Reference		BH105	BH105	BH106	BH106	BH107
Depth		0.5	3	0.5	2	0.5
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	03/06/2024	28/05/2024	03/06/2024	28/05/2024
Naphthalene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluorene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phenanthrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluoranthene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Pyrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(a)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chrysene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(b,j+k)fluoranthene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Benzo(a)pyrene	mg/kg	<0.05	<0.05	<0.05	<0.05	<0.05
Indeno(1,2,3-c,d)pyrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dibenzo(a,h)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(g,h,i)perylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve PAH's	mg/kg	<0.05	<0.05	<0.05	<0.05	<0.05
Benzo(a)pyrene TEQ calc (zero)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(half)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Surrogate p-Terphenyl-d14	%	108	125	105	122	107

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

PAHs in Soil						
Our Reference	UNITS	352254-18	352254-20	352254-21	352254-22	352254-24
Your Reference		BH107	BH108	BH109	BH109	BH110
Depth		3	0.3	0.3	1	0.25
Date Sampled		21/05/2024	23/05/2024	20/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	03/06/2024	28/05/2024	28/05/2024	03/06/2024	28/05/2024
Naphthalene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluorene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phenanthrene	mg/kg	<0.1	0.1	<0.1	<0.1	<0.1
Anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluoranthene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Pyrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(a)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chrysene	mg/kg	<0.1	0.1	<0.1	<0.1	<0.1
Benzo(b,j+k)fluoranthene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Benzo(a)pyrene	mg/kg	<0.05	<0.05	<0.05	<0.05	<0.05
Indeno(1,2,3-c,d)pyrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dibenzo(a,h)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(g,h,i)perylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve PAH's	mg/kg	<0.05	0.2	<0.05	<0.05	<0.05
Benzo(a)pyrene TEQ calc (zero)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(half)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Surrogate p-Terphenyl-d14	%	122	108	105	123	98

PAHs in Soil						
Our Reference	UNITS	352254-25	352254-26	352254-27	352254-28	352254-29
Your Reference		BH110	BH111	BH111	BH112	BH112
Depth		1	0.5	2	0.3	1
Date Sampled		20/05/2024	23/05/2024	23/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	03/06/2024	28/05/2024	03/06/2024	28/05/2024	03/06/2024
Naphthalene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluorene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phenanthrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluoranthene	mg/kg	<0.1	0.2	<0.1	<0.1	<0.1
Pyrene	mg/kg	<0.1	0.1	<0.1	<0.1	<0.1
Benzo(a)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chrysene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(b,j+k)fluoranthene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Benzo(a)pyrene	mg/kg	<0.05	0.07	<0.05	<0.05	<0.05
Indeno(1,2,3-c,d)pyrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dibenzo(a,h)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(g,h,i)perylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve PAH's	mg/kg	<0.05	0.4	<0.05	<0.05	<0.05
Benzo(a)pyrene TEQ calc (zero)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(half)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Surrogate p-Terphenyl-d14	%	118	105	122	102	120

PAHs in Soil						
Our Reference	UNITS	352254-30	352254-31	352254-32	352254-33	352254-34
Your Reference		BH113	BH113	BH114	BH114	BH115
Depth		0.5	2	0.5	1	0.2
Date Sampled		20/05/2024	20/05/2024	20/05/2024	20/05/2024	23/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	03/06/2024	28/05/2024	03/06/2024	28/05/2024
Naphthalene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluorene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phenanthrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluoranthene	mg/kg	0.1	<0.1	<0.1	<0.1	<0.1
Pyrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(a)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chrysene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(b,j+k)fluoranthene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Benzo(a)pyrene	mg/kg	<0.05	<0.05	<0.05	<0.05	<0.05
Indeno(1,2,3-c,d)pyrene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dibenzo(a,h)anthracene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(g,h,i)perylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve PAH's	mg/kg	0.1	<0.05	<0.05	<0.05	<0.05
Benzo(a)pyrene TEQ calc (zero)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(half)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Surrogate p-Terphenyl-d14	%	101	121	100	76	104

PAHs in Soil				
Our Reference	UNITS	352254-35	352254-38	352254-39
Your Reference		BH115	DUP02	DUP03
Depth		1	2	3
Date Sampled		23/05/2024	21/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	03/06/2024	03/06/2024	03/06/2024
Naphthalene	mg/kg	<0.1	<0.1	<0.1
Acenaphthylene	mg/kg	<0.1	<0.1	<0.1
Acenaphthene	mg/kg	<0.1	<0.1	<0.1
Fluorene	mg/kg	<0.1	<0.1	<0.1
Phenanthrene	mg/kg	<0.1	<0.1	<0.1
Anthracene	mg/kg	<0.1	<0.1	<0.1
Fluoranthene	mg/kg	<0.1	<0.1	<0.1
Pyrene	mg/kg	<0.1	<0.1	<0.1
Benzo(a)anthracene	mg/kg	<0.1	<0.1	<0.1
Chrysene	mg/kg	<0.1	<0.1	<0.1
Benzo(b,j+k)fluoranthene	mg/kg	<0.2	<0.2	<0.2
Benzo(a)pyrene	mg/kg	<0.05	<0.05	<0.05
Indeno(1,2,3-c,d)pyrene	mg/kg	<0.1	<0.1	<0.1
Dibenzo(a,h)anthracene	mg/kg	<0.1	<0.1	<0.1
Benzo(g,h,i)perylene	mg/kg	<0.1	<0.1	<0.1
Total +ve PAH's	mg/kg	<0.05	<0.05	<0.05
Benzo(a)pyrene TEQ calc (zero)	mg/kg	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(half)	mg/kg	<0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	<0.5	<0.5	<0.5
Surrogate p-Terphenyl-d14	%	126	122	117

Organochlorine Pesticides in soil						
Our Reference	UNITS	352254-1	352254-5	352254-10	352254-13	352254-17
Your Reference		BH102	BH103	BH105	BH106	BH107
Depth		0.5	1	0.5	0.5	0.5
Date Sampled		21/05/2024	22/05/2024	21/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
alpha-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
HCB	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
beta-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
gamma-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Heptachlor	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
delta-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aldrin	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Heptachlor Epoxide	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
gamma-Chlordane	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
alpha-chlordane	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endosulfan I	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
pp-DDE	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dieldrin	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endrin	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endosulfan II	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
pp-DDD	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endrin Aldehyde	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
pp-DDT	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endosulfan Sulphate	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Methoxychlor	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Mirex	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve DDT+DDD+DDE	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate 4-Chloro-3-NBTF	%	94	96	93	93	92

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

Organochlorine Pesticides in soil						
Our Reference	UNITS	352254-20	352254-21	352254-24	352254-26	352254-28
Your Reference		BH108	BH109	BH110	BH111	BH112
Depth		0.3	0.3	0.25	0.5	0.3
Date Sampled		23/05/2024	20/05/2024	20/05/2024	23/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
alpha-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
HCB	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
beta-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
gamma-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Heptachlor	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
delta-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aldrin	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Heptachlor Epoxide	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
gamma-Chlordane	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
alpha-chlordane	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endosulfan I	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
pp-DDE	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dieldrin	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endrin	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endosulfan II	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
pp-DDD	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endrin Aldehyde	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
pp-DDT	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endosulfan Sulphate	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Methoxychlor	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Mirex	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve DDT+DDD+DDE	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate 4-Chloro-3-NBTF	%	92	91	88	89	92

Organochlorine Pesticides in soil				
Our Reference	UNITS	352254-30	352254-32	352254-34
Your Reference		BH113	BH114	BH115
Depth		0.5	0.5	0.2
Date Sampled		20/05/2024	20/05/2024	23/05/2024
Type of sample		Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024
alpha-BHC	mg/kg	<0.1	<0.1	<0.1
HCB	mg/kg	<0.1	<0.1	<0.1
beta-BHC	mg/kg	<0.1	<0.1	<0.1
gamma-BHC	mg/kg	<0.1	<0.1	<0.1
Heptachlor	mg/kg	<0.1	<0.1	<0.1
delta-BHC	mg/kg	<0.1	<0.1	<0.1
Aldrin	mg/kg	<0.1	<0.1	<0.1
Heptachlor Epoxide	mg/kg	<0.1	<0.1	<0.1
gamma-Chlordane	mg/kg	<0.1	<0.1	<0.1
alpha-chlordane	mg/kg	<0.1	<0.1	<0.1
Endosulfan I	mg/kg	<0.1	<0.1	<0.1
pp-DDE	mg/kg	<0.1	<0.1	<0.1
Dieldrin	mg/kg	<0.1	<0.1	<0.1
Endrin	mg/kg	<0.1	<0.1	<0.1
Endosulfan II	mg/kg	<0.1	<0.1	<0.1
pp-DDD	mg/kg	<0.1	<0.1	<0.1
Endrin Aldehyde	mg/kg	<0.1	<0.1	<0.1
pp-DDT	mg/kg	<0.1	<0.1	<0.1
Endosulfan Sulphate	mg/kg	<0.1	<0.1	<0.1
Methoxychlor	mg/kg	<0.1	<0.1	<0.1
Mirex	mg/kg	<0.1	<0.1	<0.1
Total +ve DDT+DDD+DDE	mg/kg	<0.1	<0.1	<0.1
Surrogate 4-Chloro-3-NBTF	%	89	90	88

Organophosphorus Pesticides in Soil						
Our Reference	UNITS	352254-1	352254-5	352254-10	352254-13	352254-17
Your Reference		BH102	BH103	BH105	BH106	BH107
Depth		0.5	1	0.5	0.5	0.5
Date Sampled		21/05/2024	22/05/2024	21/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Dichlorvos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Mevinphos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phorate	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dimethoate	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Diazinon	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Disulfoton	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chlorpyrifos-methyl	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Parathion-Methyl	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Ronnel	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fenitrothion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Malathion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chlorpyriphos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fenthion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Parathion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Bromophos-ethyl	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Methidathion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fenamiphos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Ethion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phosalone	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Azinphos-methyl (Guthion)	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Coumaphos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate 4-Chloro-3-NBTF	%	94	96	93	93	92

Organophosphorus Pesticides in Soil						
Our Reference	UNITS	352254-20	352254-21	352254-24	352254-26	352254-28
Your Reference		BH108	BH109	BH110	BH111	BH112
Depth		0.3	0.3	0.25	0.5	0.3
Date Sampled		23/05/2024	20/05/2024	20/05/2024	23/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Dichlorvos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Mevinphos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phorate	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dimethoate	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Diazinon	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Disulfoton	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chlorpyrifos-methyl	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Parathion-Methyl	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Ronnel	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fenitrothion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Malathion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chlorpyriphos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fenthion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Parathion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Bromophos-ethyl	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Methidathion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fenamiphos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Ethion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phosalone	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Azinphos-methyl (Guthion)	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Coumaphos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate 4-Chloro-3-NBTF	%	92	91	88	89	92

Organophosphorus Pesticides in Soil				
Our Reference	UNITS	352254-30	352254-32	352254-34
Your Reference		BH113	BH114	BH115
Depth		0.5	0.5	0.2
Date Sampled		20/05/2024	20/05/2024	23/05/2024
Type of sample		Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024
Dichlorvos	mg/kg	<0.1	<0.1	<0.1
Mevinphos	mg/kg	<0.1	<0.1	<0.1
Phorate	mg/kg	<0.1	<0.1	<0.1
Dimethoate	mg/kg	<0.1	<0.1	<0.1
Diazinon	mg/kg	<0.1	<0.1	<0.1
Disulfoton	mg/kg	<0.1	<0.1	<0.1
Chlorpyrifos-methyl	mg/kg	<0.1	<0.1	<0.1
Parathion-Methyl	mg/kg	<0.1	<0.1	<0.1
Ronnel	mg/kg	<0.1	<0.1	<0.1
Fenitrothion	mg/kg	<0.1	<0.1	<0.1
Malathion	mg/kg	<0.1	<0.1	<0.1
Chlorpyriphos	mg/kg	<0.1	<0.1	<0.1
Fenthion	mg/kg	<0.1	<0.1	<0.1
Parathion	mg/kg	<0.1	<0.1	<0.1
Bromophos-ethyl	mg/kg	<0.1	<0.1	<0.1
Methidathion	mg/kg	<0.1	<0.1	<0.1
Fenamiphos	mg/kg	<0.1	<0.1	<0.1
Ethion	mg/kg	<0.1	<0.1	<0.1
Phosalone	mg/kg	<0.1	<0.1	<0.1
Azinphos-methyl (Guthion)	mg/kg	<0.1	<0.1	<0.1
Coumaphos	mg/kg	<0.1	<0.1	<0.1
Surrogate 4-Chloro-3-NBTF	%	89	90	88

PCBs in Soil						
Our Reference	UNITS	352254-1	352254-5	352254-10	352254-13	352254-17
Your Reference		BH102	BH103	BH105	BH106	BH107
Depth		0.5	1	0.5	0.5	0.5
Date Sampled		21/05/2024	22/05/2024	21/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Aroclor 1016	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1221	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1232	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1242	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1248	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1254	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1260	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve PCBs (1016-1260)	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate 2-Fluorobiphenyl	%	105	112	108	109	106

PCBs in Soil						
Our Reference	UNITS	352254-20	352254-21	352254-24	352254-26	352254-28
Your Reference		BH108	BH109	BH110	BH111	BH112
Depth		0.3	0.3	0.25	0.5	0.3
Date Sampled		23/05/2024	20/05/2024	20/05/2024	23/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Aroclor 1016	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1221	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1232	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1242	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1248	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1254	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1260	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve PCBs (1016-1260)	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate 2-Fluorobiphenyl	%	108	107	103	104	107

PCBs in Soil				
Our Reference	UNITS	352254-30	352254-32	352254-34
Your Reference		BH113	BH114	BH115
Depth		0.5	0.5	0.2
Date Sampled		20/05/2024	20/05/2024	23/05/2024
Type of sample		Soil	Soil	Soil
Date extracted	-	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024
Aroclor 1016	mg/kg	<0.1	<0.1	<0.1
Aroclor 1221	mg/kg	<0.1	<0.1	<0.1
Aroclor 1232	mg/kg	<0.1	<0.1	<0.1
Aroclor 1242	mg/kg	<0.1	<0.1	<0.1
Aroclor 1248	mg/kg	<0.1	<0.1	<0.1
Aroclor 1254	mg/kg	<0.1	<0.1	<0.1
Aroclor 1260	mg/kg	<0.1	<0.1	<0.1
Total +ve PCBs (1016-1260)	mg/kg	<0.1	<0.1	<0.1
Surrogate 2-Fluorobiphenyl	%	102	104	102

Acid Extractable metals in soil						
Our Reference	UNITS	352254-1	352254-2	352254-5	352254-6	352254-7
Your Reference		BH102	BH102	BH103	BH103	BH103
Depth		0.5	2	1	3	4
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	22/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Date analysed	-	30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Arsenic	mg/kg	<4	6	6	6	21
Cadmium	mg/kg	<0.4	<0.4	<0.4	<0.4	<0.4
Chromium	mg/kg	8	12	6	16	6
Copper	mg/kg	18	5	21	8	3
Lead	mg/kg	32	13	19	17	6
Mercury	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Nickel	mg/kg	5	<1	2	2	3
Zinc	mg/kg	54	3	23	7	9
Iron	mg/kg	[NA]	27,000	[NA]	[NA]	[NA]

Acid Extractable metals in soil						
Our Reference	UNITS	352254-10	352254-11	352254-13	352254-14	352254-17
Your Reference		BH105	BH105	BH106	BH106	BH107
Depth		0.5	3	0.5	2	0.5
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Date analysed	-	30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Arsenic	mg/kg	5	9	<4	<4	5
Cadmium	mg/kg	<0.4	<0.4	<0.4	<0.4	<0.4
Chromium	mg/kg	13	18	7	9	9
Copper	mg/kg	12	5	16	3	17
Lead	mg/kg	30	12	15	10	19
Mercury	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Nickel	mg/kg	5	1	6	<1	8
Zinc	mg/kg	39	11	30	2	51
Iron	mg/kg	[NA]	[NA]	[NA]	16,000	[NA]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

Acid Extractable metals in soil						
Our Reference	UNITS	352254-18	352254-20	352254-21	352254-22	352254-24
Your Reference		BH107	BH108	BH109	BH109	BH110
Depth		3	0.3	0.3	1	0.25
Date Sampled		21/05/2024	23/05/2024	20/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Date analysed	-	30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Arsenic	mg/kg	<4	5	<4	<4	<4
Cadmium	mg/kg	<0.4	<0.4	<0.4	<0.4	<0.4
Chromium	mg/kg	7	8	9	10	8
Copper	mg/kg	11	41	40	1	13
Lead	mg/kg	24	19	18	10	2
Mercury	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Nickel	mg/kg	3	22	16	2	6
Zinc	mg/kg	45	75	59	13	26
Iron	mg/kg	[NA]	35,000	[NA]	[NA]	[NA]

Acid Extractable metals in soil						
Our Reference	UNITS	352254-25	352254-26	352254-27	352254-28	352254-29
Your Reference		BH110	BH111	BH111	BH112	BH112
Depth		1	0.5	2	0.3	1
Date Sampled		20/05/2024	23/05/2024	23/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Date analysed	-	30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Arsenic	mg/kg	<4	4	<4	<4	<4
Cadmium	mg/kg	<0.4	<0.4	<0.4	<0.4	<0.4
Chromium	mg/kg	36	16	8	9	16
Copper	mg/kg	8	21	1	13	6
Lead	mg/kg	21	31	3	12	8
Mercury	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Nickel	mg/kg	7	15	<1	9	3
Zinc	mg/kg	69	79	<1	51	12

Acid Extractable metals in soil						
Our Reference	UNITS	352254-30	352254-31	352254-32	352254-33	352254-34
Your Reference		BH113	BH113	BH114	BH114	BH115
Depth		0.5	2	0.5	1	0.2
Date Sampled		20/05/2024	20/05/2024	20/05/2024	20/05/2024	23/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Date analysed	-	30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Arsenic	mg/kg	8	9	6	4	12
Cadmium	mg/kg	<0.4	<0.4	<0.4	<0.4	<0.4
Chromium	mg/kg	23	18	23	12	29
Copper	mg/kg	470	34	3	4	13
Lead	mg/kg	220	28	16	17	16
Mercury	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Nickel	mg/kg	12	11	4	<1	4
Zinc	mg/kg	600	91	4	1	25

Acid Extractable metals in soil						
Our Reference	UNITS	352254-35	352254-38	352254-39	352254-42	352254-43
Your Reference		BH115	DUP02	DUP03	BH102 - [TRIPPLICATE]	BH107 - [TRIPPLICATE]
Depth		1	2	3	0.5	0.5
Date Sampled		23/05/2024	21/05/2024	21/05/2024	21/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Date analysed	-	30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Arsenic	mg/kg	<4	5	<4	5	5
Cadmium	mg/kg	<0.4	<0.4	<0.4	<0.4	<0.4
Chromium	mg/kg	7	10	7	9	8
Copper	mg/kg	2	3	13	19	19
Lead	mg/kg	14	11	25	44	18
Mercury	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Nickel	mg/kg	<1	<1	3	6	6
Zinc	mg/kg	<1	3	42	51	33

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

Moisture						
Our Reference		352254-1	352254-2	352254-5	352254-6	352254-7
Your Reference	UNITS	BH102	BH102	BH103	BH103	BH103
Depth		0.5	2	1	3	4
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	22/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Moisture	%	10	10	5.7	10	23

Moisture						
Our Reference		352254-10	352254-11	352254-13	352254-14	352254-17
Your Reference	UNITS	BH105	BH105	BH106	BH106	BH107
Depth		0.5	3	0.5	2	0.5
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Moisture	%	11	15	12	15	8.6

Moisture						
Our Reference		352254-18	352254-20	352254-21	352254-22	352254-24
Your Reference	UNITS	BH107	BH108	BH109	BH109	BH110
Depth		3	0.3	0.3	1	0.25
Date Sampled		21/05/2024	23/05/2024	20/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Moisture	%	16	6.5	17	6.9	12

Moisture						
Our Reference		352254-25	352254-26	352254-27	352254-28	352254-29
Your Reference	UNITS	BH110	BH111	BH111	BH112	BH112
Depth		1	0.5	2	0.3	1
Date Sampled		20/05/2024	23/05/2024	23/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Moisture	%	9.2	11	7.8	15	6.6

<b>Moisture</b>						
Our Reference		352254-30	352254-31	352254-32	352254-33	352254-34
Your Reference	UNITS	BH113	BH113	BH114	BH114	BH115
Depth		0.5	2	0.5	1	0.2
Date Sampled		20/05/2024	20/05/2024	20/05/2024	20/05/2024	23/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Moisture	%	14	6.2	18	15	11

<b>Moisture</b>				
Our Reference		352254-35	352254-38	352254-39
Your Reference	UNITS	BH115	DUP02	DUP03
Depth		1	2	3
Date Sampled		23/05/2024	21/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil
Date prepared	-	28/05/2024	28/05/2024	28/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024
Moisture	%	3.9	12	16

PFAS in Soils Extended						
Our Reference	UNITS	352254-1	352254-5	352254-10	352254-13	352254-17
Your Reference		BH102	BH103	BH105	BH106	BH107
Depth		0.5	1	0.5	0.5	0.5
Date Sampled		21/05/2024	22/05/2024	21/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Perfluorobutanesulfonic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluoropentanesulfonic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorohexanesulfonic acid - PFHxS	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluoroheptanesulfonic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorooctanesulfonic acid PFOS	µg/kg	<0.1	0.2	<0.1	<0.1	<0.1
Perfluorodecanesulfonic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Perfluorobutanoic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Perfluoropentanoic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Perfluorohexanoic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluoroheptanoic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorooctanoic acid PFOA	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorononanoic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorodecanoic acid	µg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Perfluoroundecanoic acid	µg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Perfluorododecanoic acid	µg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Perfluorotridecanoic acid	µg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Perfluorotetradecanoic acid	µg/kg	<5	<5	<5	<5	<5
4:2 FTS	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
6:2 FTS	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
8:2 FTS	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
10:2 FTS	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Perfluorooctane sulfonamide	µg/kg	<1	<1	<1	<1	<1
N-Methyl perfluorooctane sulfonamide	µg/kg	<1	<1	<1	<1	<1
N-Ethyl perfluorooctanesulfon amide	µg/kg	<1	<1	<1	<1	<1
N-Me perfluorooctanesulfonamid oethanol	µg/kg	<1	<1	<1	<1	<1
N-Et perfluorooctanesulfonamid oethanol	µg/kg	<5	<5	<5	<5	<5
MePerfluorooctanesulf- amid oacetic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
EtPerfluorooctanesulf amid oacetic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%	95	95	97	99	98
Surrogate <sup>13</sup> C <sub>2</sub> PFOA	%	97	101	97	97	101
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFBS	%	100	100	88	92	95
Extracted ISTD <sup>18</sup> O <sub>2</sub> PFHxS	%	102	102	87	92	101
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%	99	99	85	89	94

PFAS in Soils Extended						
Our Reference	UNITS	352254-1	352254-5	352254-10	352254-13	352254-17
Your Reference		BH102	BH103	BH105	BH106	BH107
Depth		0.5	1	0.5	0.5	0.5
Date Sampled		21/05/2024	22/05/2024	21/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFBA	%	107	105	92	98	101
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFPeA	%	103	106	92	97	101
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFHxA	%	100	105	89	93	97
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFHpA	%	104	107	91	99	102
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOA	%	109	110	92	98	104
Extracted ISTD <sup>13</sup> C <sub>5</sub> PFNA	%	103	110	89	94	100
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFDA	%	107	119	92	98	107
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFUnDA	%	120	125	97	107	125
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFDoDA	%	109	119	101	101	110
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFTeDA	%	128	133	96	109	123
Extracted ISTD <sup>13</sup> C <sub>2</sub> 4:2FTS	%	83	84	72	76	79
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%	87	104	79	83	89
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%	99	129	86	82	94
Extracted ISTD <sup>13</sup> C <sub>8</sub> FOSA	%	117	115	98	105	112
Extracted ISTD d <sub>3</sub> N MeFOSA	%	98	98	84	93	100
Extracted ISTD d <sub>5</sub> N EtFOSA	%	103	102	87	96	102
Extracted ISTD d <sub>7</sub> N MeFOSE	%	101	99	89	98	100
Extracted ISTD d <sub>9</sub> N EtFOSE	%	101	97	87	96	97
Extracted ISTD d <sub>3</sub> N MeFOSAA	%	101	114	87	92	94
Extracted ISTD d <sub>5</sub> N EtFOSAA	%	99	111	84	104	96
Total Positive PFHxS & PFOS	µg/kg	<0.1	0.2	<0.1	<0.1	<0.1
Total Positive PFOS & PFOA	µg/kg	<0.1	0.2	<0.1	<0.1	<0.1
Total Positive PFAS	µg/kg	<0.1	0.2	<0.1	<0.1	<0.1

PFAS in Soils Extended						
Our Reference	UNITS	352254-21	352254-24	352254-26	352254-28	352254-32
Your Reference		BH109	BH110	BH111	BH112	BH114
Depth		0.3	0.25	0.5	0.3	0.5
Date Sampled		20/05/2024	20/05/2024	23/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Date analysed	-	29/05/2024	29/05/2024	29/05/2024	29/05/2024	29/05/2024
Perfluorobutanesulfonic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluoropentanesulfonic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorohexanesulfonic acid - PFHxS	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluoroheptanesulfonic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorooctanesulfonic acid PFOS	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorodecanesulfonic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Perfluorobutanoic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Perfluoropentanoic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Perfluorohexanoic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluoroheptanoic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorooctanoic acid PFOA	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorononanoic acid	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Perfluorodecanoic acid	µg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Perfluoroundecanoic acid	µg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Perfluorododecanoic acid	µg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Perfluorotridecanoic acid	µg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Perfluorotetradecanoic acid	µg/kg	<5	<5	<5	<5	<5
4:2 FTS	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
6:2 FTS	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
8:2 FTS	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
10:2 FTS	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Perfluorooctane sulfonamide	µg/kg	<1	<1	<1	<1	<1
N-Methyl perfluorooctane sulfonamide	µg/kg	<1	<1	<1	<1	<1
N-Ethyl perfluorooctanesulfon amide	µg/kg	<1	<1	<1	<1	<1
N-Me perfluorooctanesulfonamid oethanol	µg/kg	<1	<1	<1	<1	<1
N-Et perfluorooctanesulfonamid oethanol	µg/kg	<5	<5	<5	<5	<5
MePerfluorooctanesulf- amid oacetic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
EtPerfluorooctanesulf amid oacetic acid	µg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%	95	97	96	90	97
Surrogate <sup>13</sup> C <sub>2</sub> PFOA	%	93	100	98	100	98
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFBS	%	90	89	89	87	88
Extracted ISTD <sup>18</sup> O <sub>2</sub> PFHxS	%	92	92	91	86	92
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%	88	91	88	88	89

PFAS in Soils Extended						
Our Reference	UNITS	352254-21	352254-24	352254-26	352254-28	352254-32
Your Reference		BH109	BH110	BH111	BH112	BH114
Depth		0.3	0.25	0.5	0.3	0.5
Date Sampled		20/05/2024	20/05/2024	23/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFBA	%	95	96	96	92	92
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFPeA	%	96	93	95	92	91
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFHxA	%	92	92	92	87	89
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFHpA	%	96	96	96	91	92
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOA	%	101	100	98	93	93
Extracted ISTD <sup>13</sup> C <sub>5</sub> PFNA	%	94	94	95	88	91
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFDA	%	103	103	97	93	93
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFUnDA	%	110	112	109	108	107
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFDoDA	%	106	105	106	101	101
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFTeDA	%	113	119	112	111	106
Extracted ISTD <sup>13</sup> C <sub>2</sub> 4:2FTS	%	73	72	71	71	71
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%	75	86	81	76	76
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%	88	90	92	90	87
Extracted ISTD <sup>13</sup> C <sub>8</sub> FOSA	%	106	108	105	103	105
Extracted ISTD d <sub>3</sub> N MeFOSA	%	93	94	92	88	91
Extracted ISTD d <sub>5</sub> N EtFOSA	%	95	94	94	89	94
Extracted ISTD d <sub>7</sub> N MeFOSE	%	95	94	96	93	93
Extracted ISTD d <sub>9</sub> N EtFOSE	%	94	95	94	91	92
Extracted ISTD d <sub>3</sub> N MeFOSAA	%	92	96	93	89	71
Extracted ISTD d <sub>5</sub> N EtFOSAA	%	93	106	88	88	71
Total Positive PFHxS & PFOS	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total Positive PFOS & PFOA	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total Positive PFAS	µg/kg	<0.1	<0.1	<0.1	<0.1	<0.1

<b>Dioxins and Furans</b>						
Our Reference		352254-1	352254-5	352254-10	352254-13	352254-17
Your Reference	UNITS	BH102	BH103	BH105	BH106	BH107
Depth		0.5	1	0.5	0.5	0.5
Date Sampled		21/05/2024	22/05/2024	21/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Please see attached		#	#	#	#	#

<b>Dioxins and Furans</b>		
Our Reference		352254-21
Your Reference	UNITS	BH109
Depth		0.3
Date Sampled		20/05/2024
Type of sample		Soil
Please see attached		#

Asbestos ID - soils NEPM - ASB-001						
Our Reference	UNITS	352254-1	352254-5	352254-10	352254-13	352254-17
Your Reference		BH102	BH103	BH105	BH106	BH107
Depth		0.5	1	0.5	0.5	0.5
Date Sampled		21/05/2024	22/05/2024	21/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date analysed	-	31/05/2024	31/05/2024	31/05/2024	31/05/2024	31/05/2024
Sample mass tested	g	183.77	297	186.82	266.99	149.24
Sample Description	-	Brown fine-grained soil & rocks				
Asbestos ID in soil (AS4964) >0.1g/kg	-	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg
		Organic fibres detected				
Trace Analysis	-	No asbestos detected				
Total Asbestos <sup>#1</sup>	g/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Asbestos ID in soil <0.1g/kg*	-	No visible asbestos detected				
ACM >7mm Estimation*	g	—	—	—	—	—
FA and AF Estimation*	g	—	—	—	—	—
ACM >7mm Estimation*	%(w/w)	<0.01	<0.01	<0.01	<0.01	<0.01
FA and AF Estimation*#2	%(w/w)	<0.001	<0.001	<0.001	<0.001	<0.001
Asbestos comments	-	Nil	Nil	Nil	Nil	Nil

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

Asbestos ID - soils NEPM - ASB-001						
Our Reference		UNITS	352254-20 BH108	352254-21 BH109	352254-24 BH110	352254-26 BH111
Your Reference			0.3	0.3	0.25	0.5
Depth						0.3
Date Sampled			23/05/2024	20/05/2024	20/05/2024	23/05/2024
Type of sample			Soil	Soil	Soil	Soil
Date analysed	-		31/05/2024	31/05/2024	31/05/2024	31/05/2024
Sample mass tested	g		130.53	211.65	426.62	325.41
Sample Description	-		Grey fine-grained soil & rocks	Brown fine-grained soil & rocks	Brown fine-grained soil & rocks	Brown fine-grained soil & rocks
Asbestos ID in soil (AS4964) >0.1g/kg	-		No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg
			Organic fibres detected	Organic fibres detected	Organic fibres detected	Organic fibres detected
Trace Analysis	-		No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected
Total Asbestos#1	g/kg		<0.1	<0.1	<0.1	<0.1
Asbestos ID in soil <0.1g/kg*	-		No visible asbestos detected			
ACM >7mm Estimation*	g		—	—	—	—
FA and AF Estimation*	g		—	—	—	—
ACM >7mm Estimation*	%(w/w)		<0.01	<0.01	<0.01	<0.01
FA and AF Estimation*#2	%(w/w)		<0.001	<0.001	<0.001	<0.001
Asbestos comments	-		Nil	Nil	Nil	Nil

Asbestos ID - soils NEPM - ASB-001			
Our Reference		352254-30	352254-32
Your Reference	UNITS	BH113	BH114
Depth		0.5	0.5
Date Sampled		20/05/2024	20/05/2024
Type of sample		Soil	Soil
Date analysed	-	31/05/2024	31/05/2024
Sample mass tested	g	177.77	70
Sample Description	-	Brown fine-grained soil & rocks	Brown fine-grained soil & rocks
Asbestos ID in soil (AS4964) >0.1g/kg	-	No asbestos detected at reporting limit of 0.1g/kg Organic fibres detected	No asbestos detected at reporting limit of 0.1g/kg Organic fibres detected
Trace Analysis	-	No asbestos detected	No asbestos detected
Total Asbestos#1	g/kg	<0.1	<0.1
Asbestos ID in soil <0.1g/kg*	-	No visible asbestos detected	No visible asbestos detected
ACM >7mm Estimation*	g	—	—
FA and AF Estimation*	g	—	—
ACM >7mm Estimation*	%(w/w)	<0.01	<0.01
FA and AF Estimation*#2	%(w/w)	<0.001	<0.001
Asbestos comments	-	Nil	Nil

Chromium Suite						
Our Reference	UNITS	352254-3	352254-4	352254-7	352254-8	352254-12
Your Reference		BH102	BH102	BH103	BH103	BH105
Depth		4	6	4	6	4
Date Sampled		21/05/2024	21/05/2024	22/05/2024	22/05/2024	21/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	24/05/2024	24/05/2024	24/05/2024	24/05/2024	24/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
pH <sub>kcl</sub>	pH units	9.1	7.8	8.9	4.0	5.6
s-TAA pH 6.5	%w/w S	<0.01	<0.01	<0.01	0.03	<0.01
TAA pH 6.5	moles H <sup>+</sup> /t	<5	<5	<5	21	<5
Chromium Reducible Sulfur	%w/w	1.4	0.37	0.68	<0.005	0.54
a-Chromium Reducible Sulfur	moles H <sup>+</sup> /t	870	230	420	<3	340
S <sub>HCl</sub>	%w/w S	[NT]	[NT]	[NT]	0.044	[NT]
S <sub>KCl</sub>	%w/w S	[NT]	[NT]	[NT]	0.038	[NT]
S <sub>NAS</sub>	%w/w S	[NT]	[NT]	[NT]	0.012	[NT]
ANC <sub>BT</sub>	% CaCO <sub>3</sub>	2.8	0.60	3.5	[NT]	[NT]
s-ANC <sub>BT</sub>	%w/w S	0.88	0.19	1.1	[NT]	[NT]
s-Net Acidity	%w/w S	0.80	0.24	<0.005	0.050	0.55
a-Net Acidity	moles H <sup>+</sup> /t	500	150	<5	30	340
Liming rate	kg CaCO <sub>3</sub> /t	38	11	<0.75	2	26
a-Net Acidity without ANCE	moles H <sup>+</sup> /t	870	230	420	30	340
Liming rate without ANCE	kg CaCO <sub>3</sub> /t	65	17	32	2.2	26
s-Net Acidity without ANCE	%w/w S	1.4	0.37	0.68	0.047	0.55

Chromium Suite						
Our Reference		352254-15	352254-16	352254-19	352254-23	352254-33
Your Reference	UNITS	BH106	BH106	BH107	BH109	BH114
Depth		3	6	4	2	1
Date Sampled		22/05/2024	22/05/2024	21/05/2024	20/05/2024	20/05/2024
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	24/05/2024	24/05/2024	24/05/2024	24/05/2024	24/05/2024
Date analysed	-	28/05/2024	28/05/2024	28/05/2024	28/05/2024	28/05/2024
pH <sub>kol</sub>	pH units	3.8	5.4	6.3	5.1	3.5
s-TAA pH 6.5	%w/w S	0.12	<0.01	<0.01	<0.01	0.16
TAA pH 6.5	moles H <sup>+</sup> /t	74	<5	<5	<5	99
Chromium Reducible Sulfur	%w/w	0.02	0.04	0.38	<0.005	0.02
a-Chromium Reducible Sulfur	moles H <sup>+</sup> /t	10	26	240	<3	10
S <sub>HCl</sub>	%w/w S	0.007	[NT]	[NT]	[NT]	0.018
S <sub>KCl</sub>	%w/w S	<0.005	[NT]	[NT]	[NT]	0.013
S <sub>NAS</sub>	%w/w S	0.009	[NT]	[NT]	[NT]	0.010
ANC <sub>BT</sub>	% CaCO <sub>3</sub>	[NT]	[NT]	[NT]	[NT]	[NT]
s-ANC <sub>BT</sub>	%w/w S	[NT]	[NT]	[NT]	[NT]	[NT]
s-Net Acidity	%w/w S	0.14	0.044	0.38	0.0060	0.18
a-Net Acidity	moles H <sup>+</sup> /t	87	27	240	<5	110
Liming rate	kg CaCO <sub>3</sub> /t	7	2	18	<0.75	8.5
a-Net Acidity without ANCE	moles H <sup>+</sup> /t	87	27	240	<5	110
Liming rate without ANCE	kg CaCO <sub>3</sub> /t	6.6	2.0	18	<0.75	8.5
s-Net Acidity without ANCE	%w/w S	0.14	0.044	0.38	0.0060	0.18

Chromium Suite			
Our Reference		352254-36	352254-37
Your Reference	UNITS	BH115	BH115
Depth		5.1	10.7
Date Sampled		23/05/2024	23/05/2024
Type of sample		Soil	Soil
Date prepared	-	24/05/2024	24/05/2024
Date analysed	-	28/05/2024	28/05/2024
pH <sub>kol</sub>	pH units	4.4	5.6
s-TAA pH 6.5	%w/w S	0.02	<0.01
TAA pH 6.5	moles H <sup>+</sup> /t	11	<5
Chromium Reducible Sulfur	%w/w	<0.005	0.21
a-Chromium Reducible Sulfur	moles H <sup>+</sup> /t	<3	130
S <sub>HCl</sub>	%w/w S	0.037	[NT]
S <sub>KCl</sub>	%w/w S	0.028	[NT]
S <sub>NAS</sub>	%w/w S	0.019	[NT]
ANC <sub>BT</sub>	% CaCO <sub>3</sub>	[NT]	[NT]
s-ANC <sub>BT</sub>	%w/w S	[NT]	[NT]
s-Net Acidity	%w/w S	0.040	0.21
a-Net Acidity	moles H <sup>+</sup> /t	22	130
Liming rate	kg CaCO <sub>3</sub> /t	2	9.8
a-Net Acidity without ANCE	moles H <sup>+</sup> /t	22	130
Liming rate without ANCE	kg CaCO <sub>3</sub> /t	1.7	9.8
s-Net Acidity without ANCE	%w/w S	0.035	0.21

<b>Misc Inorg - Soil</b>			
Our Reference		352254-2	352254-14
Your Reference	UNITS	BH102	BH106
Depth		2	2
Date Sampled		21/05/2024	22/05/2024
Type of sample		Soil	Soil
Date prepared	-	24/05/2024	24/05/2024
Date analysed	-	30/05/2024	30/05/2024
pH 1:5 soil:water	pH Units	4.4	4.9
Total Organic Carbon in soil/solids	mg/kg	3,100	2,600

<b>CEC</b>			
Our Reference		352254-2	352254-14
Your Reference	UNITS	BH102	BH106
Depth		2	2
Date Sampled		21/05/2024	22/05/2024
Type of sample		Soil	Soil
Date prepared	-	03/06/2024	03/06/2024
Date analysed	-	03/06/2024	03/06/2024
Exchangeable Ca	meq/100g	0.8	1.1
Exchangeable K	meq/100g	0.2	0.2
Exchangeable Mg	meq/100g	1.3	1.2
Exchangeable Na	meq/100g	0.8	0.3
Cation Exchange Capacity	meq/100g	3.1	2.9

<b>Clay 50-120g</b>			
Our Reference		352254-2	352254-14
Your Reference	UNITS	BH102	BH106
Depth		2	2
Date Sampled		21/05/2024	22/05/2024
Type of sample		Soil	Soil
Date prepared	-	30/05/2024	30/05/2024
Date analysed	-	31/05/2024	31/05/2024
Clay in soils <2µm	% (w/w)	20	30

Asbestos ID - soils		
Our Reference		352254-34
Your Reference	UNITS	BH115
Depth		0.2
Date Sampled		23/05/2024
Type of sample		Soil
Date analysed	-	03/06/2024
Sample mass tested	g	Approx. 25g
Sample Description	-	Brown coarse-grained soil & rocks
Asbestos ID in soil	-	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected
Asbestos comments	-	Nil
Trace Analysis	-	No asbestos detected

<b>Method ID</b>	<b>Methodology Summary</b>
<b>AS1289.3.6.3</b>	Particle Size Distribution using in house method INORG-107 by way of sieving and/or hydrometer sedimentation testing. Clay fraction at <2µm reported.
<b>ASB-001</b>	Asbestos ID - Qualitative identification of asbestos in bulk samples using Polarised Light Microscopy and Dispersion Staining Techniques including Synthetic Mineral Fibre and Organic Fibre as per Australian Standard 4964-2004.
<b>ASB-001</b>	Asbestos ID - Identification of asbestos in soil samples using Polarised Light Microscopy and Dispersion Staining Techniques. Minimum 500mL soil sample was analysed as recommended by "National Environment Protection (Assessment of site contamination) Measure, Schedule B1 and "The Guidelines from the Assessment, Remediation and Management of Asbestos-Contaminated Sites in Western Australia - May 2009" with a reporting limit of 0.1g/kg (0.01% w/w) as per Australian Standard AS4964-2004. Results reported denoted with * are outside our scope of NATA accreditation.
	<p>NOTE<sup>#1</sup> Total Asbestos g/kg was analysed and reported as per Australian Standard AS4964 (This is the sum of ACM &gt;7mm, &lt;7mm and FA/AF relative to the sample mass tested)</p> <p>NOTE<sup>#2</sup> The screening level of 0.001% w/w asbestos in soil for FA and AF only applies where the FA and AF are able to be quantified by gravimetric procedures. This screening level is not applicable to free fibres.</p> <p>Estimation = Estimated asbestos weight</p> <p>Results reported with "--" is equivalent to no visible asbestos identified using Polarised Light microscopy and Dispersion Staining Techniques.</p>
<b>Inorg-001</b>	pH - Measured using pH meter and electrode. Please note that the results for water analyses are indicative only, as analysis outside of the APHA storage times.
<b>Inorg-008</b>	Moisture content determined by heating at 105+/-5 °C for a minimum of 12 hours.

Method ID	Methodology Summary
Inorg-068	<p>Chromium Reducible Sulfur - Hydrogen Sulfide is quantified by iodometric titration after distillation to determine potential acidity.</p> <p>Net acidity including ANC has a safety factor of 1.5 applied.</p> <p>Neutralising value (NV) of 100% is assumed for liming rate.</p> <p>The recommendation that the SHCL concentration be multiplied by a factor of 2 to ensure retained acidity is not underestimated, has not been applied in the SHCL result.</p> <p>However, it has been applied in the SNAS calculation:</p> $\text{SNAS \%} = (\text{SHCL-SKCL}) \times 2$
INORG-137	Total Carbon Nitrogen Sulfur by high temperature catalytic combustion with IR detection.
Metals-020	Determination of various metals by ICP-AES.
Metals-020	Determination of exchangeable cations and cation exchange capacity in soils using 1M Ammonium Chloride exchange and ICP-OES analytical finish.
Metals-021	Determination of Mercury by Cold Vapour AAS.
Org-020	<p>Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID.</p> <p><math>F_2 = (&gt;\text{C}10-\text{C}16)\text{-Naphthalene}</math> as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.</p>
Org-020	<p>Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID.</p> <p><math>F_2 = (&gt;\text{C}10-\text{C}16)\text{-Naphthalene}</math> as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.</p> <p>Note, the Total +ve TRH PQL is reflective of the lowest individual PQL and is therefore "Total +ve TRH" is simply a sum of the positive individual TRH fractions (<math>&gt;\text{C}10\text{-C}40</math>).</p>
Org-021/022/025	<p>Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD and/or GC-MS/GC-MSMS.</p> <p>Note, the Total +ve PCBs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PCBs" is simply a sum of the positive individual PCBs.</p>
Org-022/025	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS/GC-MSMS.
Org-022/025	<p>Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-MS/GC-MSMS.</p> <p>Note, the Total +ve reported DDD+DDE+DDT PQL is reflective of the lowest individual PQL and is therefore simply a sum of the positive individually report DDD+DDE+DDT.</p>

Method ID	Methodology Summary
Org-022/025	<p>Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013.</p> <p>For soil results:-</p> <ol style="list-style-type: none"> <li>1. 'EQ PQL' values are assuming all contributing PAHs reported as &lt;PQL are actually at the PQL. This is the most conservative approach and can give false positive TEQs given that PAHs that contribute to the TEQ calculation may not be present.</li> <li>2. 'EQ zero' values are assuming all contributing PAHs reported as &lt;PQL are zero. This is the least conservative approach and is more susceptible to false negative TEQs when PAHs that contribute to the TEQ calculation are present but below PQL.</li> <li>3. 'EQ half PQL' values are assuming all contributing PAHs reported as &lt;PQL are half the stipulated PQL. Hence a mid-point between the most and least conservative approaches above.</li> </ol> <p>Note, the Total +ve PAHs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PAHs" is simply a sum of the positive individual PAHs.</p>
Org-023	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS.
Org-023	<p>Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.</p>
Org-023	<p>Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.</p> <p>Note, the Total +ve Xylene PQL is reflective of the lowest individual PQL and is therefore "Total +ve Xylenes" is simply a sum of the positive individual Xylenes.</p>
Org-029	<p>Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.</p> <p>Analysis is undertaken with LC-MS/MS.</p> <p>PFAS results include the sum of branched and linear isomers where applicable.</p> <p>Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.</p> <p>Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.</p>

<b>Method ID</b>	<b>Methodology Summary</b>
<b>ORG-038</b>	<p>Water samples are extracted with DCM and concentrated. The extract is analysed by GC/MSMS for selected Dioxin and Furans.</p> <p>Soils and Sorbents are solvent extracted, followed by an extract clean-up and GC/MSMS analysis.</p> <p>1. I -TEQ(zero) and WHO-TEQ(zero) calculated where analyte components that are &lt;PQL are considered to be zero in the TEQ calculation. Where all sample analyte results are &lt;PQL, the calculated sample TEQ = 0, this is due to the calculation being an arithmetic formula and therefore does not reflect the associated PQLs.</p> <p>2. I -TEQ(0.5) and WHO-TEQ(0.5) calculated where analyte components that are &lt;PQL are considered to be 0.5 * the component PQL in the TEQ calculation.</p> <p>3. I-TEQ(PQL) and WHO-TEQ(PQL) calculated where analyte components that are &lt;PQL are considered to be equal to the component PQL in the TEQ calculation.</p> <p>13C Rec is the recovery of Isotopically labelled compound added by the Laboratory for quantification and to measure extraction efficiency.</p> <p>I-TEF - International toxic equivalency factor I-TEQ - International toxic equivalence WHO-TEF - World Health Organisation toxic equivalency factor WHO-TEQ - World Health Organisation toxic equivalence</p>

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	352254-5
Date extracted	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Date analysed	-			29/05/2024	1	29/05/2024	29/05/2024		29/05/2024	29/05/2024
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	25	Org-023	<25	1	<25	<25	0	93	95
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	25	Org-023	<25	1	<25	<25	0	93	95
Benzene	mg/kg	0.2	Org-023	<0.2	1	<0.2	<0.2	0	94	105
Toluene	mg/kg	0.5	Org-023	<0.5	1	<0.5	<0.5	0	116	90
Ethylbenzene	mg/kg	1	Org-023	<1	1	<1	<1	0	81	94
m+p-xylene	mg/kg	2	Org-023	<2	1	<2	<2	0	86	93
o-Xylene	mg/kg	1	Org-023	<1	1	<1	<1	0	90	91
Naphthalene	mg/kg	1	Org-023	<1	1	<1	<1	0	[NT]	[NT]
Surrogate aaa-Trifluorotoluene	%		Org-023	88	1	88	84	5	81	90

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	352254-24
Date extracted	-			[NT]	17	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Date analysed	-			[NT]	17	29/05/2024	29/05/2024		29/05/2024	29/05/2024
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	25	Org-023	[NT]	17	<25	<25	0	88	82
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	25	Org-023	[NT]	17	<25	<25	0	88	82
Benzene	mg/kg	0.2	Org-023	[NT]	17	<0.2	<0.2	0	86	79
Toluene	mg/kg	0.5	Org-023	[NT]	17	<0.5	<0.5	0	104	115
Ethylbenzene	mg/kg	1	Org-023	[NT]	17	<1	<1	0	79	70
m+p-xylene	mg/kg	2	Org-023	[NT]	17	<2	<2	0	85	74
o-Xylene	mg/kg	1	Org-023	[NT]	17	<1	<1	0	84	71
Naphthalene	mg/kg	1	Org-023	[NT]	17	<1	<1	0	[NT]	[NT]
Surrogate aaa-Trifluorotoluene	%		Org-023	[NT]	17	81	69	16	99	70

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
Date analysed	-			[NT]	28	29/05/2024	29/05/2024		[NT]	[NT]
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	25	Org-023	[NT]	28	<25	<25	0	[NT]	[NT]
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	25	Org-023	[NT]	28	<25	<25	0	[NT]	[NT]
Benzene	mg/kg	0.2	Org-023	[NT]	28	<0.2	<0.2	0	[NT]	[NT]
Toluene	mg/kg	0.5	Org-023	[NT]	28	<0.5	<0.5	0	[NT]	[NT]
Ethylbenzene	mg/kg	1	Org-023	[NT]	28	<1	<1	0	[NT]	[NT]
m+p-xylene	mg/kg	2	Org-023	[NT]	28	<2	<2	0	[NT]	[NT]
o-Xylene	mg/kg	1	Org-023	[NT]	28	<1	<1	0	[NT]	[NT]
Naphthalene	mg/kg	1	Org-023	[NT]	28	<1	<1	0	[NT]	[NT]
Surrogate aaa-Trifluorotoluene	%		Org-023	[NT]	28	73	84	14	[NT]	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: svTRH (C10-C40) in Soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	352254-5
Date extracted	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Date analysed	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	50	Org-020	<50	1	<50	<50	0	109	116
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	100	Org-020	<100	1	<100	<100	0	98	111
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	100	Org-020	<100	1	<100	<100	0	86	90
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	50	Org-020	<50	1	<50	<50	0	109	116
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	100	Org-020	<100	1	<100	<100	0	98	111
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	100	Org-020	<100	1	<100	<100	0	86	90
Surrogate o-Terphenyl	%		Org-020	94	1	96	88	9	95	96

QUALITY CONTROL: svTRH (C10-C40) in Soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	352254-24
Date extracted	-			[NT]	17	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Date analysed	-			[NT]	17	28/05/2024	28/05/2024		29/05/2024	28/05/2024
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	50	Org-020	[NT]	17	<50	<50	0	118	113
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	100	Org-020	[NT]	17	<100	<100	0	112	109
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	100	Org-020	[NT]	17	<100	<100	0	100	111
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	50	Org-020	[NT]	17	<50	<50	0	118	113
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	100	Org-020	[NT]	17	<100	<100	0	112	109
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	100	Org-020	[NT]	17	<100	<100	0	100	111
Surrogate o-Terphenyl	%		Org-020	[NT]	17	87	89	2	97	93

QUALITY CONTROL: svTRH (C10-C40) in Soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
Date analysed	-			[NT]	28	29/05/2024	29/05/2024		[NT]	[NT]
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	50	Org-020	[NT]	28	<50	<50	0	[NT]	[NT]
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	100	Org-020	[NT]	28	<100	<100	0	[NT]	[NT]
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	100	Org-020	[NT]	28	<100	<100	0	[NT]	[NT]
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	50	Org-020	[NT]	28	<50	<50	0	[NT]	[NT]
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	100	Org-020	[NT]	28	<100	<100	0	[NT]	[NT]
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	100	Org-020	[NT]	28	<100	<100	0	[NT]	[NT]
Surrogate o-Terphenyl	%		Org-020	[NT]	28	87	86	1	[NT]	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: PAHs in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	352254-5
Date extracted	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Date analysed	-			03/06/2024	1	28/05/2024	28/05/2024		03/06/2024	28/05/2024
Naphthalene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	98	100
Acenaphthylene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Acenaphthene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	100	104
Fluorene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	120	124
Phenanthrene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	120	124
Anthracene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Fluoranthene	mg/kg	0.1	Org-022/025	<0.1	1	0.1	<0.1	0	132	134
Pyrene	mg/kg	0.1	Org-022/025	<0.1	1	0.1	<0.1	0	120	126
Benzo(a)anthracene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Chrysene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	96	100
Benzo(b,j+k)fluoranthene	mg/kg	0.2	Org-022/025	<0.2	1	<0.2	<0.2	0	[NT]	[NT]
Benzo(a)pyrene	mg/kg	0.05	Org-022/025	<0.05	1	0.06	<0.05	18	100	94
Indeno(1,2,3-c,d)pyrene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Dibenzo(a,h)anthracene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Benzo(g,h,i)perylene	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-022/025	127	1	112	107	5	97	97

QUALITY CONTROL: PAHs in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	352254-24
Date extracted	-			[NT]	17	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Date analysed	-			[NT]	17	28/05/2024	28/05/2024		03/06/2024	28/05/2024
Naphthalene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	114	102
Acenaphthylene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Acenaphthene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	116	104
Fluorene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	114	122
Phenanthrene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	118	126
Anthracene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Fluoranthene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	118	132
Pyrene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	116	124
Benzo(a)anthracene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Chrysene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	108	98
Benzo(b,j+k)fluoranthene	mg/kg	0.2	Org-022/025	[NT]	17	<0.2	<0.2	0	[NT]	[NT]
Benzo(a)pyrene	mg/kg	0.05	Org-022/025	[NT]	17	<0.05	<0.05	0	120	88
Indeno(1,2,3-c,d)pyrene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Dibenzo(a,h)anthracene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Benzo(g,h,i)perylene	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-022/025	[NT]	17	107	105	2	123	104

QUALITY CONTROL: PAHs in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
Date analysed	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
Naphthalene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Acenaphthylene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Acenaphthene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Fluorene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Phenanthrene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Anthracene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Fluoranthene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Pyrene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Benzo(a)anthracene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Chrysene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Benzo(b,j+k)fluoranthene	mg/kg	0.2	Org-022/025	[NT]	28	<0.2	<0.2	0	[NT]	[NT]
Benzo(a)pyrene	mg/kg	0.05	Org-022/025	[NT]	28	<0.05	<0.05	0	[NT]	[NT]
Indeno(1,2,3-c,d)pyrene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Dibenzo(a,h)anthracene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Benzo(g,h,i)perylene	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-022/025	[NT]	28	102	101	1	[NT]	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: Organochlorine Pesticides in soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	352254-5
Date extracted	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Date analysed	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
alpha-BHC	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	120	126
HCB	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
beta-BHC	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	120	126
gamma-BHC	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Heptachlor	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	120	126
delta-BHC	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Aldrin	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	118	122
Heptachlor Epoxide	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	124	130
gamma-Chlordane	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
alpha-chlordane	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Endosulfan I	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
pp-DDE	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	114	120
Dieldrin	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	118	124
Endrin	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	118	124
Endosulfan II	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
pp-DDD	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	122	128
Endrin Aldehyde	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
pp-DDT	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Endosulfan Sulphate	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	112	112
Methoxychlor	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Mirex	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Surrogate 4-Chloro-3-NBTF	%		Org-022/025	97	1	94	94	0	96	94

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: Organochlorine Pesticides in soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	352254-24
Date extracted	-			[NT]	17	28/05/2024	28/05/2024		[NT]	28/05/2024
Date analysed	-			[NT]	17	28/05/2024	28/05/2024		[NT]	28/05/2024
alpha-BHC	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	122
HCB	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
beta-BHC	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	124
gamma-BHC	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Heptachlor	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	124
delta-BHC	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Aldrin	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	122
Heptachlor Epoxide	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	128
gamma-Chlordane	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
alpha-chlordane	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Endosulfan I	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
pp-DDE	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	120
Dieldrin	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	124
Endrin	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	124
Endosulfan II	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
pp-DDD	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	124
Endrin Aldehyde	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
pp-DDT	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Endosulfan Sulphate	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	102
Methoxychlor	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Mirex	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Surrogate 4-Chloro-3-NBTF	%		Org-022/025	[NT]	17	92	92	0	[NT]	90

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: Organochlorine Pesticides in soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
Date analysed	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
alpha-BHC	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
HCB	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
beta-BHC	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
gamma-BHC	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Heptachlor	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
delta-BHC	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Aldrin	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Heptachlor Epoxide	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
gamma-Chlordane	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
alpha-chlordane	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Endosulfan I	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
pp-DDE	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Dieldrin	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Endrin	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Endosulfan II	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
pp-DDD	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Endrin Aldehyde	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
pp-DDT	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Endosulfan Sulphate	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Methoxychlor	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Mirex	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Surrogate 4-Chloro-3-NBTF	%		Org-022/025	[NT]	28	92	89	3	[NT]	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: Organophosphorus Pesticides in Soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	352254-5
Date extracted	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Date analysed	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Dichlorvos	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	140	140
Mevinphos	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Phorate	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Dimethoate	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Diazinon	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Disulfoton	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Chlorpyrifos-methyl	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Parathion-Methyl	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Ronnel	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	124	130
Fenitrothion	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	124	130
Malathion	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	134	138
Chlorpyriphos	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	128	136
Fenthion	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Parathion	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	120	122
Bromophos-ethyl	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Methidathion	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Fenamiphos	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Ethion	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	130	138
Phosalone	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Azinphos-methyl (Guthion)	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Coumaphos	mg/kg	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Surrogate 4-Chloro-3-NBTF	%		Org-022/025	97	1	94	94	0	96	94

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: Organophosphorus Pesticides in Soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	352254-24
Date extracted	-			[NT]	17	28/05/2024	28/05/2024		[NT]	28/05/2024
Date analysed	-			[NT]	17	28/05/2024	28/05/2024		[NT]	28/05/2024
Dichlorvos	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	136
Mevinphos	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Phorate	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Dimethoate	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Diazinon	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Disulfoton	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Chlorpyrifos-methyl	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Parathion-Methyl	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Ronnel	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	126
Fenitrothion	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	126
Malathion	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	128
Chlorpyriphos	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	132
Fenthion	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Parathion	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	118
Bromophos-ethyl	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Methidathion	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Fenamiphos	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Ethion	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	130
Phosalone	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Azinphos-methyl (Guthion)	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Coumaphos	mg/kg	0.1	Org-022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Surrogate 4-Chloro-3-NBTF	%		Org-022/025	[NT]	17	92	92	0	[NT]	90

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: Organophosphorus Pesticides in Soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
Date analysed	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
Dichlorvos	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Mevinphos	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Phorate	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Dimethoate	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Diazinon	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Disulfoton	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Chlorpyrifos-methyl	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Parathion-Methyl	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Ronnel	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Fenitrothion	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Malathion	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Chlorpyriphos	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Fenthion	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Parathion	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Bromophos-ethyl	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Methidathion	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Fenamiphos	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Ethion	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Phosalone	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Azinphos-methyl (Guthion)	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Coumaphos	mg/kg	0.1	Org-022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Surrogate 4-Chloro-3-NBTF	%		Org-022/025	[NT]	28	92	89	3	[NT]	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: PCBs in Soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	352254-5
Date extracted	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Date analysed	-			28/05/2024	1	28/05/2024	28/05/2024		28/05/2024	28/05/2024
Aroclor 1016	mg/kg	0.1	Org-021/022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1221	mg/kg	0.1	Org-021/022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1232	mg/kg	0.1	Org-021/022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1242	mg/kg	0.1	Org-021/022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1248	mg/kg	0.1	Org-021/022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1254	mg/kg	0.1	Org-021/022/025	<0.1	1	<0.1	<0.1	0	129	91
Aroclor 1260	mg/kg	0.1	Org-021/022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Surrogate 2-Fluorobiphenyl	%		Org-021/022/025	110	1	105	110	5	104	102

QUALITY CONTROL: PCBs in Soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	352254-24
Date extracted	-			[NT]	17	28/05/2024	28/05/2024		[NT]	28/05/2024
Date analysed	-			[NT]	17	28/05/2024	28/05/2024		[NT]	28/05/2024
Aroclor 1016	mg/kg	0.1	Org-021/022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1221	mg/kg	0.1	Org-021/022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1232	mg/kg	0.1	Org-021/022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1242	mg/kg	0.1	Org-021/022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1248	mg/kg	0.1	Org-021/022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1254	mg/kg	0.1	Org-021/022/025	[NT]	17	<0.1	<0.1	0	[NT]	93
Aroclor 1260	mg/kg	0.1	Org-021/022/025	[NT]	17	<0.1	<0.1	0	[NT]	[NT]
Surrogate 2-Fluorobiphenyl	%		Org-021/022/025	[NT]	17	106	108	2	[NT]	104

QUALITY CONTROL: PCBs in Soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
Date analysed	-			[NT]	28	28/05/2024	28/05/2024		[NT]	[NT]
Aroclor 1016	mg/kg	0.1	Org-021/022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1221	mg/kg	0.1	Org-021/022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1232	mg/kg	0.1	Org-021/022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1242	mg/kg	0.1	Org-021/022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1248	mg/kg	0.1	Org-021/022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1254	mg/kg	0.1	Org-021/022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1260	mg/kg	0.1	Org-021/022/025	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Surrogate 2-Fluorobiphenyl	%		Org-021/022/025	[NT]	28	107	105	2	[NT]	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: Acid Extractable metals in soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	352254-5
Date prepared	-			29/05/2024	1	29/05/2024	29/05/2024		29/05/2024	29/05/2024
Date analysed	-			30/05/2024	1	30/05/2024	30/05/2024		30/05/2024	30/05/2024
Arsenic	mg/kg	4	Metals-020	<4	1	<4	5	22	102	95
Cadmium	mg/kg	0.4	Metals-020	<0.4	1	<0.4	<0.4	0	101	92
Chromium	mg/kg	1	Metals-020	<1	1	8	9	12	98	89
Copper	mg/kg	1	Metals-020	<1	1	18	24	29	99	93
Lead	mg/kg	1	Metals-020	<1	1	32	48	40	104	92
Mercury	mg/kg	0.1	Metals-021	<0.1	1	<0.1	0.1	0	100	97
Nickel	mg/kg	1	Metals-020	<1	1	5	8	46	101	95
Zinc	mg/kg	1	Metals-020	<1	1	54	91	51	106	93
Iron	mg/kg	10	Metals-020	<10	[NT]	[NT]	[NT]	[NT]	99	[NT]

QUALITY CONTROL: Acid Extractable metals in soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	352254-24
Date prepared	-			[NT]	17	29/05/2024	29/05/2024		29/05/2024	29/05/2024
Date analysed	-			[NT]	17	30/05/2024	30/05/2024		30/05/2024	30/05/2024
Arsenic	mg/kg	4	Metals-020	[NT]	17	5	5	0	100	101
Cadmium	mg/kg	0.4	Metals-020	[NT]	17	<0.4	<0.4	0	99	96
Chromium	mg/kg	1	Metals-020	[NT]	17	9	9	0	100	95
Copper	mg/kg	1	Metals-020	[NT]	17	17	21	21	99	106
Lead	mg/kg	1	Metals-020	[NT]	17	19	19	0	105	97
Mercury	mg/kg	0.1	Metals-021	[NT]	17	<0.1	<0.1	0	98	96
Nickel	mg/kg	1	Metals-020	[NT]	17	8	10	22	99	101
Zinc	mg/kg	1	Metals-020	[NT]	17	51	79	43	105	110
Iron	mg/kg	10	Metals-020	[NT]	[NT]	[NT]	[NT]	[NT]	114	[NT]

QUALITY CONTROL: Acid Extractable metals in soil							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date prepared	-			[NT]	28	29/05/2024	29/05/2024		[NT]	[NT]
Date analysed	-			[NT]	28	30/05/2024	30/05/2024		[NT]	[NT]
Arsenic	mg/kg	4	Metals-020	[NT]	28	<4	4	0	[NT]	[NT]
Cadmium	mg/kg	0.4	Metals-020	[NT]	28	<0.4	<0.4	0	[NT]	[NT]
Chromium	mg/kg	1	Metals-020	[NT]	28	9	9	0	[NT]	[NT]
Copper	mg/kg	1	Metals-020	[NT]	28	13	13	0	[NT]	[NT]
Lead	mg/kg	1	Metals-020	[NT]	28	12	13	8	[NT]	[NT]
Mercury	mg/kg	0.1	Metals-021	[NT]	28	<0.1	<0.1	0	[NT]	[NT]
Nickel	mg/kg	1	Metals-020	[NT]	28	9	10	11	[NT]	[NT]
Zinc	mg/kg	1	Metals-020	[NT]	28	51	60	16	[NT]	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: PFAS in Soils Extended							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	[NT]
Date prepared	-			29/05/2024	1	29/05/2024	29/05/2024		29/05/2024	[NT]
Date analysed	-			29/05/2024	1	29/05/2024	29/05/2024		29/05/2024	[NT]
Perfluorobutanesulfonic acid	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	103	[NT]
Perfluoropentanesulfonic acid	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	100	[NT]
Perfluorohexanesulfonic acid - PFHxS	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	99	[NT]
Perfluoroheptanesulfonic acid	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	101	[NT]
Perfluorooctanesulfonic acid PFOS	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	94	[NT]
Perfluorodecanesulfonic acid	µg/kg	0.2	Org-029	<0.2	1	<0.2	<0.2	0	110	[NT]
Perfluorobutanoic acid	µg/kg	0.2	Org-029	<0.2	1	<0.2	<0.2	0	94	[NT]
Perfluoropentanoic acid	µg/kg	0.2	Org-029	<0.2	1	<0.2	<0.2	0	97	[NT]
Perfluorohexanoic acid	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	97	[NT]
Perfluoroheptanoic acid	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	99	[NT]
Perfluorooctanoic acid PFOA	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	97	[NT]
Perfluorononanoic acid	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	91	[NT]
Perfluorodecanoic acid	µg/kg	0.5	Org-029	<0.5	1	<0.5	<0.5	0	99	[NT]
Perfluoroundecanoic acid	µg/kg	0.5	Org-029	<0.5	1	<0.5	<0.5	0	100	[NT]
Perfluorododecanoic acid	µg/kg	0.5	Org-029	<0.5	1	<0.5	<0.5	0	99	[NT]
Perfluorotridecanoic acid	µg/kg	0.5	Org-029	<0.5	1	<0.5	<0.5	0	95	[NT]
Perfluorotetradecanoic acid	µg/kg	5	Org-029	<5	1	<5	<5	0	110	[NT]
4:2 FTS	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	99	[NT]
6:2 FTS	µg/kg	0.1	Org-029	<0.1	1	<0.1	<0.1	0	92	[NT]
8:2 FTS	µg/kg	0.2	Org-029	<0.2	1	<0.2	<0.2	0	115	[NT]
10:2 FTS	µg/kg	0.2	Org-029	<0.2	1	<0.2	<0.2	0	118	[NT]
Perfluorooctane sulfonamide	µg/kg	1	Org-029	<1	1	<1	<1	0	95	[NT]
N-Methyl perfluorooctane sulfonamide	µg/kg	1	Org-029	<1	1	<1	<1	0	103	[NT]
N-Ethyl perfluorooctanesulfon amide	µg/kg	1	Org-029	<1	1	<1	<1	0	102	[NT]
N-Me perfluorooctanesulfonamid oethanol	µg/kg	1	Org-029	<1	1	<1	<1	0	104	[NT]
N-Et perfluorooctanesulfonamid oethanol	µg/kg	5	Org-029	<5	1	<5	<5	0	100	[NT]
MePerfluorooctanesulf- amid oacetic acid	µg/kg	0.2	Org-029	<0.2	1	<0.2	<0.2	0	105	[NT]
EtPerfluorooctanesulf amid oacetic acid	µg/kg	0.2	Org-029	<0.2	1	<0.2	<0.2	0	97	[NT]
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%		Org-029	94	1	95	94	1	94	[NT]
Surrogate <sup>13</sup> C <sub>2</sub> PFOA	%		Org-029	98	1	97	97	0	94	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: PFAS in Soils Extended							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	[NT]
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFBS	%		Org-029	97	1	100	102	2	98	[NT]
Extracted ISTD <sup>18</sup> O <sub>2</sub> PFHxS	%		Org-029	100	1	102	103	1	96	[NT]
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%		Org-029	98	1	99	100	1	100	[NT]
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFBA	%		Org-029	105	1	107	106	1	105	[NT]
Extracted ISTD <sup>13</sup> C <sub>3</sub> PFPeA	%		Org-029	105	1	103	104	1	103	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFHxA	%		Org-029	101	1	100	104	4	100	[NT]
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFHpA	%		Org-029	103	1	104	105	1	102	[NT]
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOA	%		Org-029	105	1	109	108	1	106	[NT]
Extracted ISTD <sup>13</sup> C <sub>5</sub> PFNA	%		Org-029	104	1	103	102	1	102	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFDA	%		Org-029	107	1	107	110	3	108	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFUnDA	%		Org-029	119	1	120	126	5	107	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFDoDA	%		Org-029	107	1	109	113	4	113	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> PFTeDA	%		Org-029	115	1	128	124	3	128	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> 4:2FTS	%		Org-029	80	1	83	83	0	93	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%		Org-029	91	1	87	89	2	103	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%		Org-029	99	1	99	107	8	107	[NT]
Extracted ISTD <sup>13</sup> C <sub>8</sub> FOSA	%		Org-029	115	1	117	112	4	109	[NT]
Extracted ISTD d <sub>3</sub> N MeFOSA	%		Org-029	95	1	98	99	1	97	[NT]
Extracted ISTD d <sub>5</sub> N EtFOSA	%		Org-029	96	1	103	102	1	101	[NT]
Extracted ISTD d <sub>7</sub> N MeFOSE	%		Org-029	104	1	101	104	3	102	[NT]

QUALITY CONTROL: PFAS in Soils Extended							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	[NT]
Extracted ISTD d <sub>9</sub> N EtFOSE	%		Org-029	104	1	101	101	0	100	[NT]
Extracted ISTD d <sub>3</sub> N MeFOSAA	%		Org-029	104	1	101	104	3	99	[NT]
Extracted ISTD d <sub>5</sub> N EtFOSAA	%		Org-029	106	1	99	103	4	101	[NT]

QUALITY CONTROL: Dioxins and Furans						Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]	
Please see attached			ORG-038	#	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]	

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: Chromium Suite					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	[NT]
Date prepared	-			24/05/2024	3	24/05/2024	24/05/2024		24/05/2024	[NT]
Date analysed	-			28/05/2024	3	28/05/2024	28/05/2024		28/05/2024	[NT]
pH <sub>kcl</sub>	pH units		Inorg-068	[NT]	3	9.1	9.1	0	95	[NT]
s-TAA pH 6.5	%w/w S	0.01	Inorg-068	<0.01	3	<0.01	<0.01	0	[NT]	[NT]
TAA pH 6.5	moles H <sup>+</sup> /t	5	Inorg-068	<5	3	<5	<5	0	95	[NT]
Chromium Reducible Sulfur	%w/w	0.005	Inorg-068	<0.005	3	1.4	1.4	0	97	[NT]
a-Chromium Reducible Sulfur	moles H <sup>+</sup> /t	3	Inorg-068	<3	3	870	860	1	[NT]	[NT]
S <sub>HCl</sub>	%w/w S	0.005	Inorg-068	<0.005	3	[NT]	[NT]		[NT]	[NT]
S <sub>KCl</sub>	%w/w S	0.005	Inorg-068	<0.005	3	[NT]	[NT]		[NT]	[NT]
S <sub>NAS</sub>	%w/w S	0.005	Inorg-068	<0.005	3	[NT]	[NT]		[NT]	[NT]
ANC <sub>BT</sub>	% CaCO <sub>3</sub>	0.05	Inorg-068	<0.05	3	2.8	2.8	0	100	[NT]
s-ANC <sub>BT</sub>	%w/w S	0.05	Inorg-068	<0.05	3	0.88	0.88	0	[NT]	[NT]
s-Net Acidity	%w/w S	0.005	Inorg-068	<0.005	3	0.80	0.80	0	[NT]	[NT]
a-Net Acidity	moles H <sup>+</sup> /t	5	Inorg-068	<5	3	500	500	0	[NT]	[NT]
Liming rate	kg CaCO <sub>3</sub> /t	0.75	Inorg-068	<0.75	3	38	37	3	[NT]	[NT]
a-Net Acidity without ANCE	moles H <sup>+</sup> /t	5	Inorg-068	<5	3	870	860	1	[NT]	[NT]
Liming rate without ANCE	kg CaCO <sub>3</sub> /t	0.75	Inorg-068	<0.75	3	65	65	0	[NT]	[NT]
s-Net Acidity without ANCE	%w/w S	0.005	Inorg-068	<0.005	3	1.4	1.4	0	[NT]	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: Chromium Suite					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date prepared	-			[NT]	36	24/05/2024	24/05/2024		[NT]	[NT]
Date analysed	-			[NT]	36	28/05/2024	28/05/2024		[NT]	[NT]
pH <sub>kcl</sub>	pH units		Inorg-068	[NT]	36	4.4	4.4	0	[NT]	[NT]
s-TAA pH 6.5	%w/w S	0.01	Inorg-068	[NT]	36	0.02	0.02	0	[NT]	[NT]
TAA pH 6.5	moles H <sup>+</sup> /t	5	Inorg-068	[NT]	36	11	11	0	[NT]	[NT]
Chromium Reducible Sulfur	%w/w	0.005	Inorg-068	[NT]	36	<0.005	<0.005	0	[NT]	[NT]
a-Chromium Reducible Sulfur	moles H <sup>+</sup> /t	3	Inorg-068	[NT]	36	<3	<3	0	[NT]	[NT]
S <sub>HCl</sub>	%w/w S	0.005	Inorg-068	[NT]	36	0.037	0.037	0	[NT]	[NT]
S <sub>KCl</sub>	%w/w S	0.005	Inorg-068	[NT]	36	0.028	0.026	7	[NT]	[NT]
S <sub>NAS</sub>	%w/w S	0.005	Inorg-068	[NT]	36	0.019	0.022	15	[NT]	[NT]
s-Net Acidity	%w/w S	0.005	Inorg-068	[NT]	36	0.040	0.043	7	[NT]	[NT]
a-Net Acidity	moles H <sup>+</sup> /t	5	Inorg-068	[NT]	36	22	23	4	[NT]	[NT]
Liming rate	kg CaCO <sub>3</sub> /t	0.75	Inorg-068	[NT]	36	2	2	0	[NT]	[NT]
a-Net Acidity without ANCE	moles H <sup>+</sup> /t	5	Inorg-068	[NT]	36	22	23	4	[NT]	[NT]
Liming rate without ANCE	kg CaCO <sub>3</sub> /t	0.75	Inorg-068	[NT]	36	1.7	1.7	0	[NT]	[NT]
s-Net Acidity without ANCE	%w/w S	0.005	Inorg-068	[NT]	36	0.035	0.037	6	[NT]	[NT]

QUALITY CONTROL: Misc Inorg - Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	[NT]
Date prepared	-			24/05/2024	2	24/05/2024	24/05/2024		24/05/2024	[NT]
Date analysed	-			29/05/2024	2	30/05/2024	30/05/2024		29/05/2024	[NT]
pH 1:5 soil:water	pH Units		Inorg-001	[NT]	2	4.4	4.4	0	100	[NT]
Total Organic Carbon in soil/solids	mg/kg	100	INORG-137	<100	2	3100	[NT]		87	[NT]

**Client Reference: Detailed Site Investigation:25-27 Leeds St, Rhodes**

QUALITY CONTROL: CEC						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	352254-14
Date prepared	-			03/06/2024	2	03/06/2024	03/06/2024		03/06/2024	03/06/2024
Date analysed	-			03/06/2024	2	03/06/2024	03/06/2024		03/06/2024	03/06/2024
Exchangeable Ca	meq/100g	0.1	Metals-020	<0.1	2	0.8	1	22	88	106
Exchangeable K	meq/100g	0.1	Metals-020	<0.1	2	0.2	0.2	0	104	113
Exchangeable Mg	meq/100g	0.1	Metals-020	<0.1	2	1.3	1.5	14	86	101
Exchangeable Na	meq/100g	0.1	Metals-020	<0.1	2	0.8	0.7	13	115	122

## Result Definitions

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<	Less than
>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	
The recommended maximums for analytes in urine are taken from "2018 TLVs and BEIs", as published by ACGIH (where available). Limit provided for Nickel is a precautionary guideline as per Position Paper prepared by AIOP Exposure Standards Committee, 2016.	
Guideline limits for Rinse Water Quality reported as per analytical requirements and specifications of AS 4187, Amdt 2 2019, Table 7.2	

## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Where matrix spike recoveries fall below the lower limit of the acceptance criteria (e.g. for non-labile or standard Organics <60%), positive result(s) in the parent sample will subsequently have a higher than typical estimated uncertainty (MU estimates supplied on request) and in these circumstances the sample result is likely biased significantly low.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

## **Report Comments**

### **Acid Extractable Metals in Soil:**

- The laboratory RPD acceptance criteria has been exceeded for 352254-1 for Pb and Zn. Therefore a triplicate result has been issued as laboratory sample number 352254-42.
- The laboratory RPD acceptance criteria has been exceeded for 352254-17 for Zn. Therefore a triplicate result has been issued as laboratory sample number 352254-43.

**CHROMIUM\_SUITE:**Samples were out of the recommended holding time for this analysis.

### **Asbestos-ID in soil: NEPM**

This report is consistent with the reporting recommendations in the National Environment Protection (Assessment of Site Contamination) Measure, Schedule B1, May 2013. This is reported outside our scope of NATA accreditation.

Note: All samples analysed as received. However, all samples are below the minimum recommended 500mL sample volume as per National Environment Protection (Assessment of Site Contamination) Measure, Schedule B1, May 2013.

**Asbestos:** A portion of the supplied sample was sub-sampled for asbestos according to ASB-001 asbestos subsampling procedure. We cannot guarantee that this sub-sample is indicative of the entire sample. Envirolab/MPL recommends supplying 40-60g or 500ml of sample in its own container.

Note: Sample 352254-34 was sub-sampled from a jar provided by the client.

Dioxins analysed by MPL Laboratories. Report no. PFE1917.

# Please see attached report.

## Certificate of Analysis PFE1917

### Client Details

<b>Client</b>	Envirolab (Sydney)
<b>Contact</b>	Results Receivable
<b>Address</b>	12 Ashley St, Chatswood, NSW, 2067

### Sample Details

<b>Your Reference</b>	352254
<b>Number of Samples</b>	6 Soil, 3 Swab
<b>Date Samples Received</b>	29/05/2024
<b>Date Instructions Received</b>	29/05/2024

### Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.  
Samples were analysed as received from the client. Results relate specifically to the samples as received.  
Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

### Report Details

<b>Date Results Requested by</b>	13/06/2024
<b>Date of Reissue</b>	20/06/2024 - This report supercedes previous report, see amendment history for details

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**Accredited for compliance with ISO/IEC 17025. Tests not covered by NATA are denoted with \*.**

### Authorisation Details

<b>Results Approved By</b>	Heram Halim, Operations Manager Steven Luong, Senior Chemist Travis Carey, Organics Supervisor
<b>Laboratory Manager</b>	Michael Kubiak

# Certificate of Analysis PFE1917

## Report Amendment History

Revision	Reason for Amendment
R-01	This report contains samples from job reference 352299.
R-02	This report contains samples from job reference 352254.
R-03	Corrected TEQ reporting.

# Certificate of Analysis PFE1917

## Samples in this Report

Envirolab ID	Sample ID	Description	Matrix	Date Sampled	Date Received
PFE1917-04	BH102_0.5	352254-1	Soil	21/05/2024	29/05/2024
PFE1917-05	BH103_1.0	352254-5	Soil	22/05/2024	29/05/2024
PFE1917-06	BH105_0.5	352254-10	Soil	21/05/2024	29/05/2024
PFE1917-07	BH106_0.5	352254-13	Soil	22/05/2024	29/05/2024
PFE1917-08	BH107_0.5	352254-17	Soil	21/05/2024	29/05/2024
PFE1917-09	BH109_0.5	352254-21	Soil	20/05/2024	29/05/2024

# Certificate of Analysis PFE1917

## Inorganics - Moisture (Soil) - Analysed By Envirolab Services Sydney

Envirolab ID	Units	PQL	PFE1917-04	PFE1917-05	PFE1917-06	PFE1917-07	PFE1917-08
<b>Your Reference</b>			BH102_0.5	BH103_1.0	BH105_0.5	BH106_0.5	BH107_0.5
			352254-1	352254-5	352254-10	352254-13	352254-17
<b>Date Sampled</b>			21/05/2024	22/05/2024	21/05/2024	22/05/2024	21/05/2024
Moisture	%	0.10	10	5.7	11	12	8.6
Envirolab ID	Units	PQL	PFE1917-09				
<b>Your Reference</b>			BH109_0.5				
			352254-21				
<b>Date Sampled</b>			20/05/2024				
Moisture	%	0.10	17				

# Certificate of Analysis PFE1917

## Dioxins/Furans (Soil)

<b>Envirolab ID:</b>	PFE1917-04	<b>Date Sampled:</b>	21/05/2024					
<b>Client ID:</b>	BH102_0.5							
<b>Analyte</b>								
2,3,7,8-TCDD	0.50 WHO-TEF 1	pg/g WHO-TEQ1 0.0	<0.50 WHO-TEQ2 0.25	WHO-TEQ3 0.50	I-TEF 1	I-TEQ1 0.0	I-TEQ2 0.25	I-TEQ3 0.50
1,2,3,7,8-PeCDD	2.00 WHO-TEF 1	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 1.0	WHO-TEQ3 2.0	I-TEF 0.5	I-TEQ1 0.0	I-TEQ2 0.50	I-TEQ3 1.0
1,2,3,4,7,8-HxCDD	2.00 WHO-TEF 0.1	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.10	WHO-TEQ3 0.20	I-TEF 0.1	I-TEQ1 0.0	I-TEQ2 0.10	I-TEQ3 0.20
1,2,3,6,7,8-HxCDD	2.00 WHO-TEF 0.1	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.10	WHO-TEQ3 0.20	I-TEF 0.1	I-TEQ1 0.0	I-TEQ2 0.10	I-TEQ3 0.20
1,2,3,7,8,9-HxCDD	2.00 WHO-TEF 0.1	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.10	WHO-TEQ3 0.20	I-TEF 0.1	I-TEQ1 0.0	I-TEQ2 0.10	I-TEQ3 0.20
1,2,3,4,6,7,8-HpCDD	2.00 WHO-TEF 0.01	pg/g WHO-TEQ1 1.2	120 WHO-TEQ2 1.2	WHO-TEQ3 1.2	I-TEF 0.01	I-TEQ1 1.2	I-TEQ2 1.2	I-TEQ3 1.2
OCDD	5.00 WHO-TEF 0.0003	pg/g WHO-TEQ1 0.41	1400 WHO-TEQ2 0.41	WHO-TEQ3 0.41	I-TEF 0.001	I-TEQ1 1.4	I-TEQ2 1.4	I-TEQ3 1.4
2,3,7,8-TCDF	0.50 WHO-TEF 0.1	pg/g WHO-TEQ1 0.0	<0.50 WHO-TEQ2 0.025	WHO-TEQ3 0.050	I-TEF 0.1	I-TEQ1 0.0	I-TEQ2 0.025	I-TEQ3 0.050
1,2,3,7,8-PeCDF	2.00 WHO-TEF 0.03	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.030	WHO-TEQ3 0.060	I-TEF 0.05	I-TEQ1 0.0	I-TEQ2 0.050	I-TEQ3 0.10
2,3,4,7,8-PeCDF	2.00 WHO-TEF 0.3	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.30	WHO-TEQ3 0.60	I-TEF 0.5	I-TEQ1 0.0	I-TEQ2 0.50	I-TEQ3 1.0
1,2,3,4,7,8-HxCDF	2.00 WHO-TEF 0.1	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.10	WHO-TEQ3 0.20	I-TEF 0.1	I-TEQ1 0.0	I-TEQ2 0.10	I-TEQ3 0.20
1,2,3,6,7,8-HxCDF	2.00 WHO-TEF 0.1	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.10	WHO-TEQ3 0.20	I-TEF 0.1	I-TEQ1 0.0	I-TEQ2 0.10	I-TEQ3 0.20
1,2,3,7,8,9-HxCDF	2.00 WHO-TEF 0.1	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.10	WHO-TEQ3 0.20	I-TEF 0.1	I-TEQ1 0.0	I-TEQ2 0.10	I-TEQ3 0.20
2,3,4,6,7,8-HxCDF	2.00 WHO-TEF 0.1	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.10	WHO-TEQ3 0.20	I-TEF 0.1	I-TEQ1 0.0	I-TEQ2 0.10	I-TEQ3 0.20
1,2,3,4,6,7,8-HpCDF	2.00 WHO-TEF 0.01	pg/g WHO-TEQ1 0.56	57 WHO-TEQ2 0.56	WHO-TEQ3 0.56	I-TEF 0.01	I-TEQ1 0.56	I-TEQ2 0.56	I-TEQ3 0.56
1,2,3,4,7,8,9-HpCDF	2.00 WHO-TEF 0.01	pg/g WHO-TEQ1 0.0	<2.0 WHO-TEQ2 0.010	WHO-TEQ3 0.020	I-TEF 0.01	I-TEQ1 0.0	I-TEQ2 0.010	I-TEQ3 0.020
OCDF	5.00 WHO-TEF 0.0003	pg/g WHO-TEQ1 0.0053	18 WHO-TEQ2 0.0053	WHO-TEQ3 0.0053	I-TEF 0.001	I-TEQ1 0.018	I-TEQ2 0.018	I-TEQ3 0.018

# Certificate of Analysis PFE1917

<b>Envirolab ID:</b>	PFE1917-05	<b>Date Sampled:</b>		22/05/2024					
<b>Client ID:</b>	BH103_1.0								
Analyte	PQL	Units	Result						
2,3,7,8-TCDD	0.50 <i>WHO-TEF</i> 1	pg/g <i>WHO-TEQ1</i>	<0.50 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,7,8-PeCDD	2.00 <i>WHO-TEF</i> 1	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,4,7,8-HxCDD	2.00 <i>WHO-TEF</i> 0.1	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,6,7,8-HxCDD	2.00 <i>WHO-TEF</i> 0.1	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,7,8,9-HxCDD	2.00 <i>WHO-TEF</i> 0.1	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	-
1,2,3,4,6,7,8-HpCDD	2.00 <i>WHO-TEF</i> 0.01	pg/g <i>WHO-TEQ1</i>	7.3 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
OCDD	5.00 <i>WHO-TEF</i> 0.0003	pg/g <i>WHO-TEQ1</i>	760 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
2,3,7,8-TCDF	0.50 <i>WHO-TEF</i> 0.1	pg/g <i>WHO-TEQ1</i>	<0.50 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,7,8-PeCDF	2.00 <i>WHO-TEF</i> 0.03	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
2,3,4,7,8-PeCDF	2.00 <i>WHO-TEF</i> 0.3	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,4,7,8-HxCDF	2.00 <i>WHO-TEF</i> 0.1	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,6,7,8-HxCDF	2.00 <i>WHO-TEF</i> 0.1	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,7,8,9-HxCDF	2.00 <i>WHO-TEF</i> 0.1	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
2,3,4,6,7,8-HxCDF	2.00 <i>WHO-TEF</i> 0.1	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,4,6,7,8-HpCDF	2.00 <i>WHO-TEF</i> 0.01	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
1,2,3,4,7,8,9-HpCDF	2.00 <i>WHO-TEF</i> 0.01	pg/g <i>WHO-TEQ1</i>	<2.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
OCDF	5.00 <i>WHO-TEF</i> 0.0003	pg/g <i>WHO-TEQ1</i>	<5.0 <i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	-

# Certificate of Analysis PFE1917

<b>Envirolab ID:</b>	PFE1917-06	<b>Date Sampled:</b>	21/05/2024									
<b>Client ID:</b>	BH105_0.5											
Analyte	PQL	Units	Result	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
2,3,7,8-TCDD	0.50	pg/g	<0.50									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	1	0.0	0.25	0.50				1	0.0	0.25	0.50	96.0%
1,2,3,7,8-PeCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	1	0.0	1.0	2.0				0.5	0.0	0.50	1.0	97.4%
1,2,3,4,7,8-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	90.6%
1,2,3,6,7,8-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	95.2%
1,2,3,7,8,9-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	-
1,2,3,4,6,7,8-HpCDD	2.00	pg/g	21									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.01	0.21	0.21	0.21				0.01	0.21	0.21	0.21	86.9%
OCDD	5.00	pg/g	2300									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.0003	0.68	0.68	0.68				0.001	2.3	2.3	2.3	84.7%
2,3,7,8-TCDF	0.50	pg/g	<0.50									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.1	0.0	0.025	0.050				0.1	0.0	0.025	0.050	86.8%
1,2,3,7,8-PeCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.03	0.0	0.030	0.060				0.05	0.0	0.050	0.10	84.3%
2,3,4,7,8-PeCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.3	0.0	0.30	0.60				0.5	0.0	0.50	1.0	98.7%
1,2,3,4,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	96.6%
1,2,3,6,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	87.6%
1,2,3,7,8,9-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	89.3%
2,3,4,6,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	88.1%
1,2,3,4,6,7,8-HpCDF	2.00	pg/g	3.9									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.01	0.039	0.039	0.039				0.01	0.039	0.039	0.039	83.0%
1,2,3,4,7,8,9-HpCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.01	0.0	0.010	0.020				0.01	0.0	0.010	0.020	84.4%
OCDF	5.00	pg/g	<5.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3								
	0.0003	0.0	0.00075	0.0015				0.001	0.0	0.0025	0.0050	-

# Certificate of Analysis PFE1917

<b>Envirolab ID:</b>	PFE1917-07	<b>Date Sampled:</b>	22/05/2024									
<b>Client ID:</b>	BH106_0.5											
Analyte	PQL	Units	Result	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
2,3,7,8-TCDD	0.50	pg/g	<0.50									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	
	1	0.0	0.25	0.50				1	0.0	0.25	0.50	73.2%
1,2,3,7,8-PeCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	1	0.0	1.0	2.0				0.5	0.0	0.50	1.0	70.1%
1,2,3,4,7,8-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	75.5%
1,2,3,6,7,8-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	68.2%
1,2,3,7,8,9-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	-
1,2,3,4,6,7,8-HpCDD	2.00	pg/g	13									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.13	0.13	0.13				0.01	0.13	0.13	0.13	55.7% [2]
OCDD	5.00	pg/g	580									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.0003	0.17	0.17	0.17				0.001	0.58	0.58	0.58	47.7% [2]
2,3,7,8-TCDF	0.50	pg/g	<0.50									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.025	0.050				0.1	0.0	0.025	0.050	73.5%
1,2,3,7,8-PeCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.03	0.0	0.030	0.060				0.05	0.0	0.050	0.10	69.8%
2,3,4,7,8-PeCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.3	0.0	0.30	0.60				0.5	0.0	0.50	1.0	75.6%
1,2,3,4,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	63.3%
1,2,3,6,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	58.9% [2]
1,2,3,7,8,9-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	61.5%
2,3,4,6,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	67.4%
1,2,3,4,6,7,8-HpCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.0	0.010	0.020				0.01	0.0	0.010	0.020	53.4% [2]
1,2,3,4,7,8,9-HpCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.0	0.010	0.020				0.01	0.0	0.010	0.020	52.7% [2]
OCDF	5.00	pg/g	<5.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.0003	0.0	0.00075	0.0015				0.001	0.0	0.0025	0.0050	-

# Certificate of Analysis PFE1917

<b>Envirolab ID:</b>	PFE1917-08	<b>Date Sampled:</b>	21/05/2024									
<b>Client ID:</b>	BH107_0.5											
Analyte	PQL	Units	Result	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
2,3,7,8-TCDD	0.50	pg/g	<0.50									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	
	1	0.0	0.25	0.50				1	0.0	0.25	0.50	72.0%
1,2,3,7,8-PeCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	1	0.0	1.0	2.0				0.5	0.0	0.50	1.0	74.1%
1,2,3,4,7,8-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	79.7%
1,2,3,6,7,8-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	75.7%
1,2,3,7,8,9-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	-
1,2,3,4,6,7,8-HpCDD	2.00	pg/g	12									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.12	0.12	0.12				0.01	0.12	0.12	0.12	71.5%
OCDD	5.00	pg/g	1200									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.0003	0.36	0.36	0.36				0.001	1.2	1.2	1.2	69.2%
2,3,7,8-TCDF	0.50	pg/g	<0.50									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.025	0.050				0.1	0.0	0.025	0.050	75.1%
1,2,3,7,8-PeCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.03	0.0	0.030	0.060				0.05	0.0	0.050	0.10	73.0%
2,3,4,7,8-PeCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.3	0.0	0.30	0.60				0.5	0.0	0.50	1.0	78.5%
1,2,3,4,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	78.5%
1,2,3,6,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	71.9%
1,2,3,7,8,9-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	68.9%
2,3,4,6,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	76.4%
1,2,3,4,6,7,8-HpCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.0	0.010	0.020				0.01	0.0	0.010	0.020	68.4%
1,2,3,4,7,8,9-HpCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.0	0.010	0.020				0.01	0.0	0.010	0.020	67.8%
OCDF	5.00	pg/g	<5.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.0003	0.0	0.00075	0.0015				0.001	0.0	0.0025	0.0050	-

# Certificate of Analysis PFE1917

<b>Envirolab ID:</b>	PFE1917-09	<b>Date Sampled:</b>		20/05/2024								
<b>Client ID:</b>	BH109_0.5											
Analyte	PQL	Units	Result	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
2,3,7,8-TCDD	0.50	pg/g	<0.50									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	
	1	0.0	0.25	0.50				1	0.0	0.25	0.50	63.9%
1,2,3,7,8-PeCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	1	0.0	1.0	2.0				0.5	0.0	0.50	1.0	62.2%
1,2,3,4,7,8-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	54.2% [2]
1,2,3,6,7,8-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	47.1% [2]
1,2,3,7,8,9-HxCDD	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	-
1,2,3,4,6,7,8-HpCDD	2.00	pg/g	6.1									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.061	0.061	0.061				0.01	0.061	0.061	0.061	44.3% [2]
OCDD	5.00	pg/g	800									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.0003	0.24	0.24	0.24				0.001	0.80	0.80	0.80	31.1% [2]
2,3,7,8-TCDF	0.50	pg/g	<0.50									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.025	0.050				0.1	0.0	0.025	0.050	70.1%
1,2,3,7,8-PeCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.03	0.0	0.030	0.060				0.05	0.0	0.050	0.10	59.1% [2]
2,3,4,7,8-PeCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.3	0.0	0.30	0.60				0.5	0.0	0.50	1.0	67.1%
1,2,3,4,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	54.7% [2]
1,2,3,6,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	51.9% [2]
1,2,3,7,8,9-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	53.0% [2]
2,3,4,6,7,8-HxCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.10	0.20				0.1	0.0	0.10	0.20	56.8% [2]
1,2,3,4,6,7,8-HpCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.0	0.010	0.020				0.01	0.0	0.010	0.020	37.0% [2]
1,2,3,4,7,8,9-HpCDF	2.00	pg/g	<2.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.0	0.010	0.020				0.01	0.0	0.010	0.020	35.7% [2]
OCDF	5.00	pg/g	<5.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3				I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.0003	0.0	0.00075	0.0015				0.001	0.0	0.0025	0.0050	-

# Certificate of Analysis PFE1917

## Result Comments

Identifier	Description
[2]	Surrogate recovery was outside routine acceptance criteria (60-140%) due to sample matrix effects. This may be due to the presence of carbon and/or other artefacts. An acceptable recovery was achieved for the LCS surrogates.

# Certificate of Analysis PFE1917

## Method Summary

Method ID	Methodology Summary
INORG-008	Moisture content determined by heating at 105+/-5 °C for a minimum of 12 hours.
ORG-025	Determination of semi-volatile organic compounds (SVOCs) by GC-MS-MS. Water samples are extracted by LLE and soils/solids/biota using DCM/Acetone/Methanol.
ORG-025_DIOXIN	Water samples are extracted with DCM and concentrated. The extract is analysed by GC-MSMS for Dioxin and Furans. Soils, Biota and Sorbents are solvent extracted, followed by clean-up and GC-MSMS analysis. 1. I -TEQ(zero) and WHO-TEQ(zero) calculated where analyte components that are <PQL are considered to be zero in the TEQ calculation. Where all sample analyte results are <PQL, the calculated sample TEQ = 0, this is due to the calculation being an arithmetic formula and therefore does not reflect the associated PQLs. 2. I -TEQ(0.5) and WHO-TEQ(0.5) calculated where analyte components that are <PQL are considered to be 0.5 * the component PQL in the TEQ calculation. 3. I-TEQ(PQL) and WHO-TEQ(PQL) calculated where analyte components that are <PQL are considered to be equal to the component PQL in the TEQ calculation. 13C12 Rec is the recovery of Isotopically labelled compound added by the Laboratory for quantification and to measure extraction efficiency. I-TEF - International toxic equivalency factor I-TEQ - International toxic equivalence WHO-TEF - World Health Organisation toxic equivalency factor WHO-TEQ - World Health Organisation toxic equivalence TEQ values are rounded to the same number of significant figures as the raw results for consistency and therefore may not calculate out exactly as PQL * TEF, given rounded up or down as appropriate.

# Certificate of Analysis PFE1917

## Result Definitions

Identifier	Description
<b>NR</b>	Not reported
<b>NEPM</b>	National Environment Protection Measure
<b>NS</b>	Not specified
<b>LCS</b>	Laboratory Control Sample
<b>RPD</b>	Relative Percent Difference
>	Greater than
<	Less than
<b>PQL</b>	Practical Quantitation Limit
<b>INS</b>	Insufficient sample for this test
<b>NA</b>	Test not required
<b>NT</b>	Not tested
<b>DOL</b>	Samples rejected due to particulate overload (air filters only)
<b>RFD</b>	Samples rejected due to filter damage (air filters only)
<b>RUD</b>	Samples rejected due to uneven deposition (air filters only)
<b>##</b>	Indicates a laboratory acceptance criteria outlier, for further details, see Result Comments and/or QC Comments

## Quality Control Definitions

### Blank

This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, and is determined by processing solvents and reagents in exactly the same manner as for samples.

### Surrogate Spike

Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.

### LCS (Laboratory Control Sample)

This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.

### Matrix Spike

A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.

### Duplicate

This is the complete duplicate analysis of a sample from the process batch. The sample selected should be one where the analyte concentration is easily measurable.

# Certificate of Analysis PFE1917

## Laboratory Acceptance Criteria

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Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria. Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction. Spikes for Physical and Aggregate Tests are not applicable. For VOCs in water samples, three vials are required for duplicate or spike analysis.

General Acceptance Criteria (GAC) - Analyte specific criteria applies for some analytes and is reflected in QC recovery tables.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% - see ELN-P05 QAQC tables for details (available on request); <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase. Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals; 60-140% for organics (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was typically insufficient in order to satisfy laboratory QA/QC protocols.

## Miscellaneous Information

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When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached. We have taken the sampling date as being the date received at the laboratory.

Two significant figures are reported for the majority of tests and with a high degree of confidence, for results <10\*xPQL, the second significant figure may be in doubt i.e. has a relatively high degree of uncertainty and is provided for information only.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS where sediment/solids are included by default.

Urine Analysis - The BEI values listed are taken from the 2022 edition of *TLVs and BEIs Threshold Limits by ACGIH*.

Air volume measurements are not covered by Envirolab's NATA accreditation.

# Data Quality Assessment Summary PFE1917

## Client Details

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<b>Client</b>	Envirolab (Sydney)
<b>Your Reference</b>	352254
<b>Date Issued</b>	20/06/2024

## Recommended Holding Time Compliance

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No recommended holding time exceedances

## Quality Control and QC Frequency

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QC Type	Compliant	Details
Blank	Yes	No Outliers
LCS	Yes	No Outliers
Duplicates	Yes	No Outliers
Matrix Spike	No	Matrix Spike Outliers Exist - See detailed list below
Surrogates / Extracted Internal Standards	Yes	No Outliers
QC Frequency	Yes	No Outliers

Surrogates/Extracted Internal Standards, Duplicates and/or Matrix Spikes are not always relevant/applicable to certain analyses and matrices. Therefore, said QC measures are deemed compliant in these situations by default. See Laboratory Acceptance Criteria for more information

# Data Quality Assessment Summary PFE1917

## Recommended Holding Time Compliance

Analysis	Sample Number(s)	Date Sampled	Date Extracted	Date Analysed	Compliant
1,2,3,4,6,7,8-HpCDD   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,4,6,7,8-HpCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,4,7,8,9-HpCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,4,7,8-HxCDD   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,4,7,8-HxCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,6,7,8-HxCDD   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,6,7,8-HxCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,7,8,9-HxCDD   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,7,8,9-HxCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes

# Data Quality Assessment Summary PFE1917

## Recommended Holding Time Compliance

Analysis	Sample Number(s)	Date Sampled	Date Extracted	Date Analysed	Compliant
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,7,8-PeCDD   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
1,2,3,7,8-PeCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
2,3,4,6,7,8-HxCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
2,3,4,7,8-PeCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
2,3,7,8-TCDD   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
2,3,7,8-TCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
OCDD   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
OCDF   Soil	9	20/05/2024	31/05/2024	11/06/2024	Yes
	4	21/05/2024	31/05/2024	10/06/2024	Yes
	6, 8	21/05/2024	31/05/2024	11/06/2024	Yes
	5	22/05/2024	31/05/2024	10/06/2024	Yes
	7	22/05/2024	31/05/2024	11/06/2024	Yes
Moisture   Soil	9	20/05/2024	28/05/2024	29/05/2024	Yes
	4, 6, 8	21/05/2024	28/05/2024	29/05/2024	Yes
	5, 7	22/05/2024	28/05/2024	29/05/2024	Yes

# Data Quality Assessment Summary PFE1917

## Outliers: Matrix Spike

ORG-025\_DIOXIN | Dioxins/Furans (Soil) | Batch BFE5524

Sample ID	Analyte	% Limits	% Recovery
BFE5524-MS1#	OCDD	60 - 140	##[1]

# Quality Control PFE1917

## ORG-025\_DIOXIN | Dioxins/Furans (Soil) | Batch BFE5524

Analyte	Units	PQL	Blank	DUP1 BFE5524-DUP1#	LCS %	Spike % BFE5524-MS1#
				Samp   QC   RPD %		
2,3,7,8-TCDD	pg/g	0.50	<0.50	<0.50   <0.50   [NA]	95.7	92.8
Surrogate 13C-2,3,7,8-TCDD	%		75.7	83.4 / 88.3	115	88.8
2,3,7,8-TCDD WHO-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,7,8-TCDD WHO-TEQ2	pg/g	0.250	0.250	0.250   0.250   0.00	[NA]	[NA]
2,3,7,8-TCDD WHO-TEQ3	pg/g	0.500	0.500	0.500   0.500   0.00	[NA]	[NA]
2,3,7,8-TCDD I-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,7,8-TCDD I-TEQ2	pg/g	0.250	0.250	0.250   0.250   0.00	[NA]	[NA]
2,3,7,8-TCDD I-TEQ3	pg/g	0.500	0.500	0.500   0.500   0.00	[NA]	[NA]
1,2,3,7,8-PeCDD	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	98.6	96.5
Surrogate 13C-1,2,3,7,8-PeCDD	%		84.5	93.0 / 82.5	107	92.7
1,2,3,7,8-PeCDD WHO-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8-PeCDD WHO-TEQ2	pg/g	1.00	1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,7,8-PeCDD WHO-TEQ3	pg/g	2.00	2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,7,8-PeCDD I-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8-PeCDD I-TEQ2	pg/g	0.500	0.500	0.500   0.500   0.00	[NA]	[NA]
1,2,3,7,8-PeCDD I-TEQ3	pg/g	1.00	1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDD	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	121	112
Surrogate 13C-1,2,3,4,7,8-HxCDD	%		83.4	86.2 / 89.5	103	91.7
1,2,3,4,7,8-HxCDD WHO-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8-HxCDD WHO-TEQ2	pg/g	0.100	0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDD WHO-TEQ3	pg/g	0.200	0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDD I-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8-HxCDD I-TEQ2	pg/g	0.100	0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDD I-TEQ3	pg/g	0.200	0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDD	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	88.8	88.9
Surrogate 13C-1,2,3,6,7,8-HxCDD	%		77.9	82.8 / 90.3	111	88.6
1,2,3,6,7,8-HxCDD WHO-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,6,7,8-HxCDD WHO-TEQ2	pg/g	0.100	0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDD WHO-TEQ3	pg/g	0.200	0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDD I-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,6,7,8-HxCDD I-TEQ2	pg/g	0.100	0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDD I-TEQ3	pg/g	0.200	0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDD	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	90.5	85.4
1,2,3,7,8,9-HxCDD WHO-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8,9-HxCDD WHO-TEQ2	pg/g	0.100	0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDD WHO-TEQ3	pg/g	0.200	0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDD I-TEQ1	pg/g	0.00	0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8,9-HxCDD I-TEQ2	pg/g	0.100	0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDD I-TEQ3	pg/g	0.200	0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD	pg/g	2.0	<2.0	116   93.8   21.1	99.6	84.2
Surrogate 13C-1,2,3,4,6,7,8-HpCDD	%		78.5	76.9 / 78.1	109	91.5
1,2,3,4,6,7,8-HpCDD WHO-TEQ1	pg/g	0.00	1.16	1.16   0.938   21.1	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD WHO-TEQ2	pg/g	0.0100	1.16	1.16   0.938   21.1	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD WHO-TEQ3	pg/g	0.0200	1.16	1.16   0.938   21.1	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD I-TEQ1	pg/g	0.00	1.16	1.16   0.938   21.1	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD I-TEQ2	pg/g	0.0100	1.16	1.16   0.938   21.1	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD I-TEQ3	pg/g	0.0200	1.16	1.16   0.938   21.1	[NA]	[NA]
OCDD	pg/g	5.0	<5.0	1370   1230   10.9	108	##[1]
Surrogate 13C-OCDD	%		73.8	81.3 / 81.2	104	86.0
OCDD WHO-TEQ1	pg/g	0.00	0.412	0.412   0.370   10.8	[NA]	[NA]
OCDD WHO-TEQ2	pg/g	0.000750	0.412	0.412   0.370   10.8	[NA]	[NA]
OCDD WHO-TEQ3	pg/g	0.00150	0.412	0.412   0.370   10.8	[NA]	[NA]
OCDD I-TEQ1	pg/g	0.00	1.37	1.37   1.23   10.8	[NA]	[NA]
OCDD I-TEQ2	pg/g	0.00250	1.37	1.37   1.23   10.8	[NA]	[NA]
OCDD I-TEQ3	pg/g	0.00500	1.37	1.37   1.23   10.8	[NA]	[NA]
2,3,7,8-TCDF	pg/g	0.50	<0.50	<0.50   <0.50   [NA]	104	102
Surrogate 13C-2,3,7,8-TCDF	%		79.4	96.9 / 94.9	103	84.6

# Quality Control PFE1917

## ORG-025\_DIOXIN | Dioxins/Furans (Soil) | Batch BFE5524

Analyte	Units	PQL	Blank	DUP1	LCS %	Spike %
				BFE5524-DUP1# Samp   QC   RPD %		
2,3,7,8-TCDF WHO-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,7,8-TCDF WHO-TEQ2	pg/g		0.0250	0.0250   0.0250   0.00	[NA]	[NA]
2,3,7,8-TCDF WHO-TEQ3	pg/g		0.0500	0.0500   0.0500   0.00	[NA]	[NA]
2,3,7,8-TCDF I-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,7,8-TCDF I-TEQ2	pg/g		0.0250	0.0250   0.0250   0.00	[NA]	[NA]
2,3,7,8-TCDF I-TEQ3	pg/g		0.0500	0.0500   0.0500   0.00	[NA]	[NA]
1,2,3,7,8-PeCDF	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	94.4	84.3
<i>Surrogate 13C-1,2,3,7,8-PeCDF</i>	%		76.8	83.7 / 85.4	108	91.1
1,2,3,7,8-PeCDF WHO-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8-PeCDF WHO-TEQ2	pg/g		0.0300	0.0300   0.0300   0.00	[NA]	[NA]
1,2,3,7,8-PeCDF WHO-TEQ3	pg/g		0.0600	0.0600   0.0600   0.00	[NA]	[NA]
1,2,3,7,8-PeCDF I-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8-PeCDF I-TEQ2	pg/g		0.0500	0.0500   0.0500   0.00	[NA]	[NA]
1,2,3,7,8-PeCDF I-TEQ3	pg/g		0.100	0.100   0.100   0.00	[NA]	[NA]
2,3,4,7,8-PeCDF	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	84.5	86.8
<i>Surrogate 13C-2,3,4,7,8-PeCDF</i>	%		79.9	90.3 / 89.7	118	94.1
2,3,4,7,8-PeCDF WHO-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,4,7,8-PeCDF WHO-TEQ2	pg/g		0.300	0.300   0.300   0.00	[NA]	[NA]
2,3,4,7,8-PeCDF WHO-TEQ3	pg/g		0.600	0.600   0.600   0.00	[NA]	[NA]
2,3,4,7,8-PeCDF I-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,4,7,8-PeCDF I-TEQ2	pg/g		0.500	0.500   0.500   0.00	[NA]	[NA]
2,3,4,7,8-PeCDF I-TEQ3	pg/g		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDF	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	91.3	90.1
<i>Surrogate 13C-1,2,3,4,7,8-HxCDF</i>	%		83.1	80.9 / 84.0	108	93.2
1,2,3,4,7,8-HxCDF WHO-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8-HxCDF WHO-TEQ2	pg/g		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDF WHO-TEQ3	pg/g		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDF I-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8-HxCDF I-TEQ2	pg/g		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDF I-TEQ3	pg/g		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDF	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	90.8	85.9
<i>Surrogate 13C-1,2,3,6,7,8-HxCDF</i>	%		76.1	86.5 / 82.4	114	96.0
1,2,3,6,7,8-HxCDF WHO-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,6,7,8-HxCDF WHO-TEQ2	pg/g		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDF WHO-TEQ3	pg/g		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDF I-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,6,7,8-HxCDF I-TEQ2	pg/g		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDF I-TEQ3	pg/g		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDF	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	98.1	93.0
<i>Surrogate 13C-1,2,3,7,8,9-HxCDF</i>	%		71.4	81.7 / 79.3	107	91.5
1,2,3,7,8,9-HxCDF WHO-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8,9-HxCDF WHO-TEQ2	pg/g		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDF WHO-TEQ3	pg/g		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDF I-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8,9-HxCDF I-TEQ2	pg/g		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDF I-TEQ3	pg/g		0.200	0.200   0.200   0.00	[NA]	[NA]
2,3,4,6,7,8-HxCDF	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	96.7	95.9
<i>Surrogate 13C-2,3,4,6,7,8-HxCDF</i>	%		80.8	96.9 / 80.1	104	85.7
2,3,4,6,7,8-HxCDF WHO-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,4,6,7,8-HxCDF WHO-TEQ2	pg/g		0.100	0.100   0.100   0.00	[NA]	[NA]
2,3,4,6,7,8-HxCDF WHO-TEQ3	pg/g		0.200	0.200   0.200   0.00	[NA]	[NA]
2,3,4,6,7,8-HxCDF I-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,4,6,7,8-HxCDF I-TEQ2	pg/g		0.100	0.100   0.100   0.00	[NA]	[NA]
2,3,4,6,7,8-HxCDF I-TEQ3	pg/g		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HxCDF	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	96.8	96.7
<i>Surrogate 13C-1,2,3,4,6,7,8-HxCDF</i>	%		78.2	75.9 / 77.5	105	86.3
1,2,3,4,6,7,8-HxCDF WHO-TEQ1	pg/g		0.00	0.560   0.403   32.6	[NA]	[NA]

# Quality Control PFE1917

## ORG-025\_DIOXIN | Dioxins/Furans (Soil) | Batch BFE5524

Analyte	Units	PQL	Blank	DUP1	LCS %	Spike %
				BFE5524-DUP1# Samp   QC   RPD %		
1,2,3,4,6,7,8-HpCDF WHO-TEQ2	pg/g		0.0100	0.560   0.403   32.6	[NA]	[NA]
1,2,3,4,6,7,8-HpCDF WHO-TEQ3	pg/g		0.0200	0.560   0.403   32.6	[NA]	[NA]
1,2,3,4,6,7,8-HpCDF I-TEQ1	pg/g		0.00	0.560   0.403   32.6	[NA]	[NA]
1,2,3,4,6,7,8-HpCDF I-TEQ2	pg/g		0.0100	0.560   0.403   32.6	[NA]	[NA]
1,2,3,4,6,7,8-HpCDF I-TEQ3	pg/g		0.0200	0.560   0.403   32.6	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF	pg/g	2.0	<2.0	<2.0   <2.0   [NA]	98.9	89.5
<i>Surrogate 13C-1,2,3,4,7,8,9-HpCDF</i>	%		77.3	77.3 / 78.6	104	87.9
1,2,3,4,7,8,9-HpCDF WHO-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF WHO-TEQ2	pg/g		0.0100	0.0100   0.0100   0.00	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF WHO-TEQ3	pg/g		0.0200	0.0200   0.0200   0.00	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF I-TEQ1	pg/g		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF I-TEQ2	pg/g		0.0100	0.0100   0.0100   0.00	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF I-TEQ3	pg/g		0.0200	0.0200   0.0200   0.00	[NA]	[NA]
OCDF	pg/g	5.0	<5.0	17.7   13.7   25.2	97.3	88.7
OCDF WHO-TEQ1	pg/g		0.00	0.00530   0.00411   25.2	[NA]	[NA]
OCDF WHO-TEQ2	pg/g		0.000750	0.00530   0.00411   25.2	[NA]	[NA]
OCDF WHO-TEQ3	pg/g		0.00150	0.00530   0.00411   25.2	[NA]	[NA]
OCDF I-TEQ1	pg/g		0.00	0.0177   0.0137   25.2	[NA]	[NA]
OCDF I-TEQ2	pg/g		0.00250	0.0177   0.0137   25.2	[NA]	[NA]
OCDF I-TEQ3	pg/g		0.00500	0.0177   0.0137   25.2	[NA]	[NA]

# The QC reported was not specifically part of this workorder but formed part of the QC process batch.

# Quality Control PFE1917

## ORG-025\_DIOXIN | Dioxins/Furans (Swab) | Batch BFF0424

Analyte	Units	PQL	Blank	LCS %
2,3,7,8-TCDD	pg/sample	10	<10	90.5
Surrogate 13C-2,3,7,8-TCDD	%		105	121
1,2,3,7,8-PeCDD	pg/sample	50	<50	84.1
Surrogate 13C-1,2,3,7,8-PeCDD	%		99.5	107
1,2,3,4,7,8-HxCDD	pg/sample	50	<50	114
Surrogate 13C-1,2,3,4,7,8-HxCDD	%		99.5	106
1,2,3,6,7,8-HxCDD	pg/sample	50	<50	83.2
Surrogate 13C-1,2,3,6,7,8-HxCDD	%		97.5	119
1,2,3,7,8,9-HxCDD	pg/sample	50	<50	81.7
1,2,3,4,6,7,8-HpCDD	pg/sample	50	<50	80.8
Surrogate 13C-1,2,3,4,6,7,8-HpCDD	%		102	107
OCDD	pg/sample	100	<100	82.0
Surrogate 13C-OCDD	%		96.6	120
2,3,7,8-TCDF	pg/sample	10	<10	88.8
Surrogate 13C-2,3,7,8-TCDF	%		106	113
1,2,3,7,8-PeCDF	pg/sample	50	<50	83.8
Surrogate 13C-1,2,3,7,8-PeCDF	%		92.3	107
2,3,4,7,8-PeCDF	pg/sample	50	<50	93.6
Surrogate 13C-2,3,4,7,8-PeCDF	%		105	123
1,2,3,4,7,8-HxCDF	pg/sample	50	<50	83.0
Surrogate 13C-1,2,3,4,7,8-HxCDF	%		92.7	114
1,2,3,6,7,8-HxCDF	pg/sample	50	<50	78.2
Surrogate 13C-1,2,3,6,7,8-HxCDF	%		97.5	131
1,2,3,7,8,9-HxCDF	pg/sample	50	<50	94.7
Surrogate 13C-1,2,3,7,8,9-HxCDF	%		101	120
2,3,4,6,7,8-HxCDF	pg/sample	50	<50	88.8
Surrogate 13C-2,3,4,6,7,8-HxCDF	%		97.7	123
1,2,3,4,6,7,8-HpCDF	pg/sample	50	<50	78.3
Surrogate 13C-1,2,3,4,6,7,8-HpCDF	%		90.8	116
1,2,3,4,7,8,9-HpCDF	pg/sample	50	<50	74.7
Surrogate 13C-1,2,3,4,7,8,9-HpCDF	%		93.2	123
OCDF	pg/sample	100	<100	79.3

## INORG-008 | Inorganics - Moisture (Soil) | Batch BFF0905

Analyte	Units	PQL	Blank	DUP1 BFF0905-DUP1# Samp   QC   RPD %	LCS %
Moisture	%	0.1		10.3   10.3   0.00	[NA]
% Solids	%	0.1		89.7   89.7   0.00	[NA]

# The QC reported was not specifically part of this workorder but formed part of the QC process batch.

## QC Comments

Identifier	Description
[1]	Spike recovery is not applicable due to the relatively high analyte background in the sample (>3* spike level). However, the LCS recovery is within acceptance criteria.



## CHAIN OF CUSTODY FORM - Client

[Copyright and Confidential]

Client: Reditus Consulting Contact Person: Toby Scrivener Project Mgr: Toby Scrivener Sampler: Renee Ashton Address: Unit 1A, 29-33 Waratah St, Kirrawee NSW 2232 Phone: (02) 9521 6567 Email: tobyscrivener@reditus.com.au reneashton@reditus.com.au deniseadean@reditus.com.au	Client Project Name/Number/Site etc (ie report title): Detailed Site Investigation: 25-27 Leeds St, Rhodes NSW PO No.: 22148 Envirolab Quote No.: Date results required: Or choose: standard / same day / 1 day / 2 day / 3 day Note: Inform lab in advance if urgent turnaround is required - surcharges apply Mob: 0477 882 907 Additional report format: esdat / equis / Lab Comments:
--	--

### ENVIROLAB GROUP

National phone number 1300 424 344

Sydney Lab - Envirolab Services  
12 Asnley St, Chatswood, NSW 2067  
02 9910 6200 | [sydney@envirolab.com.au](mailto:sydney@envirolab.com.au)

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08 9317 2505 | [lab@mpl.com.au](mailto:lab@mpl.com.au)

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Darwin Office - Envirolab Services  
Unit 7, 17 Willes Rd, Berrimah, NT 0820

Envirolab Sample ID	Client Sample ID or Information	Depth	Date sampled	Type of sample	Tests Required							Comments	
					Combo 6a	Dioxins & Furans	PFAS	Asbestos (%w/w)	Combo 3	TRH, BTEX, PAH, Metals	TRH, BTEX	NEPA/Eils	Chromium Reducible Sulphur (CRS)
1	BH102	0.5	21/05/2024	S	X	X	X	X					
2	BH102	2	21/05/2024	S					X				
3	BH102	4	21/05/2024	S									X
4	BH102	6	21/05/2024	S									X
5	BH103	1	22/05/2024	S	X	X	X	X					
6	BH103	3	22/05/2024	S					X				
7	BH103	4	22/05/2024	S					X				X
8	BH103	6	22/05/2024	S									X
9	BH103	7	22/05/2024	S									X
10	BH105	0.5	21/05/2024	S	X	X	X	X					
11	BH105	3	21/05/2024	S					X				
12	BH105	4	21/05/2024	S									X
13	BH106	0.5	22/05/2024	S	X	X	X	X					
14	BH106	2	22/05/2024	S					X				X
15	BH106	3	22/05/2024	S									X
16	BH106	6	22/05/2024	S									X
17	BH107	0.5	21/05/2024	S	X	X	X	X					
18	BH107	3	21/05/2024	S					X				
19	BH107	4	21/05/2024	S									X
20	BH108	0.3	23/05/2024	S	X			X					
21	BH109	0.3	20/05/2024	S	X	X	X	X					
22	BH109	1	20/05/2024	S					X				
23	BH109	2	20/05/2024	S									X
24	BH110	0.25	20/05/2024	S	X		X	X					
25	BH110	1	20/05/2024	S					X				
26	BH111	0.5	23/05/2024	S	X		X	X					
27	BH111	2	23/05/2024	S					X				
28	BH112	0.3	20/05/2024	S	X		X	X					
29	BH112	1	20/05/2024	S					X				
30	BH113	0.5	20/05/2024	S	X			X					
31	BH113	2	20/05/2024	S					X				
32	BH114	0.5	20/05/2024	S	X		X	X					
33	BH114	1	20/05/2024	S					X				X
34	BH115	0.2	23/05/2024	S	X			X					
35	BH115	1	23/05/2024	S					X				
36	BH115	5.1	23/05/2024	S									X
37	BH115	10.7	23/05/2024	S									X
38	DUP02	2	21/05/2024	S					X				
39	TRIP02	2	21/05/2024	S									Pts fwd to ALS
40	DUP03	3	21/05/2024	S					X				
41	TRIP03	3	21/05/2024	S					X				Pts fwd to ALS
	Trip Blank			S									X
	Trip Spike			S									X

Please tick the box if observed settled sediment present in water samples is to be included in the extraction and/or analysis

Released by (Company): Reditus Print Name: Renee Ashton Date & Time: 24/05/2024 Signature:	Received by (Company): TJS SYD Print Name: Sharron Tok Date & Time: 24/05/2024 1620 Signature: 27
Lab Use Only	
Job number: 332254 Cooling: Ice / Ice pack / None Temperature: 10 Security seal intact / Broken / None TAT Req - SAME day / 1 / 2 / 3 / 4 / STD	

Redistributed by our ELS SYN  
Chris C 27/5/24 1035

Thank you  
27/5/24 1600

Environmental Division  
Sydney  
Work Order Reference  
**ES2417505**



Telephone: +61 2 8784 8555



## CERTIFICATE OF ANALYSIS

Work Order	: ES2417505	Page	: 1 of 6
Client	: REDITUS CONSULTING PTY LTD.	Laboratory	: Environmental Division Sydney
Contact	: Toby Scrivener	Contact	: Customer Services ES
Address	: 1A/29-33 Waratah Street KIRRABEE	Address	: 277-289 Woodpark Road Smithfield NSW Australia 2164
Telephone	: ----	Telephone	: +61-2-8784 8555
Project	: Detailed Site Investigation: 25-27 Leeds St, Rhodes NSW	Date Samples Received	: 27-May-2024 16:00
Order number	: 22148	Date Analysis Commenced	: 29-May-2024
C-O-C number	: ----	Issue Date	: 03-Jun-2024 15:31
Sampler	: renee Ashton		
Site	:		
Quote number	: EN/333		
No. of samples received	: 2		
No. of samples analysed	: 2		

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

### Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories	Position	Accreditation Category
Ankit Joshi	Senior Chemist - Inorganics	Sydney Inorganics, Smithfield, NSW
Edwandy Fadjar	Organic Coordinator	Sydney Organics, Smithfield, NSW
Sanjeshni Jyoti	Senior Chemist Volatiles	Sydney Organics, Smithfield, NSW



Accreditation No. 825  
Accredited for compliance with  
ISO/IEC 17025 - Testing



## General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key : CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

^ = This result is computed from individual analyte detections at or above the level of reporting

Ø = ALS is not NATA accredited for these tests.

~ = Indicates an estimated value.

- Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a,h)anthracene (1.0), Benzo(g,h,i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero, for 'TEQ 1/2LOR' are treated as half the reported LOR, and for 'TEQ LOR' are treated as being equal to the reported LOR. Note: TEQ 1/2LOR and TEQ LOR will calculate as 0.6mg/Kg and 1.2mg/Kg respectively for samples with non-detects for all of the eight TEQ PAHs.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.



## Analytical Results

Sub-Matrix: SOIL (Matrix: SOIL)				Sample ID	TRIP02	TRIP03	---	---	---			
				Sampling date / time	21-May-2024 00:00	21-May-2024 00:00	---	---	---			
Compound	CAS Number	LOR	Unit	ES2417505-001	ES2417505-002	-----	-----	-----	-----			
				Result	Result	---	---	---	---			
<b>EA055: Moisture Content (Dried @ 105-110°C)</b>												
Moisture Content				1.0	%	12.2	14.3	---	---			
<b>EG005(ED093)T: Total Metals by ICP-AES</b>												
Arsenic	7440-38-2	5	mg/kg	<5	<5	---	---	---	---			
Cadmium	7440-43-9	1	mg/kg	<1	<1	---	---	---	---			
Chromium	7440-47-3	2	mg/kg	8	8	---	---	---	---			
Copper	7440-50-8	5	mg/kg	<5	17	---	---	---	---			
Lead	7439-92-1	5	mg/kg	18	46	---	---	---	---			
Nickel	7440-02-0	2	mg/kg	<2	6	---	---	---	---			
Zinc	7440-66-6	5	mg/kg	<5	58	---	---	---	---			
<b>EG035T: Total Recoverable Mercury by FIMS</b>												
Mercury	7439-97-6	0.1	mg/kg	<0.1	<0.1	---	---	---	---			
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons</b>												
Naphthalene	91-20-3	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Acenaphthylene	208-96-8	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Acenaphthene	83-32-9	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Fluorene	86-73-7	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Phenanthrene	85-01-8	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Anthracene	120-12-7	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Fluoranthene	206-44-0	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Pyrene	129-00-0	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Benz(a)anthracene	56-55-3	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Chrysene	218-01-9	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Benzo(b+j)fluoranthene	205-99-2 205-82-3	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Benzo(k)fluoranthene	207-08-9	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Benzo(a)pyrene	50-32-8	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Indeno(1,2,3.cd)pyrene	193-39-5	0.5	mg/kg	<0.5	<0.5	---	---	---	---			
Dibenz(a,h)anthracene	53-70-3	0.5	mg/kg	<0.5	<0.5	---	---	---	---			



## Analytical Results

Sub-Matrix: SOIL (Matrix: SOIL)		Sample ID	TRIP02	TRIP03	---	---	---
		Sampling date / time	21-May-2024 00:00	21-May-2024 00:00	---	---	---
Compound	CAS Number	LOR	Unit	ES2417505-001	ES2417505-002	-----	-----
			Result	Result	---	---	---
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons - Continued</b>							
Benzo(g.h.i)perylene	191-24-2	0.5	mg/kg	<0.5	<0.5	---	---
^ Sum of polycyclic aromatic hydrocarbons	---	0.5	mg/kg	<0.5	<0.5	---	---
^ Benzo(a)pyrene TEQ (zero)	---	0.5	mg/kg	<0.5	<0.5	---	---
^ Benzo(a)pyrene TEQ (half LOR)	---	0.5	mg/kg	0.6	0.6	---	---
^ Benzo(a)pyrene TEQ (LOR)	---	0.5	mg/kg	1.2	1.2	---	---
<b>EP080/071: Total Petroleum Hydrocarbons</b>							
C6 - C9 Fraction	---	10	mg/kg	<10	<10	---	---
C10 - C14 Fraction	---	50	mg/kg	<50	<50	---	---
C15 - C28 Fraction	---	100	mg/kg	<100	<100	---	---
C29 - C36 Fraction	---	100	mg/kg	<100	<100	---	---
^ C10 - C36 Fraction (sum)	---	50	mg/kg	<50	<50	---	---
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions</b>							
C6 - C10 Fraction	C6_C10	10	mg/kg	<10	<10	---	---
^ C6 - C10 Fraction minus BTEX (F1)	C6_C10-BTEX	10	mg/kg	<10	<10	---	---
>C10 - C16 Fraction	---	50	mg/kg	<50	<50	---	---
>C16 - C34 Fraction	---	100	mg/kg	<100	<100	---	---
>C34 - C40 Fraction	---	100	mg/kg	<100	<100	---	---
^ >C10 - C40 Fraction (sum)	---	50	mg/kg	<50	<50	---	---
^ >C10 - C16 Fraction minus Naphthalene (F2)	---	50	mg/kg	<50	<50	---	---
<b>EP080: BTEXN</b>							
Benzene	71-43-2	0.2	mg/kg	<0.2	<0.2	---	---
Toluene	108-88-3	0.5	mg/kg	<0.5	<0.5	---	---
Ethylbenzene	100-41-4	0.5	mg/kg	<0.5	<0.5	---	---
meta- & para-Xylene	108-38-3 106-42-3	0.5	mg/kg	<0.5	<0.5	---	---
ortho-Xylene	95-47-6	0.5	mg/kg	<0.5	<0.5	---	---
^ Sum of BTEX	---	0.2	mg/kg	<0.2	<0.2	---	---



## Analytical Results

Sub-Matrix: SOIL (Matrix: SOIL)		Sample ID	TRIP02	TRIP03	---	---	---
		Sampling date / time	21-May-2024 00:00	21-May-2024 00:00	---	---	---
Compound	CAS Number	LOR	Unit	ES2417505-001	ES2417505-002	-----	-----
			Result	Result		---	---
<b>EP080: BTEXN - Continued</b>							
^ Total Xylenes	---	0.5	mg/kg	<0.5	<0.5	---	---
Naphthalene	91-20-3	1	mg/kg	<1	<1	---	---
<b>EP075(SIM)S: Phenolic Compound Surrogates</b>							
Phenol-d6	13127-88-3	0.5	%	89.6	87.5	---	---
2-Chlorophenol-D4	93951-73-6	0.5	%	99.1	85.6	---	---
2,4,6-Tribromophenol	118-79-6	0.5	%	84.1	76.8	---	---
<b>EP075(SIM)T: PAH Surrogates</b>							
2-Fluorobiphenyl	321-60-8	0.5	%	92.7	85.4	---	---
Anthracene-d10	1719-06-8	0.5	%	95.3	89.5	---	---
4-Terphenyl-d14	1718-51-0	0.5	%	103	95.0	---	---
<b>EP080S: TPH(V)/BTEX Surrogates</b>							
1,2-Dichloroethane-D4	17060-07-0	0.2	%	79.8	79.5	---	---
Toluene-D8	2037-26-5	0.2	%	75.1	80.6	---	---
4-Bromofluorobenzene	460-00-4	0.2	%	89.0	93.1	---	---



## Surrogate Control Limits

Sub-Matrix: SOIL		Recovery Limits (%)	
Compound	CAS Number	Low	High
<b>EP075(SIM)S: Phenolic Compound Surrogates</b>			
Phenol-d6	13127-88-3	63	123
2-Chlorophenol-D4	93951-73-6	66	122
2,4,6-Tribromophenol	118-79-6	40	138
<b>EP075(SIM)T: PAH Surrogates</b>			
2-Fluorobiphenyl	321-60-8	70	122
Anthracene-d10	1719-06-8	66	128
4-Terphenyl-d14	1718-51-0	65	129
<b>EP080S: TPH(V)/BTEX Surrogates</b>			
1,2-Dichloroethane-D4	17060-07-0	63	125
Toluene-D8	2037-26-5	67	124
4-Bromofluorobenzene	460-00-4	66	131



## QA/QC Compliance Assessment to assist with Quality Review

Work Order	: ES2417505	Page	: 1 of 4
Client	: REDITUS CONSULTING PTY LTD.	Laboratory	: Environmental Division Sydney
Contact	: Toby Scrivener	Telephone	: +61-2-8784 8555
Project	: Detailed Site Investigation: 25-27 Leeds St, Rhodes NSW	Date Samples Received	: 27-May-2024
Site	:	Issue Date	: 03-Jun-2024
Sampler	: renee Ashton	No. of samples received	: 2
Order number	: 22148	No. of samples analysed	: 2

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

### ***Summary of Outliers***

#### ***Outliers : Quality Control Samples***

This report highlights outliers flagged in the Quality Control (QC) Report.

- **NO** Method Blank value outliers occur.
- **NO** Duplicate outliers occur.
- **NO** Laboratory Control outliers occur.
- **NO** Matrix Spike outliers occur.
- For all regular sample matrices, where applicable to the methodology, **NO** surrogate recovery outliers occur.

#### ***Outliers : Analysis Holding Time Compliance***

- **NO** Analysis Holding Time Outliers exist.

#### ***Outliers : Frequency of Quality Control Samples***

- **NO** Quality Control Sample Frequency Outliers exist.

## Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for VOC in soils vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive or Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: SOIL

Evaluation: ✗ = Holding time breach ; ✓ = Within holding time.

Method	Container / Client Sample ID(s)	Sample Date	Extraction / Preparation			Analysis		
			Date extracted	Due for extraction	Evaluation	Date analysed	Due for analysis	Evaluation
<b>EA055: Moisture Content (Dried @ 105-110°C)</b>								
Soil Glass Jar - Unpreserved (EA055) TRIP02,	TRIP03	21-May-2024	---	---	---	30-May-2024	04-Jun-2024	✓
<b>EG005(ED093)T: Total Metals by ICP-AES</b>								
Soil Glass Jar - Unpreserved (EG005T) TRIP02,	TRIP03	21-May-2024	30-May-2024	18-Nov-2024	✓	31-May-2024	18-Nov-2024	✓
<b>EG035T: Total Recoverable Mercury by FIMS</b>								
Soil Glass Jar - Unpreserved (EG035T) TRIP02,	TRIP03	21-May-2024	30-May-2024	18-Jun-2024	✓	03-Jun-2024	18-Jun-2024	✓
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons</b>								
Soil Glass Jar - Unpreserved (EP075(SIM)) TRIP02,	TRIP03	21-May-2024	30-May-2024	04-Jun-2024	✓	31-May-2024	09-Jul-2024	✓
<b>EP080/071: Total Petroleum Hydrocarbons</b>								
Soil Glass Jar - Unpreserved (EP080) TRIP02,	TRIP03	21-May-2024	29-May-2024	04-Jun-2024	✓	29-May-2024	04-Jun-2024	✓
Soil Glass Jar - Unpreserved (EP071) TRIP02,	TRIP03	21-May-2024	30-May-2024	04-Jun-2024	✓	31-May-2024	09-Jul-2024	✓
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions</b>								
Soil Glass Jar - Unpreserved (EP080) TRIP02,	TRIP03	21-May-2024	29-May-2024	04-Jun-2024	✓	29-May-2024	04-Jun-2024	✓
Soil Glass Jar - Unpreserved (EP071) TRIP02,	TRIP03	21-May-2024	30-May-2024	04-Jun-2024	✓	31-May-2024	09-Jul-2024	✓
<b>EP080: BTEX</b>								
Soil Glass Jar - Unpreserved (EP080) TRIP02,	TRIP03	21-May-2024	29-May-2024	04-Jun-2024	✓	29-May-2024	04-Jun-2024	✓



## Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

Matrix: SOIL

Evaluation: ✗ = Quality Control frequency not within specification ; ✓ = Quality Control frequency within specification.

Quality Control Sample Type	Analytical Methods	Method	Count		Rate (%)		Quality Control Specification
			QC	Regular	Actual	Expected	
<b>Laboratory Duplicates (DUP)</b>							
Moisture Content		EA055	2	20	10.00	10.00	✓ NEPM 2013 B3 & ALS QC Standard
PAH/Phenols (SIM)		EP075(SIM)	2	17	11.76	10.00	✓ NEPM 2013 B3 & ALS QC Standard
Total Mercury by FIMS		EG035T	2	20	10.00	10.00	✓ NEPM 2013 B3 & ALS QC Standard
Total Metals by ICP-AES		EG005T	2	20	10.00	10.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction		EP071	2	20	10.00	10.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH Volatiles/BTEX		EP080	2	12	16.67	10.00	✓ NEPM 2013 B3 & ALS QC Standard
<b>Laboratory Control Samples (LCS)</b>							
PAH/Phenols (SIM)		EP075(SIM)	1	17	5.88	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Total Mercury by FIMS		EG035T	1	20	5.00	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Total Metals by ICP-AES		EG005T	1	20	5.00	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction		EP071	1	20	5.00	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH Volatiles/BTEX		EP080	1	12	8.33	5.00	✓ NEPM 2013 B3 & ALS QC Standard
<b>Method Blanks (MB)</b>							
PAH/Phenols (SIM)		EP075(SIM)	1	17	5.88	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Total Mercury by FIMS		EG035T	1	20	5.00	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Total Metals by ICP-AES		EG005T	1	20	5.00	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction		EP071	1	20	5.00	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH Volatiles/BTEX		EP080	1	12	8.33	5.00	✓ NEPM 2013 B3 & ALS QC Standard
<b>Matrix Spikes (MS)</b>							
PAH/Phenols (SIM)		EP075(SIM)	1	17	5.88	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Total Mercury by FIMS		EG035T	1	20	5.00	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Total Metals by ICP-AES		EG005T	1	20	5.00	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction		EP071	1	20	5.00	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH Volatiles/BTEX		EP080	1	12	8.33	5.00	✓ NEPM 2013 B3 & ALS QC Standard

## Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

<b>Analytical Methods</b>	<b>Method</b>	<b>Matrix</b>	<b>Method Descriptions</b>
Moisture Content	EA055	SOIL	In house: A gravimetric procedure based on weight loss over a 12 hour drying period at 105-110 degrees C. This method is compliant with NEPM Schedule B(3).
Total Metals by ICP-AES	EG005T	SOIL	In house: Referenced to APHA 3120; USEPA SW 846 - 6010. Metals are determined following an appropriate acid digestion of the soil. The ICPAES technique ionises samples in a plasma, emitting a characteristic spectrum based on metals present. Intensities at selected wavelengths are compared against those of matrix matched standards. This method is compliant with NEPM Schedule B(3)
Total Mercury by FIMS	EG035T	SOIL	In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl <sub>2</sub> ) (Cold Vapour generation) AAS) FIM-AAS is an automated flameless atomic absorption technique. Mercury in solids are determined following an appropriate acid digestion. Ionic mercury is reduced online to atomic mercury vapour by SnCl <sub>2</sub> which is then purged into a heated quartz cell. Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM Schedule B(3)
TRH - Semivolatile Fraction	EP071	SOIL	In house: Referenced to USEPA SW 846 - 8015 Sample extracts are analysed by Capillary GC/FID and quantified against alkane standards over the range C10 - C40. Compliant with NEPM Schedule B(3).
PAH/Phenols (SIM)	EP075(SIM)	SOIL	In house: Referenced to USEPA SW 846 - 8270. Extracts are analysed by Capillary GC/MS in Selective Ion Mode (SIM) and quantification is by comparison against an established 5 point calibration curve. This method is compliant with NEPM Schedule B(3)
TRH Volatiles/BTEX	EP080	SOIL	In house: Referenced to USEPA SW 846 - 8260. Extracts are analysed by Purge and Trap, Capillary GC/MS. Quantification is by comparison against an established 5 point calibration curve. Compliant with NEPM Schedule B(3) amended.

<b>Preparation Methods</b>	<b>Method</b>	<b>Matrix</b>	<b>Method Descriptions</b>
Hot Block Digest for metals in soils sediments and sludges	EN69	SOIL	In house: Referenced to USEPA 200.2. Hot Block Acid Digestion 1.0g of sample is heated with Nitric and Hydrochloric acids, then cooled. Peroxide is added and samples heated and cooled again before being filtered and bulked to volume for analysis. Digest is appropriate for determination of selected metals in sludge, sediments, and soils. This method is compliant with NEPM Schedule B(3).
Methanolic Extraction of Soils for Purge and Trap	ORG16	SOIL	In house: Referenced to USEPA SW 846 - 5030A. 5g of solid is shaken with surrogate and 10mL methanol prior to analysis by Purge and Trap - GC/MS.
Tumbler Extraction of Solids	ORG17	SOIL	In house: Mechanical agitation (tumbler). 10g of sample, Na <sub>2</sub> SO <sub>4</sub> and surrogate are extracted with 30mL 1:1 DCM/Acetone by end over end tumble. The solvent is decanted, dehydrated and concentrated (by KD) to the desired volume for analysis.



## QUALITY CONTROL REPORT

Work Order	: ES2417505	Page	: 1 of 8
Client	: REDITUS CONSULTING PTY LTD.	Laboratory	: Environmental Division Sydney
Contact	: Toby Scrivener	Contact	: Customer Services ES
Address	: 1A/29-33 Waratah Street KIRRRAWEE	Address	: 277-289 Woodpark Road Smithfield NSW Australia 2164
Telephone	: ----	Telephone	: +61-2-8784 8555
Project	: Detailed Site Investigation: 25-27 Leeds St, Rhodes NSW	Date Samples Received	: 27-May-2024
Order number	: 22148	Date Analysis Commenced	: 29-May-2024
C-O-C number	: ----	Issue Date	: 03-Jun-2024
Sampler	: renee Ashton		
Site	:		
Quote number	: EN/333		
No. of samples received	: 2		
No. of samples analysed	: 2		



Accreditation No. 825  
Accredited for compliance with  
ISO/IEC 17025 - Testing

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Quality Control Report contains the following information:

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

### Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories	Position	Accreditation Category
Ankit Joshi	Senior Chemist - Inorganics	Sydney Inorganics, Smithfield, NSW
Edwandy Fadjar	Organic Coordinator	Sydney Organics, Smithfield, NSW
Sanjeshni Jyoti	Senior Chemist Volatiles	Sydney Organics, Smithfield, NSW

## General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key : Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

# = Indicates failed QC

\* = The final LOR has been raised due to dilution or other sample specific cause; adjusted LOR is shown in brackets. The duplicate ranges for Acceptable RPD% are applied to the final LOR where applicable.

## Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit; Result between 10 and 20 times LOR: 0% - 50%; Result > 20 times LOR: 0% - 20%.

Sub-Matrix: SOIL

Laboratory Duplicate (DUP) Report									
Laboratory sample ID	Sample ID	Method: Compound	CAS Number	LOR	Unit	Original Result	Duplicate Result	RPD (%)	Acceptable RPD (%)
<b>EG005(ED093)T: Total Metals by ICP-AES (QC Lot: 5825491)</b>									
ES2417303-001	Anonymous	EG005T: Cadmium	7440-43-9	1	mg/kg	<1	<1	0.0	No Limit
		EG005T: Chromium	7440-47-3	2	mg/kg	21	18	11.2	0% - 50%
		EG005T: Nickel	7440-02-0	2	mg/kg	13	11	12.6	No Limit
		EG005T: Arsenic	7440-38-2	5	mg/kg	<5	<5	0.0	No Limit
		EG005T: Copper	7440-50-8	5	mg/kg	24	18	28.9	No Limit
		EG005T: Lead	7439-92-1	5	mg/kg	6	6	0.0	No Limit
		EG005T: Zinc	7440-66-6	5	mg/kg	62	57	8.2	0% - 50%
ES2417501-001	Anonymous	EG005T: Cadmium	7440-43-9	1	mg/kg	<1	<1	0.0	No Limit
		EG005T: Chromium	7440-47-3	2	mg/kg	19	16	17.1	No Limit
		EG005T: Nickel	7440-02-0	2	mg/kg	8	6	18.1	No Limit
		EG005T: Arsenic	7440-38-2	5	mg/kg	5	<5	0.0	No Limit
		EG005T: Copper	7440-50-8	5	mg/kg	14	12	17.3	No Limit
		EG005T: Lead	7439-92-1	5	mg/kg	37	21	55.2	No Limit
		EG005T: Zinc	7440-66-6	5	mg/kg	45	39	14.4	No Limit
<b>EA055: Moisture Content (Dried @ 105-110°C) (QC Lot: 5825496)</b>									
ES2417314-001	Anonymous	EA055: Moisture Content	---	0.1	%	14.1	14.7	4.2	0% - 20%
ES2417501-001	Anonymous	EA055: Moisture Content	---	0.1 (1.0)*	%	10.4	10.3	1.5	0% - 50%
<b>EG035T: Total Recoverable Mercury by FIMS (QC Lot: 5825492)</b>									
ES2417303-001	Anonymous	EG035T: Mercury	7439-97-6	0.1	mg/kg	<0.1	<0.1	0.0	No Limit
ES2417501-001	Anonymous	EG035T: Mercury	7439-97-6	0.1	mg/kg	<0.1	<0.1	0.0	No Limit



Sub-Matrix: SOIL			Laboratory Duplicate (DUP) Report						
Laboratory sample ID	Sample ID	Method: Compound	CAS Number	LOR	Unit	Original Result	Duplicate Result	RPD (%)	Acceptable RPD (%)
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QC Lot: 5821839)</b>									
ES2417493-001	Anonymous	EP075(SIM): Naphthalene	91-20-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Acenaphthylene	208-96-8	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Acenaphthene	83-32-9	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Fluorene	86-73-7	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Phenanthrene	85-01-8	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Anthracene	120-12-7	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Fluoranthene	206-44-0	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Pyrene	129-00-0	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benz(a)anthracene	56-55-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Chrysene	218-01-9	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benzo(b+j)fluoranthene	205-99-2	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
			205-82-3						
		EP075(SIM): Benzo(k)fluoranthene	207-08-9	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benzo(a)pyrene	50-32-8	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Indeno(1,2,3,cd)pyrene	193-39-5	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Dibenz(a,h)anthracene	53-70-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benzo(g,h,i)perylene	191-24-2	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Sum of polycyclic aromatic hydrocarbons	----	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benzo(a)pyrene TEQ (zero)	----	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
ES2417448-001	Anonymous	EP075(SIM): Naphthalene	91-20-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Acenaphthylene	208-96-8	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Acenaphthene	83-32-9	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Fluorene	86-73-7	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Phenanthrene	85-01-8	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Anthracene	120-12-7	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Fluoranthene	206-44-0	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Pyrene	129-00-0	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benz(a)anthracene	56-55-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Chrysene	218-01-9	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benzo(b+j)fluoranthene	205-99-2	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
			205-82-3						
		EP075(SIM): Benzo(k)fluoranthene	207-08-9	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benzo(a)pyrene	50-32-8	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Indeno(1,2,3,cd)pyrene	193-39-5	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Dibenz(a,h)anthracene	53-70-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benzo(g,h,i)perylene	191-24-2	0.5	mg/kg	<0.5	<0.5	0.0	No Limit



Sub-Matrix: SOIL			Laboratory Duplicate (DUP) Report						
Laboratory sample ID	Sample ID	Method: Compound	CAS Number	LOR	Unit	Original Result	Duplicate Result	RPD (%)	Acceptable RPD (%)
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QC Lot: 5821839) - continued</b>									
ES2417448-001	Anonymous	EP075(SIM): Sum of polycyclic aromatic hydrocarbons	---	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP075(SIM): Benzo(a)pyrene TEQ (zero)	---	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
<b>EP080/071: Total Petroleum Hydrocarbons (QC Lot: 5821838)</b>									
ES2417493-001	Anonymous	EP071: C15 - C28 Fraction	---	100	mg/kg	<100	<100	0.0	No Limit
		EP071: C29 - C36 Fraction	---	100	mg/kg	<100	<100	0.0	No Limit
		EP071: C10 - C14 Fraction	---	50	mg/kg	<50	<50	0.0	No Limit
ES2417448-001	Anonymous	EP071: C15 - C28 Fraction	---	100	mg/kg	<100	<100	0.0	No Limit
		EP071: C29 - C36 Fraction	---	100	mg/kg	<100	<100	0.0	No Limit
		EP071: C10 - C14 Fraction	---	50	mg/kg	<50	<50	0.0	No Limit
<b>EP080/071: Total Petroleum Hydrocarbons (QC Lot: 5822800)</b>									
ES2417505-001	TRIP02	EP080: C6 - C9 Fraction	---	10	mg/kg	<10	<10	0.0	No Limit
ES2417133-002	Anonymous	EP080: C6 - C9 Fraction	---	10	mg/kg	<10	<10	0.0	No Limit
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5821838)</b>									
ES2417493-001	Anonymous	EP071: >C16 - C34 Fraction	---	100	mg/kg	<100	<100	0.0	No Limit
		EP071: >C34 - C40 Fraction	---	100	mg/kg	<100	<100	0.0	No Limit
		EP071: >C10 - C16 Fraction	---	50	mg/kg	<50	<50	0.0	No Limit
ES2417448-001	Anonymous	EP071: >C16 - C34 Fraction	---	100	mg/kg	<100	<100	0.0	No Limit
		EP071: >C34 - C40 Fraction	---	100	mg/kg	<100	<100	0.0	No Limit
		EP071: >C10 - C16 Fraction	---	50	mg/kg	<50	<50	0.0	No Limit
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5822800)</b>									
ES2417505-001	TRIP02	EP080: C6 - C10 Fraction	C6_C10	10	mg/kg	<10	<10	0.0	No Limit
ES2417133-002	Anonymous	EP080: C6 - C10 Fraction	C6_C10	10	mg/kg	<10	<10	0.0	No Limit
<b>EP080: BTEXN (QC Lot: 5822800)</b>									
ES2417505-001	TRIP02	EP080: Benzene	71-43-2	0.2	mg/kg	<0.2	<0.2	0.0	No Limit
		EP080: Toluene	108-88-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP080: Ethylbenzene	100-41-4	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP080: meta- & para-Xylene	108-38-3 106-42-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP080: ortho-Xylene	95-47-6	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP080: Naphthalene	91-20-3	1	mg/kg	<1	<1	0.0	No Limit
ES2417133-002	Anonymous	EP080: Benzene	71-43-2	0.2	mg/kg	<0.2	<0.2	0.0	No Limit
		EP080: Toluene	108-88-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP080: Ethylbenzene	100-41-4	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP080: meta- & para-Xylene	108-38-3 106-42-3	0.5	mg/kg	<0.5	<0.5	0.0	No Limit
		EP080: ortho-Xylene	95-47-6	0.5	mg/kg	<0.5	<0.5	0.0	No Limit

Page : 5 of 8  
Work Order : ES2417505  
Client : REDITUS CONSULTING PTY LTD.  
Project : Detailed Site Investigation: 25-27 Leeds St, Rhodes NSW



Sub-Matrix: SOIL			Laboratory Duplicate (DUP) Report						
Laboratory sample ID	Sample ID	Method: Compound	CAS Number	LOR	Unit	Original Result	Duplicate Result	RPD (%)	Acceptable RPD (%)
<b>EP080: BTEXN (QC Lot: 5822800) - continued</b>									
ES2417133-002	Anonymous	EP080: Naphthalene	91-20-3	1	mg/kg	<1	<1	0.0	No Limit



## Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

Sub-Matrix: SOIL

Method: Compound	CAS Number	LOR	Unit	Result	Method Blank (MB) Report	Laboratory Control Spike (LCS) Report		
					Spike Concentration	Spike Recovery (%)	Acceptable Limits (%)	
							Low	High
<b>EG005(ED093)T: Total Metals by ICP-AES (QCLot: 5825491)</b>								
EG005T: Arsenic	7440-38-2	5	mg/kg	<5	121.1 mg/kg	103	88.0	113
EG005T: Cadmium	7440-43-9	1	mg/kg	<1	0.74 mg/kg	124	70.0	130
EG005T: Chromium	7440-47-3	2	mg/kg	<2	19.6 mg/kg	119	68.0	132
EG005T: Copper	7440-50-8	5	mg/kg	<5	52.9 mg/kg	104	89.0	111
EG005T: Lead	7439-92-1	5	mg/kg	<5	60.8 mg/kg	101	82.0	119
EG005T: Nickel	7440-02-0	2	mg/kg	<2	15.3 mg/kg	104	80.0	120
EG005T: Zinc	7440-66-6	5	mg/kg	<5	139.3 mg/kg	91.5	66.0	133
<b>EG035T: Total Recoverable Mercury by FIMS (QCLot: 5825492)</b>								
EG035T: Mercury	7439-97-6	0.1	mg/kg	<0.1	0.087 mg/kg	94.8	70.0	125
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QCLot: 5821839)</b>								
EP075(SIM): Naphthalene	91-20-3	0.5	mg/kg	<0.5	6 mg/kg	91.9	77.0	125
EP075(SIM): Acenaphthylene	208-96-8	0.5	mg/kg	<0.5	6 mg/kg	93.3	72.0	124
EP075(SIM): Acenaphthene	83-32-9	0.5	mg/kg	<0.5	6 mg/kg	98.4	73.0	127
EP075(SIM): Fluorene	86-73-7	0.5	mg/kg	<0.5	6 mg/kg	89.7	72.0	126
EP075(SIM): Phenanthrene	85-01-8	0.5	mg/kg	<0.5	6 mg/kg	90.9	75.0	127
EP075(SIM): Anthracene	120-12-7	0.5	mg/kg	<0.5	6 mg/kg	93.0	77.0	127
EP075(SIM): Fluoranthene	206-44-0	0.5	mg/kg	<0.5	6 mg/kg	96.8	73.0	127
EP075(SIM): Pyrene	129-00-0	0.5	mg/kg	<0.5	6 mg/kg	94.1	74.0	128
EP075(SIM): Benz(a)anthracene	56-55-3	0.5	mg/kg	<0.5	6 mg/kg	94.7	69.0	123
EP075(SIM): Chrysene	218-01-9	0.5	mg/kg	<0.5	6 mg/kg	89.1	75.0	127
EP075(SIM): Benzo(b+j)fluoranthene	205-99-2	0.5	mg/kg	<0.5	6 mg/kg	89.7	68.0	116
	205-82-3							
EP075(SIM): Benzo(k)fluoranthene	207-08-9	0.5	mg/kg	<0.5	6 mg/kg	93.9	74.0	126
EP075(SIM): Benzo(a)pyrene	50-32-8	0.5	mg/kg	<0.5	6 mg/kg	98.6	70.0	126
EP075(SIM): Indeno(1.2.3.cd)pyrene	193-39-5	0.5	mg/kg	<0.5	6 mg/kg	79.3	61.0	121
EP075(SIM): Dibenz(a,h)anthracene	53-70-3	0.5	mg/kg	<0.5	6 mg/kg	77.3	62.0	118
EP075(SIM): Benzo(g.h.i)perylene	191-24-2	0.5	mg/kg	<0.5	6 mg/kg	79.6	63.0	121
<b>EP080/071: Total Petroleum Hydrocarbons (QCLot: 5821838)</b>								
EP071: C10 - C14 Fraction	----	50	mg/kg	<50	300 mg/kg	112	75.0	129
EP071: C15 - C28 Fraction	----	100	mg/kg	<100	450 mg/kg	113	77.0	131
EP071: C29 - C36 Fraction	----	100	mg/kg	<100	300 mg/kg	114	71.0	129



Sub-Matrix: SOIL				Method Blank (MB) Report	Laboratory Control Spike (LCS) Report			
					Spike Concentration	Spike Recovery (%)	Acceptable Limits (%)	
Method: Compound	CAS Number	LOR	Unit		Result		Low	High
<b>EP080/071: Total Petroleum Hydrocarbons (QC Lot: 5822800)</b>								
EP080: C6 - C9 Fraction	---	10	mg/kg	<10	26 mg/kg	89.5	72.2	131
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5821838)</b>								
EP071: >C10 - C16 Fraction	---	50	mg/kg	<50	375 mg/kg	99.2	77.0	125
EP071: >C16 - C34 Fraction	---	100	mg/kg	<100	525 mg/kg	110	74.0	138
EP071: >C34 - C40 Fraction	---	100	mg/kg	<100	225 mg/kg	120	63.0	131
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5822800)</b>								
EP080: C6 - C10 Fraction	C6_C10	10	mg/kg	<10	31 mg/kg	81.3	72.4	133
<b>EP080: BTEXN (QC Lot: 5822800)</b>								
EP080: Benzene	71-43-2	0.2	mg/kg	<0.2	1 mg/kg	82.3	76.0	124
EP080: Toluene	108-88-3	0.5	mg/kg	<0.5	1 mg/kg	79.2	78.5	121
EP080: Ethylbenzene	100-41-4	0.5	mg/kg	<0.5	1 mg/kg	82.4	77.4	121
EP080: meta- & para-Xylene	108-38-3 106-42-3	0.5	mg/kg	<0.5	2 mg/kg	85.0	78.2	121
EP080: ortho-Xylene	95-47-6	0.5	mg/kg	<0.5	1 mg/kg	92.9	81.3	121
EP080: Naphthalene	91-20-3	1	mg/kg	<1	1 mg/kg	92.0	78.8	122

### Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.

Sub-Matrix: SOIL				Matrix Spike (MS) Report			
				Spike	Spike Recovery (%)	Acceptable Limits (%)	
Laboratory sample ID	Sample ID	Method: Compound	CAS Number	Concentration	MS	Low	High
<b>EG005(ED093)T: Total Metals by ICP-AES (QC Lot: 5825491)</b>							
ES2417303-001	Anonymous	EG005T: Arsenic	7440-38-2	50 mg/kg	102	70.0	130
		EG005T: Cadmium	7440-43-9	50 mg/kg	93.3	70.0	130
		EG005T: Chromium	7440-47-3	50 mg/kg	94.4	68.0	132
		EG005T: Copper	7440-50-8	250 mg/kg	96.7	70.0	130
		EG005T: Lead	7439-92-1	250 mg/kg	94.5	70.0	130
		EG005T: Nickel	7440-02-0	50 mg/kg	96.3	70.0	130
		EG005T: Zinc	7440-66-6	250 mg/kg	87.1	66.0	133
<b>EG035T: Total Recoverable Mercury by FIMS (QC Lot: 5825492)</b>							
ES2417303-001	Anonymous	EG035T: Mercury	7439-97-6	5 mg/kg	97.4	70.0	130
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QC Lot: 5821839)</b>							
ES2417448-001	Anonymous	EP075(SIM): Acenaphthene	83-32-9	10 mg/kg	90.9	70.0	130
		EP075(SIM): Pyrene	129-00-0	10 mg/kg	97.7	70.0	130



Sub-Matrix: SOIL				Matrix Spike (MS) Report			
Laboratory sample ID	Sample ID	Method: Compound	CAS Number	Spike	Spike Recovery(%)	Acceptable Limits (%)	
				Concentration	MS	Low	High
<b>EP080/071: Total Petroleum Hydrocarbons (QC Lot: 5821838)</b>							
ES2417448-001	Anonymous	EP071: C10 - C14 Fraction	---	480 mg/kg	98.8	73.0	137
		EP071: C15 - C28 Fraction	---	3100 mg/kg	103	53.0	131
		EP071: C29 - C36 Fraction	---	2060 mg/kg	119	52.0	132
<b>EP080/071: Total Petroleum Hydrocarbons (QC Lot: 5822800)</b>							
ES2417133-002	Anonymous	EP080: C6 - C9 Fraction	---	32.5 mg/kg	92.1	60.4	142
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5821838)</b>							
ES2417448-001	Anonymous	EP071: >C10 - C16 Fraction	---	860 mg/kg	89.2	73.0	137
		EP071: >C16 - C34 Fraction	---	4320 mg/kg	109	53.0	131
		EP071: >C34 - C40 Fraction	---	890 mg/kg	131	52.0	132
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5822800)</b>							
ES2417133-002	Anonymous	EP080: C6 - C10 Fraction	C6_C10	37.5 mg/kg	87.0	61.1	142
<b>EP080: BTEXN (QC Lot: 5822800)</b>							
ES2417133-002	Anonymous	EP080: Benzene	71-43-2	2.5 mg/kg	87.9	62.1	122
		EP080: Toluene	108-88-3	2.5 mg/kg	89.6	66.6	119
		EP080: Ethylbenzene	100-41-4	2.5 mg/kg	94.7	67.4	123
		EP080: meta- & para-Xylene	108-38-3 106-42-3	2.5 mg/kg	99.8	66.4	121
		EP080: ortho-Xylene	95-47-6	2.5 mg/kg	110	70.7	121
		EP080: Naphthalene	91-20-3	2.5 mg/kg	108	61.1	115

Updated: 1/7/24 0732


**ENVIROLAB**  
 services  
**mpl**

# CHAIN OF CUSTODY FORM - Client

[Copyright and Confidential]

Client: Reditus Consulting					Client Project Name/Number/Site etc (ie report title): 22148															
Contact Person: Toby Scrivener					PO No.: 22148															
Project Mgr: Toby Scrivener					Envirolab Quote No. :															
Sampler: Kyle Sier					Date results required: standard															
Address: Unit 1A, Level 1, 29-33 Waratah Street Kirrawee NSW					Or choose: standard / same day / 1 day / 2 day / 3 day Note: Inform lab in advance if urgent turnaround is required - surcharges apply															
Phone: 02 9521 6567 Mob: 0478117515					Additional report format: esdat															
Email Invoice To: accounts@reditusconsulting.com tobyscrivener@reditus.com.au					Lab Comments:															
Email Results To: tobyscrivener@reditus.com.au, kylesier@reditusconsulting.com reditusconsulting@esdat.com.au																				
Sample information					Tests Required										Comments					
Envirolab Sample ID	Client Sample ID or information	Depth	Date sampled	Type of sample	Combo 3	PFAS Short Suite (Trace)	Cations	Anions	Ionic Balance	Nutrients	Dioxins & Furans									Provide as much information about the sample as you can
1	RMW01	-	30/05/2024	Water	x	x	x	x	x	x	x									
2	RMW02	-	30/05/2024	Water	x		x	x	x	x	x									
3	RMW03	-	30/05/2024	Water	x		x	x	x	x	x									
4	RMW04	-	30/05/2024	Water	x	x	x	x	x	x	x	x								
	RMW05	-	31/05/2024	Water	x		x	x	x	x	x									
	MW1-B	-	31/05/2024	Water	x	x														
	MW2-B	-	31/05/2024	Water	x	x														
5	MW3-B		30/05/2024	Water	x	x					x									
	MW4-B	-	31/05/2024	Water	x	x														
	EW1	-	31/05/2024	Water	x	x														
	DUP1	-	31/05/2024	Water	x	x														
	TRIP1	-	31/05/2024	Water	x	x													Please forward to ALS	
	DUP2	-	31/05/2024	Water		x														
	TRIP2	-	31/05/2024	Water		x													Please forward to ALS	
	BLANK	-	31/05/2024	Water	x	x														
					Sample Count	13	12	5	5	5	3	f	c	t	e	c	s	o	r	

Please tick the box if observed settled sediment present in water samples is to be included in the extraction and/or analysis

Relinquished by (Company): Reditus	Received by (Company): EW MD	Lab Use Only	
Print Name: Kyle Sier	Print Name: OW	Job number: 352732-A	Cooling: Ice / Ice pack / None
Date & Time: 31/05/2024	Date & Time: 30/5/24 18:00	Temperature: 8	Security seal: Intact / Broken / None
Signature: K.Sier	Signature: OW	TAT Req - SAME day / 1 / 2 / 3 / 4 / STD	

**Anna Bui**

---

**From:** tobyscrivener@reditusconsulting.com  
**Sent:** Monday, 3 June 2024 10:41 AM  
**To:** 'Kyle Sier'; Samplereceipt  
**Subject:** RE: COC - 22148

**CAUTION:** This email originated from outside of the organisation. Do not act on instructions, click links or open attachments unless you recognise the sender and know the content is authentic and safe.

Hi all – further to Kyle's email (below), could I please also add the following to samples RMW01, RMW02, RMW03, RMW04 and RMW05:

3 4

- Dissolved –
  - Aluminium (Al),
  - antimony (Sb),
  - arsenic (As),
  - barium (Ba),
  - beryllium (Be),
  - boron (B),
  - cadmium (Cd),
  - chromium (Cr),
  - cobalt (Co),
  - copper (Cu),
  - iron (Fe),
  - lead (Pb),
  - lithium (Li),
  - manganese (Mn),
  - mercury (Hg),
  - molybdenum (Mo),
  - nickel (Ni),
  - selenium (Se),
  - silica (dissolved SiO<sub>2</sub>),
  - silver (Ag),
  - strontium (Sr),
  - uranium (U),
  - vanadium (V),
  - zinc (Zn)
- Alkalinity (bicarbonate, carbonate, hydroxide and total)
- Total dissolved solids (TDS), total hardness

Thanks,

Regards,

**Toby Scrivener**  
Principal Environmental Engineer  
EIANZ Certified Environmental Practitioner - Site Contamination Specialist



Unit 1A, 29-33 Waratah Street  
Kirrawee NSW 2232

## CERTIFICATE OF ANALYSIS 352732-A

### **Client Details**

<b>Client</b>	Reditus Consulting
<b>Attention</b>	Toby Scrivener
<b>Address</b>	Shop 1, 29-33 Waratah St, KIRRRAWEE, NSW, 2232

### **Sample Details**

<b>Your Reference</b>	<b><u>22148</u></b>
<b>Number of Samples</b>	Additional analysis
<b>Date samples received</b>	30/05/2024
<b>Date completed instructions received</b>	03/06/2024

### **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.  
Samples were analysed as received from the client. Results relate specifically to the samples as received.  
Results are reported on a dry weight basis for solids and on an as received basis for other matrices.  
**Please refer to the last page of this report for any comments relating to the results.**

### **Report Details**

<b>Date results requested by</b>	13/06/2024
<b>Date of Issue</b>	13/06/2024
NATA Accreditation Number 2901. This document shall not be reproduced except in full.	
Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	

#### **Results Approved By**

Diego Bigolin, Inorganics Supervisor  
Dragana Tomas, Senior Chemist  
Giovanni Agosti, Group Technical Manager  
Sean McAlary, Chemist (FAS)  
Stuart Chen, Asbestos Approved Identifier/Report coordinator  
Timothy Toll, Senior Chemist

#### **Authorised By**

Nancy Zhang, Laboratory Manager

vTRH(C6-C10)/BTEXN in Water						
Our Reference	UNITS	352732-A-1	352732-A-2	352732-A-3	352732-A-4	352732-A-5
Your Reference		RMW01	RMW02	RMW03	RMW04	MW3-B
Date Sampled		30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water	Water	Water
Date extracted	-	11/06/2024	11/06/2024	11/06/2024	11/06/2024	11/06/2024
Date analysed	-	11/06/2024	11/06/2024	11/06/2024	11/06/2024	11/06/2024
TRH C <sub>6</sub> - C <sub>9</sub>	µg/L	<10	81	<10	<10	<10
TRH C <sub>6</sub> - C <sub>10</sub>	µg/L	<10	93	<10	<10	<10
TRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	µg/L	<10	47	<10	<10	<10
Benzene	µg/L	<1	<1	<1	<1	<1
Toluene	µg/L	<1	46	<1	<1	<1
Ethylbenzene	µg/L	<1	<1	<1	<1	<1
m+p-xylene	µg/L	<2	<2	<2	<2	<2
o-xylene	µg/L	<1	<1	<1	<1	<1
Naphthalene	µg/L	<1	<1	<1	<1	<1
Surrogate Dibromofluoromethane	%	116	111	117	118	118
Surrogate Toluene-d8	%	99	99	99	98	99
Surrogate 4-Bromofluorobenzene	%	99	99	99	98	98

svTRH (C10-C40) in Water						
Our Reference	UNITS	352732-A-1	352732-A-2	352732-A-3	352732-A-4	352732-A-5
Your Reference		RMW01	RMW02	RMW03	RMW04	MW3-B
Date Sampled		30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water	Water	Water
Date extracted	-	04/06/2024	04/06/2024	04/06/2024	04/06/2024	04/06/2024
Date analysed	-	05/06/2024	05/06/2024	05/06/2024	05/06/2024	05/06/2024
TRH C <sub>10</sub> - C <sub>14</sub>	µg/L	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	µg/L	<100	<100	<100	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	µg/L	<100	<100	<100	<100	<100
Total +ve TRH (C10-C36)	µg/L	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub>	µg/L	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	µg/L	<50	<50	<50	<50	<50
TRH >C <sub>16</sub> - C <sub>34</sub>	µg/L	<100	<100	<100	<100	<100
TRH >C <sub>34</sub> - C <sub>40</sub>	µg/L	<100	<100	<100	<100	<100
Total +ve TRH (>C10-C40)	µg/L	<50	<50	<50	<50	<50
Surrogate o-Terphenyl	%	73	87	91	100	79

PAHs in Water						
Our Reference	UNITS	352732-A-1	352732-A-2	352732-A-3	352732-A-4	352732-A-5
Your Reference		RMW01	RMW02	RMW03	RMW04	MW3-B
Date Sampled		30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water	Water	Water
Date extracted	-	04/06/2024	04/06/2024	04/06/2024	04/06/2024	04/06/2024
Date analysed	-	04/06/2024	04/06/2024	04/06/2024	04/06/2024	04/06/2024
Naphthalene	µg/L	<0.1	0.2	<0.1	<0.1	<0.1
Acenaphthylene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Fluorene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Phenanthrene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Anthracene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Fluoranthene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Pyrene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(a)anthracene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Chrysene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(b,j+k)fluoranthene	µg/L	<0.2	<0.2	<0.2	<0.2	<0.2
Benzo(a)pyrene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Indeno(1,2,3-c,d)pyrene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Dibenzo(a,h)anthracene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(g,h,i)perylene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(a)pyrene TEQ	µg/L	<0.5	<0.5	<0.5	<0.5	<0.5
Total +ve PAH's	µg/L	<0.1	0.22	<0.1	<0.1	<0.1
Surrogate p-Terphenyl-d14	%	62	68	72	76	81

All metals in water-dissolved						
Our Reference	UNITS	352732-A-1	352732-A-2	352732-A-3	352732-A-4	352732-A-5
Your Reference		RMW01	RMW02	RMW03	RMW04	MW3-B
Date Sampled		30/05/2024	30/05/2024	30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water	Water	Water
Date prepared	-	05/06/2024	05/06/2024	05/06/2024	05/06/2024	05/06/2024
Date analysed	-	05/06/2024	05/06/2024	05/06/2024	05/06/2024	05/06/2024
Aluminium-Dissolved	µg/L	9,200	5,600	16,000	100	[NA]
Antimony-Dissolved	µg/L	<1	<1	<1	<1	[NA]
Arsenic-Dissolved	µg/L	4	4	9	3	<1
Barium-Dissolved	µg/L	60	25	96	46	[NA]
Beryllium-Dissolved	µg/L	1	5	6	<0.5	[NA]
Boron-Dissolved	µg/L	1,000	1,600	1,400	2,000	[NA]
Cadmium-Dissolved	µg/L	0.5	0.9	1.1	<0.1	0.1
Chromium-Dissolved	µg/L	3	1	3	<1	<1
Cobalt-Dissolved	µg/L	25	73	92	51	[NA]
Copper-Dissolved	µg/L	<1	12	120	<1	1
Iron-Dissolved	µg/L	120,000	66,000	50,000	31,000	[NA]
Lead-Dissolved	µg/L	3	44	19	<1	<1
Lithium-Dissolved	µg/L	4	21	24	2	[NA]
Manganese-Dissolved	µg/L	480	2,500	2,600	1,900	[NA]
Mercury-Dissolved	µg/L	<0.05	<0.05	<0.05	<0.05	<0.05
Molybdenum-Dissolved	µg/L	<1	<1	<1	<1	[NA]
Nickel-Dissolved	µg/L	30	61	130	9	<1
Selenium-Dissolved	µg/L	<1	<1	<1	<1	[NA]
Silver-Dissolved	µg/L	<1	<1	<1	<1	[NA]
Strontium-Dissolved	µg/L	5,200	1,800	2,700	1,000	[NA]
Uranium-Dissolved	µg/L	<0.5	2.0	1.3	<0.5	[NA]
Vanadium-Dissolved	µg/L	5	<1	4	<1	[NA]
Zinc-Dissolved	µg/L	490	420	620	32	32

Ion Balance					
Our Reference	UNITS	352732-A-1	352732-A-2	352732-A-3	352732-A-4
Your Reference		RMW01	RMW02	RMW03	RMW04
Date Sampled		30/05/2024	30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water	Water
Date prepared	-	07/06/2024	07/06/2024	07/06/2024	07/06/2024
Date analysed	-	07/06/2024	07/06/2024	07/06/2024	07/06/2024
Calcium - Dissolved	mg/L	290	91	130	71
Potassium - Dissolved	mg/L	96	120	120	96
Sodium - Dissolved	mg/L	3,200	4,000	5,000	3,100
Magnesium - Dissolved	mg/L	430	310	400	140
Hardness (calc) equivalent CaCO <sub>3</sub>	mg/L	2,500	1,500	2,000	750
Hydroxide Alkalinity (OH <sup>-</sup> ) as CaCO <sub>3</sub>	mg/L	<5	<5	<5	<5
Bicarbonate Alkalinity as CaCO <sub>3</sub>	mg/L	<5	<5	<5	140
Carbonate Alkalinity as CaCO <sub>3</sub>	mg/L	<5	<5	<5	<5
Total Alkalinity as CaCO <sub>3</sub>	mg/L	<5	<5	<5	140
Sulphate, SO <sub>4</sub>	mg/L	980	1,400	1,600	1,200
Chloride, Cl	mg/L	6,500	6,300	8,000	5,000
Ionic Balance	%	-3.0	0	0	-6.0

<b>Miscellaneous Inorganics</b>					
Our Reference	UNITS	352732-A-1	352732-A-2	352732-A-3	352732-A-4
Your Reference		RMW01	RMW02	RMW03	RMW04
Date Sampled		30/05/2024	30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water	Water
Date prepared	-	03/06/2024	03/06/2024	03/06/2024	03/06/2024
Date analysed	-	03/06/2024	03/06/2024	03/06/2024	03/06/2024
Total Dissolved Solids (grav)	mg/L	11,000	12,000	16,000	9,400
Ammonia as N in water	mg/L	0.21	0.13	0.15	0.26
Nitrate as N in water	mg/L	<0.005	0.01	0.02	<0.005
Nitrite as N in water	mg/L	<0.005	<0.005	0.007	<0.005
NOx as N in water	mg/L	<0.005	0.01	0.03	0.008
Total Nitrogen in water	mg/L	1.1	0.3	0.4	0.3
TKN in water	mg/L	1.1	0.3	0.3	0.3
Phosphate as P in water	mg/L	<0.005	<0.005	<0.005	<0.005
Organic Nitrogen as N	mg/L	0.9	<0.2	<0.2	<0.2

Metals in Waters - Acid extractable					
Our Reference		352732-A-1	352732-A-2	352732-A-3	352732-A-4
Your Reference	UNITS	RMW01	RMW02	RMW03	RMW04
Date Sampled		30/05/2024	30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water	Water
Date prepared	-	05/06/2024	05/06/2024	05/06/2024	05/06/2024
Date analysed	-	05/06/2024	05/06/2024	05/06/2024	05/06/2024
Phosphorus - Total	mg/L	<0.05	<0.05	<0.05	<0.05

Metals in Water - Dissolved					
Our Reference		352732-A-1	352732-A-2	352732-A-3	352732-A-4
Your Reference	UNITS	RMW01	RMW02	RMW03	RMW04
Date Sampled		30/05/2024	30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water	Water
Date digested	-	04/06/2024	04/06/2024	04/06/2024	04/06/2024
Date analysed	-	04/06/2024	04/06/2024	04/06/2024	04/06/2024
Silicon*- Dissolved	mg/L	15	12	12	3.7

PFAS in Water TRACE Short				
Our Reference	UNITS	352732-A-1	352732-A-4	352732-A-5
Your Reference		RMW01	RMW04	MW3-B
Date Sampled		30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water
Date prepared	-	05/06/2024	05/06/2024	05/06/2024
Date analysed	-	05/06/2024	05/06/2024	05/06/2024
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.068	<0.0002	0.002
Perfluorooctanesulfonic acid PFOS	µg/L	0.0031	0.0003	0.010
Perfluorooctanoic acid PFOA	µg/L	0.0029	0.0002	0.002
6:2 FTS	µg/L	<0.0004	<0.0004	<0.0004
8:2 FTS	µg/L	<0.0004	<0.0004	<0.0004
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%	94	95	102
Surrogate <sup>13</sup> C <sub>2</sub> PFOA	%	92	92	94
Extracted ISTD <sup>18</sup> O <sub>2</sub> PFHxS	%	81	82	88
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%	77	83	74
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOA	%	107	100	126
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%	153	125	177
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%	173	191	#
Total Positive PFHxS & PFOS	µg/L	0.071	0.0003	0.012
Total Positive PFOS & PFOA	µg/L	0.0060	0.0006	0.012
Total Positive PFAS	µg/L	0.074	0.0006	0.014

<b>Dioxins and Furans</b>				
Our Reference		352732-A-1	352732-A-4	352732-A-5
Your Reference	UNITS	RMW01	RMW04	MW3-B
Date Sampled		30/05/2024	30/05/2024	30/05/2024
Type of sample		Water	Water	Water
See attached report		#	#	#

Method ID	Methodology Summary
Inorg-006	Alkalinity - determined titrimetrically in accordance with APHA latest edition, 2320-B.
Inorg-018	Total Dissolved Solids - determined gravimetrically. The solids are dried at 180+/-10°C.  NOTE: Where the EC of the sample is <100µS/cm, the TDS will typically be below 70mg/L (as the sample is very likely to be at least drinking water quality). Therefore to ensure data quality for TDS, the TDS is typically calculated as per the equation below:-  TDS = EC * 0.6
Inorg-040	The concentrations of the major ions (mg/L) are converted to milliequivalents and summed. The ionic balance should be within +/- 15% ie total anions = total cations +/-15%.
Inorg-055	Nitrate - determined colourimetrically. Waters samples are filtered on receipt prior to analysis. Soils are analysed following a water extraction.
Inorg-055	Nitrite - determined colourimetrically based on APHA latest edition NO2- B. Waters samples are filtered on receipt prior to analysis. Soils are analysed following a water extraction.
Inorg-055/062/127	Total Nitrogen - Calculation sum of TKN and oxidised Nitrogen. Alternatively analysed by combustion and chemiluminescence.
Inorg-057	Ammonia - determined colourimetrically, based on APHA latest edition 4500-NH3 F. Waters samples are filtered on receipt prior to analysis. Soils are analysed following a KCl extraction.
Inorg-060	Phosphate determined colourimetrically based on EPA365.1 and APHA latest edition 4500 P E. Waters samples are filtered on receipt prior to analysis. Soils are analysed following a water extraction.
Inorg-062	TKN - determined colourimetrically based on APHA latest edition 4500 Norg. Alternatively, TKN can be derived from calculation (Total N - NOx).
Inorg-081	Anions - a range of Anions are determined by Ion Chromatography, in accordance with APHA latest edition, 4110-B. Waters samples are filtered on receipt prior to analysis. Alternatively determined by colourimetry/turbidity using Discrete Analyser.
Metals-020	Determination of various metals by ICP-AES.
Metals-021	Determination of Mercury by Cold Vapour AAS.
Metals-022	Determination of various metals by ICP-MS.  Please note for Bromine and Iodine, any forms of these elements that are present are included together in the one result reported for each of these two elements.  Salt forms (e.g. FeO, PbO, ZnO) are determined stoichiometrically from the base metal concentration.
Org-020	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.

Method ID	Methodology Summary
Org-022/025	<p>Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS/GC-MSMS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013.</p>
Org-023	Water samples are analysed directly by purge and trap GC-MS.
Org-023	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.
Org-029	<p>Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.</p> <p>Analysis is undertaken with LC-MS/MS.</p> <p>PFAS results include the sum of branched and linear isomers where applicable.</p> <p>Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.</p> <p>Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.</p>

Method ID	Methodology Summary
ORG-038	<p>Water samples are extracted with DCM and concentrated. The extract is analysed by GC/MSMS for selected Dioxin and Furans.</p> <p>Soils and Sorbents are solvent extracted, followed by an extract clean-up and GC/MSMS analysis.</p> <ol style="list-style-type: none"> <li>1. I -TEQ(zero) and WHO-TEQ(zero) calculated where analyte components that are &lt;PQL are considered to be zero in the TEQ calculation. Where all sample analyte results are &lt;PQL, the calculated sample TEQ = 0, this is due to the calculation being an arithmetic formula and therefore does not reflect the associated PQLs.</li> <li>2. I -TEQ(0.5) and WHO-TEQ(0.5) calculated where analyte components that are &lt;PQL are considered to be 0.5 * the component PQL in the TEQ calculation.</li> <li>3. I-TEQ(PQL) and WHO-TEQ(PQL) calculated where analyte components that are &lt;PQL are considered to be equal to the component PQL in the TEQ calculation.</li> </ol> <p>13C Rec is the recovery of Isotopically labelled compound added by the Laboratory for quantification and to measure extraction efficiency.</p> <p>I-TEF - International toxic equivalency factor  I-TEQ - International toxic equivalence  WHO-TEF - World Health Organisation toxic equivalency factor  WHO-TEQ - World Health Organisation toxic equivalence</p>

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Water							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date extracted	-			11/06/2024	[NT]	[NT]	[NT]	[NT]	11/06/2024	[NT]
Date analysed	-			11/06/2024	[NT]	[NT]	[NT]	[NT]	11/06/2024	[NT]
TRH C <sub>6</sub> - C <sub>9</sub>	µg/L	10	Org-023	<10	[NT]	[NT]	[NT]	[NT]	83	[NT]
TRH C <sub>6</sub> - C <sub>10</sub>	µg/L	10	Org-023	<10	[NT]	[NT]	[NT]	[NT]	83	[NT]
Benzene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	85	[NT]
Toluene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	84	[NT]
Ethylbenzene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	82	[NT]
m+p-xylene	µg/L	2	Org-023	<2	[NT]	[NT]	[NT]	[NT]	83	[NT]
o-xylene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	82	[NT]
Naphthalene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate Dibromofluoromethane	%		Org-023	109	[NT]	[NT]	[NT]	[NT]	129	[NT]
Surrogate Toluene-d8	%		Org-023	100	[NT]	[NT]	[NT]	[NT]	108	[NT]
Surrogate 4-Bromofluorobenzene	%		Org-023	98	[NT]	[NT]	[NT]	[NT]	106	[NT]

QUALITY CONTROL: svTRH (C10-C40) in Water							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W2	352732-A-2
Date extracted	-			04/06/2024	1	04/06/2024	04/06/2024		04/06/2024	04/06/2024
Date analysed	-			05/06/2024	1	05/06/2024	05/06/2024		05/06/2024	05/06/2024
TRH C <sub>10</sub> - C <sub>14</sub>	µg/L	50	Org-020	<50	1	<50	<50	0	98	92
TRH C <sub>15</sub> - C <sub>28</sub>	µg/L	100	Org-020	<100	1	<100	<100	0	103	97
TRH C <sub>29</sub> - C <sub>36</sub>	µg/L	100	Org-020	<100	1	<100	<100	0	100	80
TRH >C <sub>10</sub> - C <sub>16</sub>	µg/L	50	Org-020	<50	1	<50	<50	0	98	92
TRH >C <sub>16</sub> - C <sub>34</sub>	µg/L	100	Org-020	<100	1	<100	<100	0	103	97
TRH >C <sub>34</sub> - C <sub>40</sub>	µg/L	100	Org-020	<100	1	<100	<100	0	100	80
Surrogate o-Terphenyl	%		Org-020	90	1	73	60	20	113	96

QUALITY CONTROL: PAHs in Water					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	352732-A-2
Date extracted	-			04/06/2024	1	04/06/2024	04/06/2024		04/06/2024	04/06/2024
Date analysed	-			04/06/2024	1	04/06/2024	04/06/2024		04/06/2024	04/06/2024
Naphthalene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	109	120
Acenaphthylene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Acenaphthene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	115	118
Fluorene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	108	121
Phenanthrene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	110	106
Anthracene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Fluoranthene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	105	103
Pyrene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	103	102
Benzo(a)anthracene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Chrysene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	103	103
Benzo(b,j+k)fluoranthene	µg/L	0.2	Org-022/025	<0.2	1	<0.2	<0.2	0	[NT]	[NT]
Benzo(a)pyrene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	91	98
Indeno(1,2,3-c,d)pyrene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Dibenzo(a,h)anthracene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Benzo(g,h,i)perylene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-022/025	85	1	62	62	0	81	78

QUALITY CONTROL: All metals in water-dissolved					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W4	352732-A-2
Date prepared	-			05/06/2024	1	05/06/2024	05/06/2024		05/06/2024	05/06/2024
Date analysed	-			05/06/2024	1	05/06/2024	05/06/2024		05/06/2024	05/06/2024
Aluminium-Dissolved	µg/L	10	Metals-022	<10	1	9200	9300	1	96	[NT]
Antimony-Dissolved	µg/L	1	Metals-022	<1	1	<1	<1	0	94	[NT]
Arsenic-Dissolved	µg/L	1	Metals-022	<1	1	4	4	0	101	[NT]
Barium-Dissolved	µg/L	1	Metals-022	<1	1	60	58	3	107	[NT]
Beryllium-Dissolved	µg/L	0.5	Metals-022	<0.5	1	1	1	0	83	[NT]
Boron-Dissolved	µg/L	20	Metals-022	<20	1	1000	1100	10	112	[NT]
Cadmium-Dissolved	µg/L	0.1	Metals-022	<0.1	1	0.5	0.5	0	98	[NT]
Chromium-Dissolved	µg/L	1	Metals-022	<1	1	3	3	0	98	[NT]
Cobalt-Dissolved	µg/L	1	Metals-022	<1	1	25	26	4	103	[NT]
Copper-Dissolved	µg/L	1	Metals-022	<1	1	<1	<1	0	98	[NT]
Iron-Dissolved	µg/L	10	Metals-022	<10	1	120000	140000	15	99	[NT]
Lead-Dissolved	µg/L	1	Metals-022	<1	1	3	3	0	98	[NT]
Lithium-Dissolved	µg/L	1	Metals-022	<1	1	4	4	0	92	[NT]
Manganese-Dissolved	µg/L	5	Metals-022	<5	1	480	490	2	97	[NT]
Mercury-Dissolved	µg/L	0.05	Metals-021	<0.05	1	<0.05	<0.05	0	101	77
Molybdenum-Dissolved	µg/L	1	Metals-022	<1	1	<1	<1	0	99	[NT]
Nickel-Dissolved	µg/L	1	Metals-022	<1	1	30	30	0	101	[NT]
Selenium-Dissolved	µg/L	1	Metals-022	<1	1	<1	<1	0	98	[NT]
Silver-Dissolved	µg/L	1	Metals-022	<1	1	<1	<1	0	104	[NT]
Strontium-Dissolved	µg/L	1	Metals-022	<1	1	5200	5200	0	99	[NT]
Uranium-Dissolved	µg/L	0.5	Metals-022	<0.5	1	<0.5	<0.5	0	91	[NT]
Vanadium-Dissolved	µg/L	1	Metals-022	<1	1	5	5	0	99	[NT]
Zinc-Dissolved	µg/L	1	Metals-022	<1	1	490	500	2	98	[NT]

QUALITY CONTROL: Ion Balance						Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]	
Date prepared	-			07/06/2024	[NT]	[NT]	[NT]	[NT]	07/06/2024	[NT]	
Date analysed	-			07/06/2024	[NT]	[NT]	[NT]	[NT]	07/06/2024	[NT]	
Calcium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	105	[NT]	
Potassium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	98	[NT]	
Sodium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	101	[NT]	
Magnesium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	102	[NT]	
Hydroxide Alkalinity (OH <sup>-</sup> ) as CaCO <sub>3</sub>	mg/L	5	Inorg-006	<5	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]	
Bicarbonate Alkalinity as CaCO <sub>3</sub>	mg/L	5	Inorg-006	<5	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]	
Carbonate Alkalinity as CaCO <sub>3</sub>	mg/L	5	Inorg-006	<5	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]	
Total Alkalinity as CaCO <sub>3</sub>	mg/L	5	Inorg-006	<5	[NT]	[NT]	[NT]	[NT]	109	[NT]	
Sulphate, SO <sub>4</sub>	mg/L	1	Inorg-081	<1	[NT]	[NT]	[NT]	[NT]	115	[NT]	
Chloride, Cl	mg/L	1	Inorg-081	<1	[NT]	[NT]	[NT]	[NT]	109	[NT]	

QUALITY CONTROL: Miscellaneous Inorganics							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	352732-A-2
Date prepared	-			03/06/2024	1	03/06/2024	03/06/2024		03/06/2024	03/06/2024
Date analysed	-			03/06/2024	1	03/06/2024	03/06/2024		03/06/2024	03/06/2024
Total Dissolved Solids (grav)	mg/L	5	Inorg-018	<5	1	11000	[NT]		96	[NT]
Ammonia as N in water	mg/L	0.005	Inorg-057	<0.005	1	0.21	0.23	9	90	#
Nitrate as N in water	mg/L	0.005	Inorg-055	<0.005	1	<0.005	<0.005	0	95	#
Nitrite as N in water	mg/L	0.005	Inorg-055	<0.005	1	<0.005	<0.005	0	99	#
NOx as N in water	mg/L	0.005	Inorg-055	<0.005	1	<0.005	<0.005	0	95	#
Total Nitrogen in water	mg/L	0.1	Inorg-055/062/127	<0.1	1	1.1	1.1	0	89	[NT]
TKN in water	mg/L	0.1	Inorg-062	<0.1	1	1.1	1.1	0	[NT]	[NT]
Phosphate as P in water	mg/L	0.005	Inorg-060	<0.005	1	<0.005	<0.005	0	107	#
Organic Nitrogen as N	mg/L	0.2	Inorg-055/062/127	<0.2	1	0.9	0.8	12	[NT]	[NT]

QUALITY CONTROL: Metals in Waters - Acid extractable							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			05/06/2024	1	05/06/2024	05/06/2024		05/06/2024	[NT]
Date analysed	-			05/06/2024	1	05/06/2024	05/06/2024		05/06/2024	[NT]
Phosphorus - Total	mg/L	0.05	Metals-020	<0.05	1	<0.05	<0.05	0	93	[NT]

QUALITY CONTROL: Metals in Water - Dissolved							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date digested	-			04/06/2024	[NT]	[NT]	[NT]	[NT]	04/06/2024	[NT]
Date analysed	-			04/06/2024	[NT]	[NT]	[NT]	[NT]	04/06/2024	[NT]
Silicon*- Dissolved	mg/L	0.2	Metals-020	<0.2	[NT]	[NT]	[NT]	[NT]	96	[NT]

QUALITY CONTROL: PFAS in Water TRACE Short						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W2	[NT]
Date prepared	-			05/06/2024	[NT]	[NT]	[NT]	[NT]	05/06/2024	[NT]
Date analysed	-			05/06/2024	[NT]	[NT]	[NT]	[NT]	05/06/2024	[NT]
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.0002	Org-029	<0.0002	[NT]	[NT]	[NT]	[NT]	100	[NT]
Perfluorooctanesulfonic acid PFOS	µg/L	0.0002	Org-029	<0.0002	[NT]	[NT]	[NT]	[NT]	101	[NT]
Perfluorooctanoic acid PFOA	µg/L	0.0002	Org-029	<0.0002	[NT]	[NT]	[NT]	[NT]	100	[NT]
6:2 FTS	µg/L	0.0004	Org-029	<0.0004	[NT]	[NT]	[NT]	[NT]	96	[NT]
8:2 FTS	µg/L	0.0004	Org-029	<0.0004	[NT]	[NT]	[NT]	[NT]	92	[NT]
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%		Org-029	98	[NT]	[NT]	[NT]	[NT]	99	[NT]
Surrogate <sup>13</sup> C <sub>2</sub> PFOA	%		Org-029	99	[NT]	[NT]	[NT]	[NT]	95	[NT]
Extracted ISTD <sup>18</sup> O <sub>2</sub> PFHxS	%		Org-029	83	[NT]	[NT]	[NT]	[NT]	83	[NT]
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%		Org-029	69	[NT]	[NT]	[NT]	[NT]	75	[NT]
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOA	%		Org-029	110	[NT]	[NT]	[NT]	[NT]	111	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%		Org-029	175	[NT]	[NT]	[NT]	[NT]	164	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%		Org-029	190	[NT]	[NT]	[NT]	[NT]	186	[NT]

QUALITY CONTROL: Dioxins and Furans						Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]	
See attached report			ORG-038	#	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]	

**Result Definitions**

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<	Less than
>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	
The recommended maximums for analytes in urine are taken from "2018 TLVs and BEIs", as published by ACGIH (where available). Limit provided for Nickel is a precautionary guideline as per Position Paper prepared by AIOP Exposure Standards Committee, 2016.	
Guideline limits for Rinse Water Quality reported as per analytical requirements and specifications of AS 4187, Amdt 2 2019, Table 7.2	

## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Where matrix spike recoveries fall below the lower limit of the acceptance criteria (e.g. for non-labile or standard Organics <60%), positive result(s) in the parent sample will subsequently have a higher than typical estimated uncertainty (MU estimates supplied on request) and in these circumstances the sample result is likely biased significantly low.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

## **Report Comments**

NO<sub>2</sub>/NO<sub>3</sub>/PO<sub>4</sub> - out of recommended holding time

MISC\_INORG: # Percent recovery not reported due to matrix interferences. Samples were diluted and reanalysed and the poor recovery was confirmed. However, an acceptable recovery was obtained for the LCS.

Total metals: no unfiltered, preserved sample was received, therefore analysis was conducted from the unpreserved sample bottle.  
Note: there is a possibility some elements may be underestimated.

For PFAS Extracted Internal Standards denoted with # or outside the 50-150% acceptance range, the respective target analyte results may be unaffected, in other circumstances the PQL has been raised to accommodate the outlier(s).

Dioxins analysed by MPL Laboratories. Report no. PFF0035.

## Certificate of Analysis PFF0035

### Client Details

<b>Client</b>	Envirolab (Sydney)
<b>Contact</b>	Results Receivable
<b>Address</b>	12 Ashley St, Chatswood, NSW, 2067

### Sample Details

<b>Your Reference</b>	352732
<b>Number of Samples</b>	3 Water
<b>Date Samples Received</b>	04/06/2024
<b>Date Instructions Received</b>	04/06/2024

### Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.  
Samples were analysed as received from the client. Results relate specifically to the samples as received.  
Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

### Report Details

<b>Date Results Requested by</b>	13/06/2024
<b>Date of Issue</b>	13/06/2024

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**Accredited for compliance with ISO/IEC 17025. Tests not covered by NATA are denoted with \*.**

### Authorisation Details

<b>Results Approved By</b>	Huong Patfield, Organics Chemist
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<b>Laboratory Manager</b>	Michael Kubiak
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# Certificate of Analysis PFF0035

## Samples in this Report

Envirolab ID	Sample ID	Matrix	Date Sampled	Date Received
PFF0035-01	352732-1	Water	04/06/2024	04/06/2024
PFF0035-02	352732-4	Water	04/06/2024	04/06/2024
PFF0035-03	352732-5	Water	04/06/2024	04/06/2024

## Sample Comments

General Comment      No sampling date(s) was/were provided by client. Therefore the sampling date(s) is/are assigned as the date(s) of sample receipt to the laboratory.

# Certificate of Analysis PFF0035

## Dioxins/Furans (Water)

<b>Envirolab ID:</b>	PFF0035-01		<b>Date Sampled:</b>	04/06/2024					
<b>Client ID:</b>	352732-1								
<b>Analyte</b>									
2,3,7,8-TCDD	5.00	pg/L	<5.0						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	1	0.0	2.5	5.0	1	0.0	2.5	5.0	80.2%
1,2,3,7,8-PeCDD	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	1	0.0	10	20	0.5	0.0	5.0	10	88.9%
1,2,3,4,7,8-HxCDD	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0	83.8%
1,2,3,6,7,8-HxCDD	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0	98.1%
1,2,3,7,8,9-HxCDD	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0	-
1,2,3,4,6,7,8-HpCDD	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.0	0.10	0.20	0.01	0.0	0.10	0.20	87.0%
OCDD	50.00	pg/L	120						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.0003	0.036	0.036	0.036	0.001	0.12	0.12	0.12	86.8%
2,3,7,8-TCDF	5.00	pg/L	<5.0						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	0.25	0.50	0.1	0.0	0.25	0.50	79.4%
1,2,3,7,8-PeCDF	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.03	0.0	0.30	0.60	0.05	0.0	0.50	1.0	84.2%
2,3,4,7,8-PeCDF	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.3	0.0	3.0	6.0	0.5	0.0	5.0	10	91.8%
1,2,3,4,7,8-HxCDF	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0	90.0%
1,2,3,6,7,8-HxCDF	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0	95.4%
1,2,3,7,8,9-HxCDF	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0	91.5%
2,3,4,6,7,8-HxCDF	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0	91.8%
1,2,3,4,6,7,8-HpCDF	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.0	0.10	0.20	0.01	0.0	0.10	0.20	89.6%
1,2,3,4,7,8,9-HpCDF	20.00	pg/L	<20						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.01	0.0	0.10	0.20	0.01	0.0	0.10	0.20	87.4%
OCDF	50.00	pg/L	<50						
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
	0.0003	0.0	0.0075	0.015	0.001	0.0	0.025	0.050	-

# Certificate of Analysis PFF0035

<b>Envirolab ID:</b>	PFF0035-02	<b>Date Sampled:</b>		04/06/2024							
<b>Client ID:</b>	352732-4										
<b>Analyte</b>		<b>PQL</b>	<b>Units</b>	<b>Result</b>							
2,3,7,8-TCDD	5.00	pg/L	<5.0	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	1	0.0	2.5		5.0		1	0.0	2.5	5.0	92.5%
1,2,3,7,8-PeCDD	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	1	0.0	10		20		0.5	0.0	5.0	10	102%
1,2,3,4,7,8-HxCDD	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.1	0.0	1.0		2.0		0.1	0.0	1.0	2.0	99.3%
1,2,3,6,7,8-HxCDD	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.1	0.0	1.0		2.0		0.1	0.0	1.0	2.0	110%
1,2,3,7,8,9-HxCDD	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.1	0.0	1.0		2.0		0.1	0.0	1.0	2.0	-
1,2,3,4,6,7,8-HpCDD	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.01	0.0	0.10		0.20		0.01	0.0	0.10	0.20	107%
OCDD	50.00	pg/L	<50	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.0003	0.0079	0.0079		0.0079		0.001	0.026	0.026	0.026	103%
2,3,7,8-TCDF	5.00	pg/L	<5.0	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.1	0.0	0.25		0.50		0.1	0.0	0.25	0.50	92.6%
1,2,3,7,8-PeCDF	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.03	0.0	0.30		0.60		0.05	0.0	0.50	1.0	100%
2,3,4,7,8-PeCDF	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.3	0.0	3.0		6.0		0.5	0.0	5.0	10	102%
1,2,3,4,7,8-HxCDF	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.1	0.0	1.0		2.0		0.1	0.0	1.0	2.0	112%
1,2,3,6,7,8-HxCDF	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.1	0.0	1.0		2.0		0.1	0.0	1.0	2.0	108%
1,2,3,7,8,9-HxCDF	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.1	0.0	1.0		2.0		0.1	0.0	1.0	2.0	98.2%
2,3,4,6,7,8-HxCDF	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.1	0.0	1.0		2.0		0.1	0.0	1.0	2.0	105%
1,2,3,4,6,7,8-HpCDF	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.01	0.0	0.10		0.20		0.01	0.0	0.10	0.20	109%
1,2,3,4,7,8,9-HpCDF	20.00	pg/L	<20	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.01	0.0	0.10		0.20		0.01	0.0	0.10	0.20	103%
OCDF	50.00	pg/L	<50	<i>WHO-TEQ1</i>	<i>WHO-TEQ2</i>	<i>WHO-TEQ3</i>	<i>I-TEF</i>	<i>I-TEQ1</i>	<i>I-TEQ2</i>	<i>I-TEQ3</i>	<i>Recovery</i>
	<i>WHO-TEF</i>										
	0.0003	0.0075	0.015		0.025		0.001	0.0	0.025	0.050	-

# Certificate of Analysis PFF0035

<b>Envirolab ID:</b>	PFF0035-03	<b>Date Sampled:</b>		04/06/2024								
<b>Client ID:</b>	352732-5											
Analyte	PQL	Units	Result	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3	Recovery
2,3,7,8-TCDD	5.00	pg/L	<5.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				
	1	0.0	2.5	5.0	1	0.0	2.5	5.0				112%
1,2,3,7,8-PeCDD	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	1	0.0	10	20	0.5	0.0	5.0	10				112%
1,2,3,4,7,8-HxCDD	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0				120%
1,2,3,6,7,8-HxCDD	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0				124%
1,2,3,7,8,9-HxCDD	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0				-
1,2,3,4,6,7,8-HpCDD	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.01	0.0	0.10	0.20	0.01	0.0	0.10	0.20				113%
OCDD	50.00	pg/L	57									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.0003	0.017	0.017	0.017	0.001	0.057	0.057	0.057				108%
2,3,7,8-TCDF	5.00	pg/L	<5.0									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.1	0.0	0.25	0.50	0.1	0.0	0.25	0.50				116%
1,2,3,7,8-PeCDF	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.03	0.0	0.30	0.60	0.05	0.0	0.50	1.0				118%
2,3,4,7,8-PeCDF	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.3	0.0	3.0	6.0	0.5	0.0	5.0	10				121%
1,2,3,4,7,8-HxCDF	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0				119%
1,2,3,6,7,8-HxCDF	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0				114%
1,2,3,7,8,9-HxCDF	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0				111%
2,3,4,6,7,8-HxCDF	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.1	0.0	1.0	2.0	0.1	0.0	1.0	2.0				119%
1,2,3,4,6,7,8-HpCDF	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.01	0.0	0.10	0.20	0.01	0.0	0.10	0.20				117%
1,2,3,4,7,8,9-HpCDF	20.00	pg/L	<20									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.01	0.0	0.10	0.20	0.01	0.0	0.10	0.20				112%
OCDF	50.00	pg/L	<50									
	WHO-TEF	WHO-TEQ1	WHO-TEQ2	WHO-TEQ3	I-TEF	I-TEQ1	I-TEQ2	I-TEQ3				Recovery
	0.0003	0.0	0.0075	0.015	0.001	0.0	0.025	0.050				-

# Certificate of Analysis PFF0035

## Method Summary

Method ID	Methodology Summary
ORG-025	Determination of semi-volatile organic compounds (SVOCs) by GC-MS-MS. Water samples are extracted by LLE and soils/solids/biota using DCM/Acetone/Methanol.
ORG-025_DIOXIN	Water samples are extracted with DCM and concentrated. The extract is analysed by GC-MSMS for Dioxin and Furans. Soils, Biota and Sorbents are solvent extracted, followed by clean-up and GC-MSMS analysis. 1. I -TEQ(zero) and WHO-TEQ(zero) calculated where analyte components that are <PQL are considered to be zero in the TEQ calculation. Where all sample analyte results are <PQL, the calculated sample TEQ = 0, this is due to the calculation being an arithmetic formula and therefore does not reflect the associated PQLs. 2. I -TEQ(0.5) and WHO-TEQ(0.5) calculated where analyte components that are <PQL are considered to be 0.5 * the component PQL in the TEQ calculation. 3. I-TEQ(PQL) and WHO-TEQ(PQL) calculated where analyte components that are <PQL are considered to be equal to the component PQL in the TEQ calculation. 13C12 Rec is the recovery of Isotopically labelled compound added by the Laboratory for quantification and to measure extraction efficiency. I-TEF - International toxic equivalency factor I-TEQ - International toxic equivalence WHO-TEF - World Health Organisation toxic equivalency factor WHO-TEQ - World Health Organisation toxic equivalence TEQ values are rounded to the same number of significant figures as the raw results for consistency and therefore may not calculate out exactly as PQL * TEF, given rounded up or down as appropriate.

# Certificate of Analysis PFF0035

## Result Definitions

Identifier	Description
<b>NR</b>	Not reported
<b>NEPM</b>	National Environment Protection Measure
<b>NS</b>	Not specified
<b>LCS</b>	Laboratory Control Sample
<b>RPD</b>	Relative Percent Difference
>	Greater than
<	Less than
<b>PQL</b>	Practical Quantitation Limit
<b>INS</b>	Insufficient sample for this test
<b>NA</b>	Test not required
<b>NT</b>	Not tested
<b>DOL</b>	Samples rejected due to particulate overload (air filters only)
<b>RFD</b>	Samples rejected due to filter damage (air filters only)
<b>RUD</b>	Samples rejected due to uneven deposition (air filters only)
<b>##</b>	Indicates a laboratory acceptance criteria outlier, for further details, see Result Comments and/or QC Comments

## Quality Control Definitions

### Blank

This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, and is determined by processing solvents and reagents in exactly the same manner as for samples.

### Surrogate Spike

Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.

### LCS (Laboratory Control Sample)

This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.

### Matrix Spike

A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.

### Duplicate

This is the complete duplicate analysis of a sample from the process batch. The sample selected should be one where the analyte concentration is easily measurable.

# Certificate of Analysis PFF0035

## Laboratory Acceptance Criteria

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Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria. Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction. Spikes for Physical and Aggregate Tests are not applicable. For VOCs in water samples, three vials are required for duplicate or spike analysis.

General Acceptance Criteria (GAC) - Analyte specific criteria applies for some analytes and is reflected in QC recovery tables.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% - see ELN-P05 QAQC tables for details (available on request); <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase. Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals; 60-140% for organics (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was typically insufficient in order to satisfy laboratory QA/QC protocols.

## Miscellaneous Information

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When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached. We have taken the sampling date as being the date received at the laboratory.

Two significant figures are reported for the majority of tests and with a high degree of confidence, for results <10\*xPQL, the second significant figure may be in doubt i.e. has a relatively high degree of uncertainty and is provided for information only.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS where sediment/solids are included by default.

Urine Analysis - The BEI values listed are taken from the 2022 edition of *TLVs and BEIs Threshold Limits by ACGIH*.

Air volume measurements are not covered by Envirolab's NATA accreditation.

# Data Quality Assessment Summary PFF0035

## Client Details

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<b>Client</b>	Envirolab (Sydney)
<b>Your Reference</b>	352732
<b>Date Issued</b>	13/06/2024

## Recommended Holding Time Compliance

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No recommended holding time exceedances

## Quality Control and QC Frequency

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QC Type	Compliant	Details
Blank	Yes	No Outliers
LCS	Yes	No Outliers
Duplicates	Yes	No Outliers
Matrix Spike	Yes	No Outliers
Surrogates / Extracted Internal Standards	Yes	No Outliers
QC Frequency	Yes	No Outliers

Surrogates/Extracted Internal Standards, Duplicates and/or Matrix Spikes are not always relevant/applicable to certain analyses and matrices. Therefore, said QC measures are deemed compliant in these situations by default. See Laboratory Acceptance Criteria for more information

# Data Quality Assessment Summary PFF0035

## Recommended Holding Time Compliance

Analysis	Sample Number(s)	Date Sampled	Date Extracted	Date Analysed	Compliant
1,2,3,4,6,7,8-HpCDD   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,4,6,7,8-HpCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,4,7,8,9-HpCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,4,7,8-HxCDD   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,4,7,8-HxCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,6,7,8-HxCDD   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,6,7,8-HXCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,7,8,9-HxCDD   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,7,8,9-HxCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,7,8-PeCDD   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
1,2,3,7,8-PeCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
2,3,4,6,7,8-HxCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
2,3,4,7,8-PeCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
2,3,7,8-TCDD   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
2,3,7,8-TCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
OCDD   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes
OCDF   Water	1-2	04/06/2024	07/06/2024	12/06/2024	Yes
	3	04/06/2024	07/06/2024	13/06/2024	Yes

No sampling date(s) was/were provided by client. Therefore the sampling date(s) is/are assigned as the date(s) of sample receipt to the laboratory.

# Quality Control PFF0035

## ORG-025\_DIOXIN | Dioxins/Furans (Water) | Batch BFF0816

Analyte	Units	PQL	Blank	DUP1	LCS %	Spike %
				PFF0035-01 Samp   QC   RPD %		
2,3,7,8-TCDD	pg/L	5.0	<5.0	<5.0   <5.0   [NA]	97.1	98.0
Surrogate 13C-2,3,7,8-TCDD	%		97.9	80.2 / 84.3	97.1	61.2
2,3,7,8-TCDD WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,7,8-TCDD WHO-TEQ2	pg/L		2.50	2.50   2.50   0.00	[NA]	[NA]
2,3,7,8-TCDD WHO-TEQ3	pg/L		5.00	5.00   5.00   0.00	[NA]	[NA]
2,3,7,8-TCDD I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,7,8-TCDD I-TEQ2	pg/L		2.50	2.50   2.50   0.00	[NA]	[NA]
2,3,7,8-TCDD I-TEQ3	pg/L		5.00	5.00   5.00   0.00	[NA]	[NA]
1,2,3,7,8-PeCDD	pg/L	20	<20	<20   <20   [NA]	100	83.0
Surrogate 13C-1,2,3,7,8-PeCDD	%		90.5	88.9 / 90.6	95.2	72.9
1,2,3,7,8-PeCDD WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8-PeCDD WHO-TEQ2	pg/L		10.0	10.0   10.0   0.00	[NA]	[NA]
1,2,3,7,8-PeCDD WHO-TEQ3	pg/L		20.0	20.0   20.0   0.00	[NA]	[NA]
1,2,3,7,8-PeCDD I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8-PeCDD I-TEQ2	pg/L		5.00	5.00   5.00   0.00	[NA]	[NA]
1,2,3,7,8-PeCDD I-TEQ3	pg/L		10.0	10.0   10.0   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDD	pg/L	20	<20	<20   <20   [NA]	99.7	75.3
Surrogate 13C-1,2,3,4,7,8-HxCDD	%		102	83.8 / 88.7	91.9	73.9
1,2,3,4,7,8-HxCDD WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8-HxCDD WHO-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDD WHO-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDD I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8-HxCDD I-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDD I-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDD	pg/L	20	<20	<20   <20   [NA]	96.5	73.7
Surrogate 13C-1,2,3,6,7,8-HxCDD	%		89.7	98.1 / 101	98.8	83.2
1,2,3,6,7,8-HxCDD WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,6,7,8-HxCDD WHO-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDD WHO-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDD I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,6,7,8-HxCDD I-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDD I-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDD	pg/L	20	<20	<20   <20   [NA]	96.3	72.2
1,2,3,7,8,9-HxCDD WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8,9-HxCDD WHO-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDD WHO-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDD I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8,9-HxCDD I-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDD I-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD	pg/L	20	<20	<20   <20   [NA]	98.2	71.0
Surrogate 13C-1,2,3,4,6,7,8-HpCDD	%		97.1	87.0 / 95.4	100	78.8
1,2,3,4,6,7,8-HpCDD WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD WHO-TEQ2	pg/L		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD WHO-TEQ3	pg/L		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD I-TEQ2	pg/L		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HpCDD I-TEQ3	pg/L		0.200	0.200   0.200   0.00	[NA]	[NA]
OCDD	pg/L	50	<50	121   168   32.4	101	70.3
Surrogate 13C-OCDD	%		94.0	86.8 / 90.3	94.5	78.6
OCDD WHO-TEQ1	pg/L		0.00	0.0363   0.0503   32.4	[NA]	[NA]
OCDD WHO-TEQ2	pg/L		0.00750	0.0363   0.0503   32.4	[NA]	[NA]
OCDD WHO-TEQ3	pg/L		0.0150	0.0363   0.0503   32.4	[NA]	[NA]
OCDD I-TEQ1	pg/L		0.00	0.121   0.168   32.4	[NA]	[NA]
OCDD I-TEQ2	pg/L		0.0250	0.121   0.168   32.4	[NA]	[NA]
OCDD I-TEQ3	pg/L		0.0500	0.121   0.168   32.4	[NA]	[NA]
2,3,7,8-TCDF	pg/L	5.0	<5.0	<5.0   <5.0   [NA]	107	103
Surrogate 13C-2,3,7,8-TCDF	%		99.0	79.4 / 82.8	96.2	64.4

# Quality Control PFF0035

## ORG-025\_DIOXIN | Dioxins/Furans (Water) | Batch BFF0816

Analyte	Units	PQL	Blank	DUP1	LCS %	Spike %
				PFF0035-01 Samp   QC   RPD %		
2,3,7,8-TCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,7,8-TCDF WHO-TEQ2	pg/L		0.250	0.250   0.250   0.00	[NA]	[NA]
2,3,7,8-TCDF WHO-TEQ3	pg/L		0.500	0.500   0.500   0.00	[NA]	[NA]
2,3,7,8-TCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,7,8-TCDF I-TEQ2	pg/L		0.250	0.250   0.250   0.00	[NA]	[NA]
2,3,7,8-TCDF I-TEQ3	pg/L		0.500	0.500   0.500   0.00	[NA]	[NA]
1,2,3,7,8-PeCDF	pg/L	20	<20	<20   <20   [NA]	101	81.7
<i>Surrogate 13C-1,2,3,7,8-PeCDF</i>	%		95.7	84.2 / 89.6	94.0	74.3
1,2,3,7,8-PeCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8-PeCDF WHO-TEQ2	pg/L		0.300	0.300   0.300   0.00	[NA]	[NA]
1,2,3,7,8-PeCDF WHO-TEQ3	pg/L		0.600	0.600   0.600   0.00	[NA]	[NA]
1,2,3,7,8-PeCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8-PeCDF I-TEQ2	pg/L		0.500	0.500   0.500   0.00	[NA]	[NA]
1,2,3,7,8-PeCDF I-TEQ3	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
2,3,4,7,8-PeCDF	pg/L	20	<20	<20   <20   [NA]	101	86.7
<i>Surrogate 13C-2,3,4,7,8-PeCDF</i>	%		94.9	91.8 / 88.9	92.3	68.2
2,3,4,7,8-PeCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,4,7,8-PeCDF WHO-TEQ2	pg/L		3.00	3.00   3.00   0.00	[NA]	[NA]
2,3,4,7,8-PeCDF WHO-TEQ3	pg/L		6.00	6.00   6.00   0.00	[NA]	[NA]
2,3,4,7,8-PeCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,4,7,8-PeCDF I-TEQ2	pg/L		5.00	5.00   5.00   0.00	[NA]	[NA]
2,3,4,7,8-PeCDF I-TEQ3	pg/L		10.0	10.0   10.0   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDF	pg/L	20	<20	<20   <20   [NA]	99.5	68.9
<i>Surrogate 13C-1,2,3,4,7,8-HxCDF</i>	%		90.4	90.0 / 94.8	95.4	77.8
1,2,3,4,7,8-HxCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8-HxCDF WHO-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDF WHO-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8-HxCDF I-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,4,7,8-HxCDF I-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDF	pg/L	20	<20	<20   <20   [NA]	103	61.8
<i>Surrogate 13C-1,2,3,6,7,8-HxCDF</i>	%		89.8	95.4 / 96.1	96.4	86.0
1,2,3,6,7,8-HxCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,6,7,8-HxCDF WHO-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDF WHO-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,6,7,8-HxCDF I-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,6,7,8-HxCDF I-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDF	pg/L	20	<20	<20   <20   [NA]	104	104
<i>Surrogate 13C-1,2,3,7,8,9-HxCDF</i>	%		98.5	91.5 / 87.7	96.8	74.2
1,2,3,7,8,9-HxCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8,9-HxCDF WHO-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDF WHO-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,7,8,9-HxCDF I-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
1,2,3,7,8,9-HxCDF I-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
2,3,4,6,7,8-HxCDF	pg/L	20	<20	<20   <20   [NA]	102	75.4
<i>Surrogate 13C-2,3,4,6,7,8-HxCDF</i>	%		89.8	91.8 / 94.2	98.2	78.2
2,3,4,6,7,8-HxCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,4,6,7,8-HxCDF WHO-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
2,3,4,6,7,8-HxCDF WHO-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
2,3,4,6,7,8-HxCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
2,3,4,6,7,8-HxCDF I-TEQ2	pg/L		1.00	1.00   1.00   0.00	[NA]	[NA]
2,3,4,6,7,8-HxCDF I-TEQ3	pg/L		2.00	2.00   2.00   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HpCDF	pg/L	20	<20	<20   <20   [NA]	99.6	66.9
<i>Surrogate 13C-1,2,3,4,6,7,8-HpCDF</i>	%		97.7	89.6 / 93.8	95.4	79.4
1,2,3,4,6,7,8-HpCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]

# Quality Control PFF0035

## ORG-025\_DIOXIN | Dioxins/Furans (Water) | Batch BFF0816

Analyte	Units	PQL	Blank	DUP1	LCS %	Spike %
				PFF0035-01 Samp   QC   RPD %		
1,2,3,4,6,7,8-HpCDF WHO-TEQ2	pg/L		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HpCDF WHO-TEQ3	pg/L		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HpCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,6,7,8-HpCDF I-TEQ2	pg/L		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,6,7,8-HpCDF I-TEQ3	pg/L		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF	pg/L	20	<20	<20   <20   [NA]	99.8	69.1
<i>Surrogate 13C-1,2,3,4,7,8,9-HpCDF</i>	%		96.3	87.4 / 89.2	95.8	78.9
1,2,3,4,7,8,9-HpCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF WHO-TEQ2	pg/L		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF WHO-TEQ3	pg/L		0.200	0.200   0.200   0.00	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF I-TEQ2	pg/L		0.100	0.100   0.100   0.00	[NA]	[NA]
1,2,3,4,7,8,9-HpCDF I-TEQ3	pg/L		0.200	0.200   0.200   0.00	[NA]	[NA]
OCDF	pg/L	50	<50	<50   <50   [NA]	101	66.2
OCDF WHO-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
OCDF WHO-TEQ2	pg/L		0.00750	0.00750   0.00750   0.00	[NA]	[NA]
OCDF WHO-TEQ3	pg/L		0.0150	0.0150   0.0150   0.00	[NA]	[NA]
OCDF I-TEQ1	pg/L		0.00	0.00   0.00   [NA]	[NA]	[NA]
OCDF I-TEQ2	pg/L		0.0250	0.0250   0.0250   0.00	[NA]	[NA]
OCDF I-TEQ3	pg/L		0.0500	0.0500   0.0500   0.00	[NA]	[NA]



# CHAIN OF CUSTODY FORM - Client

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Company:	Reditus			Client Project Name/Number/Site etc (ie report title):	22148					
Contact Person:	Kyle Sier			PO No. (if applicable):	22148					
Project Mgr:	Toby Scrivener			Envirolab Quote No.:						
Sampler:	Kyle Sier			Date results required:	Microbiological TAT					
Address:	29 - 33 Warratah St, Kirrawee NSW			Or choose:	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	
Phone:	-	Mob:	0478 117 515	Note: Inform lab in advance if urgent turnaround is required - surcharges apply						
Email Results to:	kyle.sier@reditus.com.au toby.scrivener@reditus.com.au			Additional report format:	<input type="checkbox"/> Esdat		<input type="checkbox"/> Equis			
Email Invoice to:	accounts@reditusconsulting.com			Lab Comments:						

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Darwin Office - Envirolab Services  
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08 8967 1201 | <darwin@envirolab.com.au

Sample Information					Tests Required						Comments		
Envirolab Sample ID (Lab use only)	Client Sample ID or Information	Depth	Date Sampled	Type of Sample	Faecal Coliforms	Faecal Streptococci	E. Coli	HOLD					Provide as much information about the sample as you can
1	MW4-B	-	31/5/21	Water				X					
2	MW1-B	-						X					
3	MW2-B	-						X					
4	EW1	-						X					
5	RMW05	-			X	X	X						
6	DUP1	-						X					
7	TRIP1	-						X					
8	DUP42	-						X					
9	TRIP2	-						X					
10	BLANK	-						X					

Please tick the box if observed settled sediment present in water samples is to be included in the extraction and/or analysis

Relinquished by (Company): Reditus Consulting	Received by (Company): PLXND	Lab Use Only	
Print Name: Kyle Sier	Print Name: GINNAR	Job number: 352854	Cooling: Ice / Ice pack / None
Date & Time: 31/5/21 → 5:59 pm	Date & Time: 31/5/21 1800	Temperature: 10°C	Security seal: Intact / Broken / None
Signature:	Signature:	TAT Req - SAME day / 1 / 2 / 3 / 4 / STD	

## CERTIFICATE OF ANALYSIS 352854

### **Client Details**

<b>Client</b>	Reditus Consulting
<b>Attention</b>	Toby Scrivener, Kyle Sier
<b>Address</b>	Shop 1, 29-33 Waratah St, KIRRRAWEE, NSW, 2232

### **Sample Details**

<b>Your Reference</b>	<b><u>22148</u></b>
<b>Number of Samples</b>	10 Water
<b>Date samples received</b>	31/05/2024
<b>Date completed instructions received</b>	31/05/2024

### **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.  
Samples were analysed as received from the client. Results relate specifically to the samples as received.  
Results are reported on a dry weight basis for solids and on an as received basis for other matrices.  
**Please refer to the last page of this report for any comments relating to the results.**

### **Report Details**

<b>Date results requested by</b>	07/06/2024
<b>Date of Issue</b>	04/06/2024
NATA Accreditation Number 2901. This document shall not be reproduced except in full.	
Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	

### **Results Approved By**

Stuart Chen, Asbestos Approved Identifier/Report coordinator

### **Authorised By**

Nancy Zhang, Laboratory Manager

<b>Microbiologocal Testing</b>		
Our Reference		352854-5
Your Reference	UNITS	RMW05
Date Sampled		31/05/2024
Type of sample		Water
Date of testing	-	01/06/2024
Faecal Coliforms	cfu/100mL	80
E. coli	cfu/100mL	80
Enterococci	cfu/100mL	<1

<b>Method ID</b>	<b>Methodology Summary</b>
<b>Ext-008</b>	Subcontracted to Sonic Food & Water Testing. NATA Accreditation No. 4034.

**Result Definitions**

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<	Less than
>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	
The recommended maximums for analytes in urine are taken from "2018 TLVs and BEIs", as published by ACGIH (where available). Limit provided for Nickel is a precautionary guideline as per Position Paper prepared by AIOP Exposure Standards Committee, 2016.	
Guideline limits for Rinse Water Quality reported as per analytical requirements and specifications of AS 4187, Amdt 2 2019, Table 7.2	

## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Where matrix spike recoveries fall below the lower limit of the acceptance criteria (e.g. for non-labile or standard Organics <60%), positive result(s) in the parent sample will subsequently have a higher than typical estimated uncertainty (MU estimates supplied on request) and in these circumstances the sample result is likely biased significantly low.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

## **Report Comments**

Microbiology analysed by Sonic Food & Water Testing. Report no. W2413062

The time between collection and the commencement of testing should not exceed 24 hours. Samples tested outside this time may have their results compromised.

Updated: 1/7/24 0732


**ENVIROLAB**  
 GROUP  
**MPL**

# CHAIN OF CUSTODY FORM - Client

[Copyright and Confidential]

Client: Reditus Consulting Contact Person: Toby Scrivener Project Mgr: Toby Scrivener Sampler: Kyle Sier Address: Unit 1A, Level 1, 29-33 Waratah Street Kirrawee NSW  Phone: 02 9521 6567      Mob: 0478117515  Email Invoice To: accounts@reditusconsulting.com tobyscrivener@reditus.com.au  Email Results To: tobyscrivener@reditus.com.au, kylesier@reditusconsulting.com reditusconsulting@esdat.com.au					Client Project Name/Number/Site etc (ie report title):  22148  PO No.: 22148  Envirolab Quote No.:  Date results required: standard  Or choose: standard / same day / 1 day / 2 day / 3 day Note: Inform lab in advance if urgent turnaround is required - surcharges apply  Additional report format: esdat  Lab Comments:								
Sample Information					Tests Required					Comments			
Envirolab Sample ID	Client Sample ID or Information	Depth	Date sampled	Type of sample	Combo 3	PFAS Short Suite (Trace)	Cations	Anions	Ionic Balance	Nutrients	Dioxins & Furans	Provide as much information about the sample as you can  RMW01 RMW02 RMW03 RMW04 RMW05 MW1-B MW2-B MW3-B MW4-B EW1 DUP1 TRIP1 DUP2 TRIP2 BLANK	
		-	30/05/2024	Water	X	X	X	X	X	X	X		
		-	30/05/2024	Water	X		X	X	X	X			
		-	30/05/2024	Water	X		X	X	X	X			
		-	30/05/2024	Water	X	X	X	X	X	X	X		
S		-	31/05/2024	Water	X		X	X	X	X			
2		-	31/05/2024	Water	X	X							
3		-	31/05/2024	Water	X	X							
			30/05/2024	Water	X	X					X		
1		-	31/05/2024	Water	X	X							
4		-	31/05/2024	Water	X	X							
6		-	31/05/2024	Water	X	X							
7		-	31/05/2024	Water	X	X							
8		-	31/05/2024	Water		X							
9		-	31/05/2024	Water		X							
10		-	31/05/2024	Water	X	X							
					Sample Count	13	12	5	5	5	3		
<input type="checkbox"/> Please tick the box if observed settled sediment present in water samples is to be included in the extraction and/or analysis													
Relinquished by (Company): Reditus		Received by (Company): El J MD			Lab Use Only								
Print Name: Kyle Sier		Print Name: O. W.			Job number: 352854-A Cooling: Ice / Ice pack / None								
Date & Time: 31/05/2024		Date & Time: 31/5/24 1800			Temperature: 10 Security seal: Intact / Broken / None								
Signature: K.Sier		Signature: OW			TAT Req - SAME day / 1 / 2 / 3 / 4 / STD								

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**Anna Bui**

---

**From:** tobyscrivener@reditusconsulting.com  
**Sent:** Monday, 3 June 2024 10:41 AM  
**To:** 'Kyle Sier'; Samplereceipt  
**Subject:** RE: COC - 22148

**CAUTION:** This email originated from outside of the organisation. Do not act on instructions, click links or open attachments unless you recognise the sender and know the content is authentic and safe.

Hi all – further to Kyle's email (below), could I please also add the following to samples RMW01, RMW02, RMW03, RMW04 and RMW05:

5

- Dissolved –
  - Aluminium (Al),
  - antimony (Sb),
  - arsenic (As),
  - barium (Ba),
  - beryllium (Be),
  - boron (B),
  - cadmium (Cd),
  - chromium (Cr),
  - cobalt (Co),
  - copper (Cu),
  - iron (Fe),
  - lead (Pb),
  - lithium (Li),
  - manganese (Mn),
  - mercury (Hg),
  - molybdenum (Mo),
  - nickel (Ni),
  - selenium (Se),
  - silica (dissolved SiO<sub>2</sub>),
  - silver (Ag),
  - strontium (Sr),
  - uranium (U),
  - vanadium (V),
  - zinc (Zn)
- Alkalinity (bicarbonate, carbonate, hydroxide and total)
- Total dissolved solids (TDS), total hardness

Thanks,

Regards,

352854-A

**Toby Scrivener**  
Principal Environmental Engineer  
EIANZ Certified Environmental Practitioner - Site Contamination Specialist



Unit 1A, 29-33 Waratah Street  
Kirrawee NSW 2232

## CERTIFICATE OF ANALYSIS 352854-A

### **Client Details**

<b>Client</b>	Reditus Consulting
<b>Attention</b>	Toby Scrivener
<b>Address</b>	Shop 1, 29-33 Waratah St, KIRRRAWEE, NSW, 2232

### **Sample Details**

<b>Your Reference</b>	<b><u>22148</u></b>
<b>Number of Samples</b>	Additional analysis
<b>Date samples received</b>	31/05/2024
<b>Date completed instructions received</b>	03/06/2024

### **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.  
Samples were analysed as received from the client. Results relate specifically to the samples as received.  
Results are reported on a dry weight basis for solids and on an as received basis for other matrices.  
**Please refer to the last page of this report for any comments relating to the results.**

### **Report Details**

<b>Date results requested by</b>	11/06/2024
<b>Date of Issue</b>	11/06/2024
NATA Accreditation Number 2901. This document shall not be reproduced except in full.	
Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	

#### **Results Approved By**

Diego Bigolin, Inorganics Supervisor  
Dragana Tomas, Senior Chemist  
Giovanni Agosti, Group Technical Manager  
Jack Wallis, Chemist (FAS)  
Loren Bardwell, Development Chemist  
Sean McAlary, Chemist (FAS)

#### **Authorised By**

Nancy Zhang, Laboratory Manager

vTRH(C6-C10)/BTEXN in Water						
Our Reference	UNITS	352854-A-1	352854-A-2	352854-A-3	352854-A-4	352854-A-5
Your Reference		MW4-B	MW1-B	MW2-B	EW1	RMW05
Date Sampled		31/05/2024	31/05/2024	31/05/2024	31/05/2024	31/05/2024
Type of sample		Water	Water	Water	Water	Water
Date extracted	-	11/06/2024	11/06/2024	11/06/2024	11/06/2024	11/06/2024
Date analysed	-	11/06/2024	11/06/2024	11/06/2024	11/06/2024	11/06/2024
TRH C <sub>6</sub> - C <sub>9</sub>	µg/L	47	<10	<10	<10	<10
TRH C <sub>6</sub> - C <sub>10</sub>	µg/L	50	<10	<10	<10	<10
TRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	µg/L	50	<10	<10	<10	<10
Benzene	µg/L	<1	<1	<1	<1	<1
Toluene	µg/L	<1	<1	<1	<1	<1
Ethylbenzene	µg/L	<1	<1	<1	<1	<1
m+p-xylene	µg/L	<2	<2	<2	<2	<2
o-xylene	µg/L	<1	<1	<1	<1	<1
Naphthalene	µg/L	<1	<1	<1	<1	<1
Surrogate Dibromofluoromethane	%	106	112	112	111	112
Surrogate Toluene-d8	%	90	94	94	94	94
Surrogate 4-Bromofluorobenzene	%	105	107	107	106	106

vTRH(C6-C10)/BTEXN in Water			
Our Reference	UNITS	352854-A-6	352854-A-10
Your Reference		DUP1	BLANK
Date Sampled		31/05/2024	31/05/2024
Type of sample		Water	Water
Date extracted	-	11/06/2024	11/06/2024
Date analysed	-	11/06/2024	11/06/2024
TRH C <sub>6</sub> - C <sub>9</sub>	µg/L	<10	<10
TRH C <sub>6</sub> - C <sub>10</sub>	µg/L	<10	<10
TRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	µg/L	<10	<10
Benzene	µg/L	<1	<1
Toluene	µg/L	<1	<1
Ethylbenzene	µg/L	<1	<1
m+p-xylene	µg/L	<2	<2
o-xylene	µg/L	<1	<1
Naphthalene	µg/L	<1	<1
Surrogate Dibromofluoromethane	%	112	112
Surrogate Toluene-d8	%	93	94
Surrogate 4-Bromofluorobenzene	%	107	107

svTRH (C10-C40) in Water						
Our Reference	UNITS	352854-A-1	352854-A-2	352854-A-3	352854-A-4	352854-A-5
Your Reference		MW4-B	MW1-B	MW2-B	EW1	RMW05
Date Sampled		31/05/2024	31/05/2024	31/05/2024	31/05/2024	31/05/2024
Type of sample		Water	Water	Water	Water	Water
Date extracted	-	04/06/2024	04/06/2024	04/06/2024	04/06/2024	04/06/2024
Date analysed	-	04/06/2024	04/06/2024	04/06/2024	04/06/2024	04/06/2024
TRH C <sub>10</sub> - C <sub>14</sub>	µg/L	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	µg/L	<100	<100	<100	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	µg/L	<100	<100	<100	<100	<100
Total +ve TRH (C10-C36)	µg/L	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub>	µg/L	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	µg/L	<50	<50	<50	<50	<50
TRH >C <sub>16</sub> - C <sub>34</sub>	µg/L	<100	<100	<100	<100	<100
TRH >C <sub>34</sub> - C <sub>40</sub>	µg/L	<100	<100	<100	<100	<100
Total +ve TRH (>C10-C40)	µg/L	<50	<50	<50	<50	<50
Surrogate o-Terphenyl	%	61	93	77	80	75

svTRH (C10-C40) in Water			
Our Reference	UNITS	352854-A-6	352854-A-10
Your Reference		DUP1	BLANK
Date Sampled		31/05/2024	31/05/2024
Type of sample		Water	Water
Date extracted	-	11/06/2024	04/06/2024
Date analysed	-	11/06/2024	04/06/2024
TRH C <sub>10</sub> - C <sub>14</sub>	µg/L	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	µg/L	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	µg/L	<100	<100
Total +ve TRH (C10-C36)	µg/L	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub>	µg/L	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	µg/L	<50	<50
TRH >C <sub>16</sub> - C <sub>34</sub>	µg/L	<100	<100
TRH >C <sub>34</sub> - C <sub>40</sub>	µg/L	<100	<100
Total +ve TRH (>C10-C40)	µg/L	<50	<50
Surrogate o-Terphenyl	%	76	71

PAHs in Water						
Our Reference	UNITS	352854-A-1	352854-A-2	352854-A-3	352854-A-4	352854-A-5
Your Reference		MW4-B	MW1-B	MW2-B	EW1	RMW05
Date Sampled		31/05/2024	31/05/2024	31/05/2024	31/05/2024	31/05/2024
Type of sample		Water	Water	Water	Water	Water
Date extracted	-	04/06/2024	04/06/2024	04/06/2024	04/06/2024	04/06/2024
Date analysed	-	04/06/2024	04/06/2024	04/06/2024	04/06/2024	04/06/2024
Naphthalene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthylene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Fluorene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Phenanthrene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Anthracene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Fluoranthene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Pyrene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(a)anthracene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Chrysene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(b,j+k)fluoranthene	µg/L	<0.2	<0.2	<0.2	<0.2	<0.2
Benzo(a)pyrene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Indeno(1,2,3-c,d)pyrene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Dibenzo(a,h)anthracene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(g,h,i)perylene	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Benzo(a)pyrene TEQ	µg/L	<0.5	<0.5	<0.5	<0.5	<0.5
Total +ve PAH's	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate p-Terphenyl-d14	%	65	83	74	83	82

PAHs in Water			
Our Reference	UNITS	352854-A-6	352854-A-10
Your Reference		DUP1	BLANK
Date Sampled		31/05/2024	31/05/2024
Type of sample		Water	Water
Date extracted	-	04/06/2024	04/06/2024
Date analysed	-	04/06/2024	04/06/2024
Naphthalene	µg/L	<0.1	<0.1
Acenaphthylene	µg/L	<0.1	<0.1
Acenaphthene	µg/L	<0.1	<0.1
Fluorene	µg/L	<0.1	<0.1
Phenanthrene	µg/L	<0.1	<0.1
Anthracene	µg/L	<0.1	<0.1
Fluoranthene	µg/L	<0.1	<0.1
Pyrene	µg/L	<0.1	<0.1
Benzo(a)anthracene	µg/L	<0.1	<0.1
Chrysene	µg/L	<0.1	<0.1
Benzo(b,j+k)fluoranthene	µg/L	<0.2	<0.2
Benzo(a)pyrene	µg/L	<0.1	<0.1
Indeno(1,2,3-c,d)pyrene	µg/L	<0.1	<0.1
Dibenzo(a,h)anthracene	µg/L	<0.1	<0.1
Benzo(g,h,i)perylene	µg/L	<0.1	<0.1
Benzo(a)pyrene TEQ	µg/L	<0.5	<0.5
Total +ve PAH's	µg/L	<0.1	<0.1
Surrogate p-Terphenyl-d14	%	76	70

All metals in water-dissolved						
Our Reference	UNITS	352854-A-1	352854-A-2	352854-A-3	352854-A-4	352854-A-5
Your Reference		MW4-B	MW1-B	MW2-B	EW1	RMW05
Date Sampled		31/05/2024	31/05/2024	31/05/2024	31/05/2024	31/05/2024
Type of sample		Water	Water	Water	Water	Water
Date prepared	-	05/06/2024	05/06/2024	05/06/2024	05/06/2024	05/06/2024
Date analysed	-	05/06/2024	05/06/2024	05/06/2024	05/06/2024	05/06/2024
Aluminium-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	60
Antimony-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	<1
Arsenic-Dissolved	µg/L	<1	2	2	<1	<1
Barium-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	50
Beryllium-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	3
Boron-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	60
Cadmium-Dissolved	µg/L	<0.1	<0.1	<0.1	<0.1	<0.1
Chromium-Dissolved	µg/L	<1	<1	<1	<1	<1
Cobalt-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	91
Copper-Dissolved	µg/L	2	<1	<1	<1	<1
Iron-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	48,000
Lead-Dissolved	µg/L	<1	<1	<1	<1	<1
Lithium-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	36
Manganese-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	4,300
Mercury-Dissolved	µg/L	<0.05	<0.05	<0.05	<0.05	<0.05
Molybdenum-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	<1
Nickel-Dissolved	µg/L	<1	2	25	120	94
Selenium-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	<1
Silver-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	<1
Strontium-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	88
Uranium-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	<0.5
Vanadium-Dissolved	µg/L	[NA]	[NA]	[NA]	[NA]	<1
Zinc-Dissolved	µg/L	7	9	4	68	400

<b>All metals in water-dissolved</b>			
Our Reference		352854-A-6	352854-A-10
Your Reference	UNITS	DUP1	BLANK
Date Sampled		31/05/2024	31/05/2024
Type of sample		Water	Water
Date prepared	-	05/06/2024	05/06/2024
Date analysed	-	05/06/2024	05/06/2024
Arsenic-Dissolved	µg/L	2	<1
Cadmium-Dissolved	µg/L	<0.1	<0.1
Chromium-Dissolved	µg/L	<1	<1
Copper-Dissolved	µg/L	<1	<1
Lead-Dissolved	µg/L	<1	<1
Mercury-Dissolved	µg/L	<0.05	<0.05
Nickel-Dissolved	µg/L	2	<1
Zinc-Dissolved	µg/L	6	<1

<b>Ion Balance</b>		
Our Reference		352854-A-5
Your Reference	UNITS	RMW05
Date Sampled		31/05/2024
Type of sample		Water
Date prepared	-	07/06/2024
Date analysed	-	07/06/2024
Calcium - Dissolved	mg/L	10
Potassium - Dissolved	mg/L	7.8
Sodium - Dissolved	mg/L	850
Magnesium - Dissolved	mg/L	120
Hardness (calc) equivalent CaCO <sub>3</sub>	mg/L	520
Hydroxide Alkalinity (OH <sup>-</sup> ) as CaCO <sub>3</sub>	mg/L	<5
Bicarbonate Alkalinity as CaCO <sub>3</sub>	mg/L	75
Carbonate Alkalinity as CaCO <sub>3</sub>	mg/L	<5
Total Alkalinity as CaCO <sub>3</sub>	mg/L	75
Sulphate, SO <sub>4</sub>	mg/L	310
Chloride, Cl	mg/L	1,500
Ionic Balance	%	-3.0

<b>Miscellaneous Inorganics</b>		
Our Reference	UNITS	352854-A-5
Your Reference		RMW05
Date Sampled		31/05/2024
Type of sample		Water
Date prepared	-	03/06/2024
Date analysed	-	03/06/2024
Total Dissolved Solids (grav)	mg/L	3,100
Ammonia as N in water	mg/L	0.016
Nitrate as N in water	mg/L	0.31
Nitrite as N in water	mg/L	0.021
NOx as N in water	mg/L	0.3
Total Nitrogen in water	mg/L	0.5
TKN in water	mg/L	0.2
Phosphate as P in water	mg/L	<0.005
Organic Nitrogen as N	mg/L	<0.2

<b>Metals in Waters - Acid extractable</b>		
Our Reference		352854-A-5
Your Reference	UNITS	RMW05
Date Sampled		31/05/2024
Type of sample		Water
Date prepared	-	04/06/2024
Date analysed	-	04/06/2024
Phosphorus - Total	mg/L	<0.05

<b>Metals in Water - Dissolved</b>		
Our Reference		352854-A-5
Your Reference	UNITS	RMW05
Date Sampled		31/05/2024
Type of sample		Water
Date digested	-	04/06/2024
Date analysed	-	04/06/2024
Silicon*- Dissolved	mg/L	11

PFAS in Water TRACE Short						
Our Reference	UNITS	352854-A-1	352854-A-2	352854-A-3	352854-A-4	352854-A-6
Your Reference		MW4-B	MW1-B	MW2-B	EW1	DUP1
Date Sampled		31/05/2024	31/05/2024	31/05/2024	31/05/2024	31/05/2024
Type of sample		Water	Water	Water	Water	Water
Date prepared	-	05/06/2024	05/06/2024	05/06/2024	05/06/2024	05/06/2024
Date analysed	-	05/06/2024	05/06/2024	05/06/2024	05/06/2024	05/06/2024
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.0039	0.0081	0.001	<0.0002	0.0074
Perfluorooctanesulfonic acid PFOS	µg/L	0.013	0.001	0.002	<0.0002	0.001
Perfluorooctanoic acid PFOA	µg/L	0.018	0.0023	0.0020	<0.0002	0.0021
6:2 FTS	µg/L	<0.0004	0.0007	0.0007	<0.0004	0.0005
8:2 FTS	µg/L	<0.0004	<0.0004	<0.0004	<0.0004	<0.0004
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%	92	114	97	107	95
Surrogate <sup>13</sup> C <sub>2</sub> PFOA	%	89	91	95	92	93
Extracted ISTD <sup>18</sup> O <sub>2</sub> PFHxS	%	84	70	83	81	79
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%	89	71	84	69	82
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOA	%	96	91	98	105	98
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%	125	102	157	94	156
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%	164	92	164	120	186
Total Positive PFHxS & PFOS	µg/L	0.017	0.0094	0.0032	<0.0002	0.0085
Total Positive PFOS & PFOA	µg/L	0.031	0.0035	0.0038	<0.0002	0.0031
Total Positive PFAS	µg/L	0.035	0.012	0.0060	<0.0002	0.011

PFAS in Water TRACE Short			
Our Reference	UNITS	352854-A-8	352854-A-10
Your Reference		DUP2	BLANK
Date Sampled		31/05/2024	31/05/2024
Type of sample		Water	Water
Date prepared	-	05/06/2024	05/06/2024
Date analysed	-	05/06/2024	05/06/2024
Perfluorohexanesulfonic acid - PFHxS	µg/L	<0.0002	<0.0002
Perfluorooctanesulfonic acid PFOS	µg/L	0.0002	<0.0002
Perfluorooctanoic acid PFOA	µg/L	<0.0002	<0.0002
6:2 FTS	µg/L	<0.0004	<0.0004
8:2 FTS	µg/L	<0.0004	<0.0004
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%	112	97
Surrogate <sup>13</sup> C <sub>2</sub> PFOA	%	90	96
Extracted ISTD <sup>18</sup> O <sub>2</sub> PFHxS	%	80	91
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%	76	84
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOA	%	105	130
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%	103	#
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%	117	#
Total Positive PFHxS & PFOS	µg/L	0.0002	<0.0002
Total Positive PFOS & PFOA	µg/L	0.0002	<0.0002
Total Positive PFAS	µg/L	0.0002	<0.0002

Method ID	Methodology Summary
Inorg-006	Alkalinity - determined titrimetrically in accordance with APHA latest edition, 2320-B.
Inorg-018	Total Dissolved Solids - determined gravimetrically. The solids are dried at 180+/-10°C.  NOTE: Where the EC of the sample is <100µS/cm, the TDS will typically be below 70mg/L (as the sample is very likely to be at least drinking water quality). Therefore to ensure data quality for TDS, the TDS is typically calculated as per the equation below:-  $\text{TDS} = \text{EC} * 0.6$
Inorg-040	The concentrations of the major ions (mg/L) are converted to milliequivalents and summed. The ionic balance should be within +/- 15% ie total anions = total cations +/-15%.
Inorg-055	Nitrate - determined colourimetrically. Waters samples are filtered on receipt prior to analysis. Soils are analysed following a water extraction.
Inorg-055	Nitrite - determined colourimetrically based on APHA latest edition NO2- B. Waters samples are filtered on receipt prior to analysis. Soils are analysed following a water extraction.
Inorg-055/062/127	Total Nitrogen - Calculation sum of TKN and oxidised Nitrogen. Alternatively analysed by combustion and chemiluminescence.
Inorg-057	Ammonia - determined colourimetrically, based on APHA latest edition 4500-NH3 F. Waters samples are filtered on receipt prior to analysis. Soils are analysed following a KCl extraction.
Inorg-060	Phosphate determined colourimetrically based on EPA365.1 and APHA latest edition 4500 P E. Waters samples are filtered on receipt prior to analysis. Soils are analysed following a water extraction.
Inorg-062	TKN - determined colourimetrically based on APHA latest edition 4500 Norg. Alternatively, TKN can be derived from calculation (Total N - NOx).
Inorg-081	Anions - a range of Anions are determined by Ion Chromatography, in accordance with APHA latest edition, 4110-B. Waters samples are filtered on receipt prior to analysis. Alternatively determined by colourimetry/turbidity using Discrete Analyser.
Metals-020	Determination of various metals by ICP-AES.
Metals-021	Determination of Mercury by Cold Vapour AAS.
Metals-022	Determination of various metals by ICP-MS.  Please note for Bromine and Iodine, any forms of these elements that are present are included together in the one result reported for each of these two elements.  Salt forms (e.g. FeO, PbO, ZnO) are determined stoichiometrically from the base metal concentration.
Org-020	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.

Method ID	Methodology Summary
Org-022/025	<p>Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS/GC-MSMS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013.</p>
Org-023	Water samples are analysed directly by purge and trap GC-MS.
Org-023	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.
Org-029	<p>Soil samples are extracted with basified Methanol. Waters and soil extracts are directly injected and/or concentrated/extracted using SPE. TCLPs/ASLP leachates are centrifuged, the supernatant is then analysed (including amendment with solvent) - as per the option in AS4439.3.</p> <p>Analysis is undertaken with LC-MS/MS.</p> <p>PFAS results include the sum of branched and linear isomers where applicable.</p> <p>Please note that PFAS results are corrected for Extracted Internal Standards (QSM 5.4 Table B-15 terminology), which are mass labelled analytes added prior to sample preparation to assess matrix effects and verify processing of the sample. PFAS analytes without a commercially available mass labelled analogue are corrected vs a closely eluting mass labelled PFAS compound. Surrogates are also reported, in this context they are mass labelled PFAS compounds added prior to extraction but are used as monitoring compounds only (not used for result correction). Envicarb (or similar) is used discretionally to remove interfering matrix components.</p> <p>Please contact the laboratory if estimates of Measurement Uncertainty are required as per WA DER.</p>

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Water							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date extracted	-			11/06/2024	[NT]	[NT]	[NT]	[NT]	11/06/2024	[NT]
Date analysed	-			11/06/2024	[NT]	[NT]	[NT]	[NT]	11/06/2024	[NT]
TRH C <sub>6</sub> - C <sub>9</sub>	µg/L	10	Org-023	<10	[NT]	[NT]	[NT]	[NT]	94	[NT]
TRH C <sub>6</sub> - C <sub>10</sub>	µg/L	10	Org-023	<10	[NT]	[NT]	[NT]	[NT]	94	[NT]
Benzene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	103	[NT]
Toluene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	89	[NT]
Ethylbenzene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	86	[NT]
m+p-xylene	µg/L	2	Org-023	<2	[NT]	[NT]	[NT]	[NT]	97	[NT]
o-xylene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	93	[NT]
Naphthalene	µg/L	1	Org-023	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate Dibromofluoromethane	%		Org-023	107	[NT]	[NT]	[NT]	[NT]	123	[NT]
Surrogate Toluene-d8	%		Org-023	95	[NT]	[NT]	[NT]	[NT]	94	[NT]
Surrogate 4-Bromofluorobenzene	%		Org-023	107	[NT]	[NT]	[NT]	[NT]	98	[NT]

QUALITY CONTROL: svTRH (C10-C40) in Water							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	352854-A-2
Date extracted	-			04/06/2024	1	04/06/2024	04/06/2024		04/06/2024	04/06/2024
Date analysed	-			04/06/2024	1	04/06/2024	04/06/2024		04/06/2024	04/06/2024
TRH C <sub>10</sub> - C <sub>14</sub>	µg/L	50	Org-020	<50	1	<50	<50	0	114	97
TRH C <sub>15</sub> - C <sub>28</sub>	µg/L	100	Org-020	<100	1	<100	<100	0	104	98
TRH C <sub>29</sub> - C <sub>36</sub>	µg/L	100	Org-020	<100	1	<100	<100	0	100	82
TRH >C <sub>10</sub> - C <sub>16</sub>	µg/L	50	Org-020	<50	1	<50	<50	0	114	97
TRH >C <sub>16</sub> - C <sub>34</sub>	µg/L	100	Org-020	<100	1	<100	<100	0	104	98
TRH >C <sub>34</sub> - C <sub>40</sub>	µg/L	100	Org-020	<100	1	<100	<100	0	100	82
Surrogate o-Terphenyl	%		Org-020	89	1	61	70	14	88	99

QUALITY CONTROL: PAHs in Water					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	352854-A-2
Date extracted	-			04/06/2024	1	04/06/2024	04/06/2024		04/06/2024	04/06/2024
Date analysed	-			04/06/2024	1	04/06/2024	04/06/2024		04/06/2024	04/06/2024
Naphthalene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	109	118
Acenaphthylene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Acenaphthene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	115	118
Fluorene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	108	118
Phenanthrene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	110	102
Anthracene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Fluoranthene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	105	100
Pyrene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	103	100
Benzo(a)anthracene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Chrysene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	103	103
Benzo(b,j+k)fluoranthene	µg/L	0.2	Org-022/025	<0.2	1	<0.2	<0.2	0	[NT]	[NT]
Benzo(a)pyrene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	91	94
Indeno(1,2,3-c,d)pyrene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Dibenzo(a,h)anthracene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Benzo(g,h,i)perylene	µg/L	0.1	Org-022/025	<0.1	1	<0.1	<0.1	0	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-022/025	85	1	65	75	14	81	75

QUALITY CONTROL: All metals in water-dissolved					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W4	[NT]
Date prepared	-			05/06/2024	1	05/06/2024	05/06/2024		05/06/2024	[NT]
Date analysed	-			05/06/2024	1	05/06/2024	05/06/2024		05/06/2024	[NT]
Aluminium-Dissolved	µg/L	10	Metals-022	<10	[NT]	[NT]	[NT]	[NT]	96	[NT]
Antimony-Dissolved	µg/L	1	Metals-022	<1	[NT]	[NT]	[NT]	[NT]	94	[NT]
Arsenic-Dissolved	µg/L	1	Metals-022	<1	1	<1	[NT]		101	[NT]
Barium-Dissolved	µg/L	1	Metals-022	<1	[NT]	[NT]	[NT]	[NT]	107	[NT]
Beryllium-Dissolved	µg/L	0.5	Metals-022	<0.5	[NT]	[NT]	[NT]	[NT]	83	[NT]
Boron-Dissolved	µg/L	20	Metals-022	<20	[NT]	[NT]	[NT]	[NT]	112	[NT]
Cadmium-Dissolved	µg/L	0.1	Metals-022	<0.1	1	<0.1	[NT]		98	[NT]
Chromium-Dissolved	µg/L	1	Metals-022	<1	1	<1	[NT]		98	[NT]
Cobalt-Dissolved	µg/L	1	Metals-022	<1	[NT]	[NT]	[NT]	[NT]	103	[NT]
Copper-Dissolved	µg/L	1	Metals-022	<1	1	2	[NT]		98	[NT]
Iron-Dissolved	µg/L	10	Metals-022	<10	[NT]	[NT]	[NT]	[NT]	99	[NT]
Lead-Dissolved	µg/L	1	Metals-022	<1	1	<1	[NT]		98	[NT]
Lithium-Dissolved	µg/L	1	Metals-022	<1	[NT]	[NT]	[NT]	[NT]	92	[NT]
Manganese-Dissolved	µg/L	5	Metals-022	<5	[NT]	[NT]	[NT]	[NT]	97	[NT]
Mercury-Dissolved	µg/L	0.05	Metals-021	<0.05	1	<0.05	<0.05	0	101	[NT]
Molybdenum-Dissolved	µg/L	1	Metals-022	<1	[NT]	[NT]	[NT]	[NT]	99	[NT]
Nickel-Dissolved	µg/L	1	Metals-022	<1	1	<1	[NT]		101	[NT]
Selenium-Dissolved	µg/L	1	Metals-022	<1	[NT]	[NT]	[NT]	[NT]	98	[NT]
Silver-Dissolved	µg/L	1	Metals-022	<1	[NT]	[NT]	[NT]	[NT]	104	[NT]
Strontium-Dissolved	µg/L	1	Metals-022	<1	[NT]	[NT]	[NT]	[NT]	99	[NT]
Uranium-Dissolved	µg/L	0.5	Metals-022	<0.5	[NT]	[NT]	[NT]	[NT]	91	[NT]
Vanadium-Dissolved	µg/L	1	Metals-022	<1	[NT]	[NT]	[NT]	[NT]	99	[NT]
Zinc-Dissolved	µg/L	1	Metals-022	<1	1	7	[NT]		98	[NT]

QUALITY CONTROL: Ion Balance						Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]	
Date prepared	-			07/06/2024	[NT]	[NT]	[NT]	[NT]	07/06/2024	[NT]	
Date analysed	-			07/06/2024	[NT]	[NT]	[NT]	[NT]	07/06/2024	[NT]	
Calcium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	105	[NT]	
Potassium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	98	[NT]	
Sodium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	101	[NT]	
Magnesium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	102	[NT]	
Hydroxide Alkalinity (OH <sup>-</sup> ) as CaCO <sub>3</sub>	mg/L	5	Inorg-006	<5	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]	
Bicarbonate Alkalinity as CaCO <sub>3</sub>	mg/L	5	Inorg-006	<5	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]	
Carbonate Alkalinity as CaCO <sub>3</sub>	mg/L	5	Inorg-006	<5	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]	
Total Alkalinity as CaCO <sub>3</sub>	mg/L	5	Inorg-006	<5	[NT]	[NT]	[NT]	[NT]	109	[NT]	
Sulphate, SO <sub>4</sub>	mg/L	1	Inorg-081	<1	[NT]	[NT]	[NT]	[NT]	105	[NT]	
Chloride, Cl	mg/L	1	Inorg-081	<1	[NT]	[NT]	[NT]	[NT]	100	[NT]	

QUALITY CONTROL: Miscellaneous Inorganics					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			03/06/2024	[NT]	[NT]	[NT]	[NT]	03/06/2024	[NT]
Date analysed	-			03/06/2024	[NT]	[NT]	[NT]	[NT]	03/06/2024	[NT]
Total Dissolved Solids (grav)	mg/L	5	Inorg-018	<5	[NT]	[NT]	[NT]	[NT]	96	[NT]
Ammonia as N in water	mg/L	0.005	Inorg-057	<0.005	[NT]	[NT]	[NT]	[NT]	92	[NT]
Nitrate as N in water	mg/L	0.005	Inorg-055	<0.005	[NT]	[NT]	[NT]	[NT]	97	[NT]
Nitrite as N in water	mg/L	0.005	Inorg-055	<0.005	[NT]	[NT]	[NT]	[NT]	99	[NT]
NOx as N in water	mg/L	0.005	Inorg-055	<0.005	[NT]	[NT]	[NT]	[NT]	97	[NT]
Total Nitrogen in water	mg/L	0.1	Inorg-055/062/127	<0.1	[NT]	[NT]	[NT]	[NT]	91	[NT]
TKN in water	mg/L	0.1	Inorg-062	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Phosphate as P in water	mg/L	0.005	Inorg-060	<0.005	[NT]	[NT]	[NT]	[NT]	99	[NT]
Organic Nitrogen as N	mg/L	0.2	Inorg-055/062/127	<0.2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]

QUALITY CONTROL: Metals in Waters - Acid extractable							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			04/06/2024	[NT]	[NT]	[NT]	[NT]	04/06/2024	[NT]
Date analysed	-			04/06/2024	[NT]	[NT]	[NT]	[NT]	04/06/2024	[NT]
Phosphorus - Total	mg/L	0.05	Metals-020	<0.05	[NT]	[NT]	[NT]	[NT]	96	[NT]

QUALITY CONTROL: Metals in Water - Dissolved							Duplicate		Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date digested	-			04/06/2024	[NT]	[NT]	[NT]	[NT]	04/06/2024	[NT]
Date analysed	-			04/06/2024	[NT]	[NT]	[NT]	[NT]	04/06/2024	[NT]
Silicon*- Dissolved	mg/L	0.2	Metals-020	<0.2	[NT]	[NT]	[NT]	[NT]	96	[NT]

QUALITY CONTROL: PFAS in Water TRACE Short						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			05/06/2024	[NT]	[NT]	[NT]	[NT]	05/06/2024	[NT]
Date analysed	-			05/06/2024	[NT]	[NT]	[NT]	[NT]	05/06/2024	[NT]
Perfluorohexanesulfonic acid - PFHxS	µg/L	0.0002	Org-029	<0.0002	[NT]	[NT]	[NT]	[NT]	94	[NT]
Perfluorooctanesulfonic acid PFOS	µg/L	0.0002	Org-029	<0.0002	[NT]	[NT]	[NT]	[NT]	102	[NT]
Perfluorooctanoic acid PFOA	µg/L	0.0002	Org-029	<0.0002	[NT]	[NT]	[NT]	[NT]	100	[NT]
6:2 FTS	µg/L	0.0004	Org-029	<0.0004	[NT]	[NT]	[NT]	[NT]	91	[NT]
8:2 FTS	µg/L	0.0004	Org-029	<0.0004	[NT]	[NT]	[NT]	[NT]	100	[NT]
Surrogate <sup>13</sup> C <sub>8</sub> PFOS	%		Org-029	98	[NT]	[NT]	[NT]	[NT]	102	[NT]
Surrogate <sup>13</sup> C <sub>2</sub> PFOA	%		Org-029	99	[NT]	[NT]	[NT]	[NT]	98	[NT]
Extracted ISTD <sup>18</sup> O <sub>2</sub> PFHxS	%		Org-029	83	[NT]	[NT]	[NT]	[NT]	88	[NT]
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOS	%		Org-029	69	[NT]	[NT]	[NT]	[NT]	74	[NT]
Extracted ISTD <sup>13</sup> C <sub>4</sub> PFOA	%		Org-029	110	[NT]	[NT]	[NT]	[NT]	98	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> 6:2FTS	%		Org-029	99	[NT]	[NT]	[NT]	[NT]	127	[NT]
Extracted ISTD <sup>13</sup> C <sub>2</sub> 8:2FTS	%		Org-029	113	[NT]	[NT]	[NT]	[NT]	149	[NT]

**Result Definitions**

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<	Less than
>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	
The recommended maximums for analytes in urine are taken from "2018 TLVs and BEIs", as published by ACGIH (where available). Limit provided for Nickel is a precautionary guideline as per Position Paper prepared by AIOP Exposure Standards Committee, 2016.	
Guideline limits for Rinse Water Quality reported as per analytical requirements and specifications of AS 4187, Amdt 2 2019, Table 7.2	

## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Where matrix spike recoveries fall below the lower limit of the acceptance criteria (e.g. for non-labile or standard Organics <60%), positive result(s) in the parent sample will subsequently have a higher than typical estimated uncertainty (MU estimates supplied on request) and in these circumstances the sample result is likely biased significantly low.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

## **Report Comments**

NO<sub>2</sub>/NO<sub>3</sub>/PO<sub>4</sub> - out of recommended holding time

Total metals: no unfiltered, preserved sample was received, therefore analysis was conducted from the unpreserved sample bottle.  
Note: there is a possibility some elements may be underestimated.

For PFAS Extracted Internal Standards denoted with # or outside the 50-150% acceptance range, the respective target analyte results may be unaffected, in other circumstances the PQL has been raised to accommodate the outlier(s).

Updated: 1/7/24 07:32



# CHAIN OF CUSTODY FORM - Client

## ENVIROLAB GROUP



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## Sample information

Envirolab Sample ID	Client Sample ID or Information	Depth	Date sampled	Type of sample	Tests Required												Comments
					Combos 3	PAHs (Trace)	Sulfate	Cations	Anions	Nutrients	Furanics	Dioxins & PCBs	Bacteria	Organic	Inorganic	Trace	
RMW01	-	-	30/05/2024	Water	X	X	X	X	X	X	X	X	X				
RMW02	-	-	30/05/2024	Water	X	X	X	X	X	X	X	X	X				
RMW03	-	-	30/05/2024	Water	X	X	X	X	X	X	X	X	X				
RMW04	-	-	30/05/2024	Water	X	X	X	X	X	X	X	X	X				
RMW05	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
1 MW1-B	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
2 MW2-B	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
MW3-B	-	-	30/05/2024	Water	X	X	X	X	X	X	X	X	X				
MW4-B	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
4 EW1	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
5 DUP1	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
6 TRIP1	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
7 DUP2	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
8 TRIP2	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
9 BLANK	-	-	31/05/2024	Water	X	X	X	X	X	X	X	X	X				
10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
					Sample Count	13	12	6	6	5	6	3					

 Please tick the box if observed settled sediment present in water samples is to be included in the extraction and/or analysis

MD

JMD

Lab Use Only

Print Name:	Kyle Sier	Print Name:	O. L	Job number:	352854-A	Cooling:	Ice / Ice pack / None
Date & Time:	31/05/2024 13:00	Date & Time:	31/5/24 1600	Temperature:	10	Security seal:	Intact / Broken / None
Signature:	K. Sier	Signature:	O.L.	TAT Reg - SAME day / 1 / 2 / 3 / 4 / STD			

Relinquished by (Company): Reditus	Received by (Company): ELS S10
Print Name:	Kyle Sier
Date & Time:	31/05/2024 13:00
Signature:	K. Sier



## SAMPLE RECEIPT NOTIFICATION (SRN)

Work Order : ES2418204

Client	: REDITUS CONSULTING PTY LTD.	Laboratory	: Environmental Division Sydney
Contact	: Toby Scrivener	Contact	: Customer Services ES
Address	: 1A/29-33 Waratah Street KIRRABEE	Address	: 277-289 Woodpark Road Smithfield NSW Australia 2164
E-mail	: tobyscrivener@reditusconsulting.co m.au	E-mail	: ALSEnviro.Sydney@ALSGlobal.com
Telephone	: ----	Telephone	: +61-2-8784 8555
Facsimile	: ----	Facsimile	: +61-2-8784 8500
Project	: 22148	Page	: 1 of 3
Order number	: 22148	Quote number	: EP2023REDITUS0001 (EN/333)
C-O-C number	: ----	QC Level	: NEPM 2013 B3 & ALS QC Standard
Site	:		
Sampler	: KYLE SIER		

### Dates

Date Samples Received	: 03-Jun-2024 17:05	Issue Date	: 03-Jun-2024
Client Requested Due	: 12-Jun-2024	Scheduled Reporting Date	: <b>12-Jun-2024</b>
Date			

### Delivery Details

Mode of Delivery	: Carrier	Security Seal	: Intact.
No. of coolers/boxes	: 1	Temperature	: 10.8, 5.6, 7.6°C - Ice Bricks present
Receipt Detail	:	No. of samples received / analysed	: 2 / 2

### General Comments

- This report contains the following information:
  - Sample Container(s)/Preservation Non-Compliances
  - Summary of Sample(s) and Requested Analysis
  - Proactive Holding Time Report
  - Requested Deliverables
- **Please refer to the Proactive Holding Time Report table below which summarises breaches of recommended holding times that have occurred prior to samples/instructions being received at the laboratory. The laboratory will process these samples unless instructions are received from you indicating you do not wish to proceed. The absence of this summary table indicates that all samples have been received within the recommended holding times for the analysis requested.**
- **Sample(s) requiring volatile organic compound analysis received in airtight containers (ZHE).**
- Please direct any queries you have regarding this work order to the above ALS laboratory contact.
- Analytical work for this work order will be conducted at ALS Sydney.
- Sample Disposal - Aqueous (3 weeks), Solid (2 months ± 1 week) from receipt of samples.
- Please be aware that APHA/NEPM recommends water and soil samples be chilled to less than or equal to 6°C for chemical analysis, and less than or equal to 10°C but unfrozen for Microbiological analysis. Where samples are received above this temperature, it should be taken into consideration when interpreting results. Refer to ALS EnviroMail 85 for ALS recommendations of the best practice for chilling samples after sampling and for maintaining a cool temperature during transit.



## Sample Container(s)/Preservation Non-Compliances

All comparisons are made against pretreatment/preservation AS, APHA, USEPA standards.

- No sample container / preservation non-compliance exists.

## Summary of Sample(s) and Requested Analysis

Some items described below may be part of a laboratory process necessary for the execution of client requested tasks. Packages may contain additional analyses, such as the determination of moisture content and preparation tasks, that are included in the package.

If no sampling time is provided, the sampling time will default 00:00 on the date of sampling. If no sampling date is provided, the sampling date will be assumed by the laboratory and displayed in brackets without a time component

Matrix: WATER

Laboratory sample ID	Sampling date / time	Sample ID	WATER EP231 PFAS - Short Suite (12 analytes)	WATER W-26 TRH/BTEX/NPAH/8 Metals
ES2418204-001	31-May-2024 00:00	TRIP1	✓	✓
ES2418204-002	31-May-2024 00:00	TRIP2	✓	

## Proactive Holding Time Report

Sample(s) have been received within the recommended holding times for the requested analysis.

## Requested Deliverables

### ACCOUNTS

- A4 - AU Tax Invoice (INV) Email accounts@reditusconsulting.com

### Esdat Deliverables

- EDI Format - ESDAT (ESDAT) Email reditusconsulting@esdat.com.au

### KYLE SIER

- \*AU Certificate of Analysis - NATA (COA) Email kylesier@reditusconsulting.com
- \*AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) Email kylesier@reditusconsulting.com
- \*AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) Email kylesier@reditusconsulting.com
- A4 - AU Sample Receipt Notification - Environmental HT (SRN) Email kylesier@reditusconsulting.com
- Chain of Custody (CoC) (COC) Email kylesier@reditusconsulting.com
- EDI Format - ENMRG (ENMRG) Email kylesier@reditusconsulting.com
- EDI Format - ESDAT (ESDAT) Email kylesier@reditusconsulting.com

### REDITUS

- EDI Format - ESDAT (ESDAT) Email reditusconsulting@esdat.com.au

### Toby Scrivener

- \*AU Certificate of Analysis - NATA (COA) Email tobyscrivener@reditusconsulting.co.m.au
- \*AU Interpretive QC Report - DEFAULT (Anon QCI Rep) (QCI) Email tobyscrivener@reditusconsulting.co.m.au
- \*AU QC Report - DEFAULT (Anon QC Rep) - NATA (QC) Email tobyscrivener@reditusconsulting.co.m.au
- A4 - AU Sample Receipt Notification - Environmental HT (SRN) Email tobyscrivener@reditusconsulting.co.m.au
- A4 - AU Tax Invoice (INV) Email tobyscrivener@reditusconsulting.co.m.au
- Chain of Custody (CoC) (COC) Email tobyscrivener@reditusconsulting.co.m.au
- EDI Format - ENMRG (ENMRG) Email tobyscrivener@reditusconsulting.co.m.au
- EDI Format - ESDAT (ESDAT) Email tobyscrivener@reditusconsulting.co.m.au

Issue Date : 03-Jun-2024  
Page : 3 of 3  
Work Order : ES2418204 Amendment 0  
Client : REDITUS CONSULTING PTY LTD.

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## CERTIFICATE OF ANALYSIS

Work Order	: ES2418204	Page	: 1 of 7
Client	: REDITUS CONSULTING PTY LTD.	Laboratory	: Environmental Division Sydney
Contact	: Toby Scrivener	Contact	: Customer Services ES
Address	: 1A/29-33 Waratah Street KIRRAWEE	Address	: 277-289 Woodpark Road Smithfield NSW Australia 2164
Telephone	: ----	Telephone	: +61-2-8784 8555
Project	: 22148	Date Samples Received	: 03-Jun-2024 17:05
Order number	: 22148	Date Analysis Commenced	: 04-Jun-2024
C-O-C number	: ----	Issue Date	: 12-Jun-2024 16:59
Sampler	: KYLE SIER		
Site	:		
Quote number	: EN/333		
No. of samples received	: 2		
No. of samples analysed	: 2		

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Certificate of Analysis contains the following information:

- General Comments
- Analytical Results
- Surrogate Control Limits

Additional information pertinent to this report will be found in the following separate attachments: Quality Control Report, QA/QC Compliance Assessment to assist with Quality Review and Sample Receipt Notification.

### Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories	Position	Accreditation Category
Ankit Joshi	Senior Chemist - Inorganics	Sydney Inorganics, Smithfield, NSW
Edwandy Fadjar	Organic Coordinator	Sydney Organics, Smithfield, NSW
Franco Lentini	LCMS Coordinator	Sydney Organics, Smithfield, NSW



Accreditation No. 825  
Accredited for compliance with  
ISO/IEC 17025 - Testing

## General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis.

Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

When sampling time information is not provided by the client, sampling dates are shown without a time component. In these instances, the time component has been assumed by the laboratory for processing purposes.

Where a result is required to meet compliance limits the associated uncertainty must be considered. Refer to the ALS Contract for details.

Key : CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

^ = This result is computed from individual analyte detections at or above the level of reporting

Ø = ALS is not NATA accredited for these tests.

~ = Indicates an estimated value.

- EP075 (SIM): Where reported, Benzo(a)pyrene Toxicity Equivalent Quotient (TEQ) per the NEPM (2013) is the sum total of the concentration of the eight carcinogenic PAHs multiplied by their Toxicity Equivalence Factor (TEF) relative to Benzo(a)pyrene. TEF values are provided in brackets as follows: Benz(a)anthracene (0.1), Chrysene (0.01), Benzo(b+j) & Benzo(k)fluoranthene (0.1), Benzo(a)pyrene (1.0), Indeno(1.2.3.cd)pyrene (0.1), Dibenz(a,h)anthracene (1.0), Benzo(g.h.i)perylene (0.01). Less than LOR results for 'TEQ Zero' are treated as zero.
- EP231X - Per- and Polyfluoroalkyl Substances (PFAS): Samples received in 20mL or 125mL bottles have been tested in accordance with the QSM5.4 compliant, NATA accredited method. 60mL or 250mL bottles have been tested to the legacy QSM 5.1 aligned, NATA accredited method.
- EP080: Where reported, Total Xylenes is the sum of the reported concentrations of m&p-Xylene and o-Xylene at or above the LOR.
- EP075(SIM): Where reported, Total Cresol is the sum of the reported concentrations of 2-Methylphenol and 3- & 4-Methylphenol at or above the LOR.
- EP231X: PFAS results for sample ES2418204-001 confirmed by re-extraction and re-analysis.
- EP231: Stable isotope enriched internal standards are added to samples prior to extraction. Target compounds have a direct analogous internal standard with the exception of PFPeS, PFHpA, PFDS, PFTDA and 10:2 FTS. These compounds use an internal standard that is chemically related and has a retention time close to that of the target compound. The DQO for internal standard response is 50-150% of that established at initial calibration or as per tables in USEPA 1633 where listed. PFOS is quantified using a certified, traceable standard consisting of linear and branched PFOS isomers. These practices are in line with recommendations in the National Environmental Management Plan for PFAS and also conform to QSM 5.4 (US DoD) requirements.



## Analytical Results

Sub-Matrix: WATER (Matrix: WATER)				Sample ID	TRIP1	TRIP2	---	---	---
Compound	CAS Number	LOR	Unit	Sampling date / time	31-May-2024 00:00	31-May-2024 00:00	---	---	---
				ES2418204-001	ES2418204-002	-----	-----	-----	-----
				Result	Result	---	---	---	---
<b>EG020F: Dissolved Metals by ICP-MS</b>									
Arsenic	7440-38-2	0.001	mg/L	<0.001	---	---	---	---	---
Cadmium	7440-43-9	0.0001	mg/L	<0.0001	---	---	---	---	---
Chromium	7440-47-3	0.001	mg/L	<0.001	---	---	---	---	---
Copper	7440-50-8	0.001	mg/L	<0.001	---	---	---	---	---
Lead	7439-92-1	0.001	mg/L	<0.001	---	---	---	---	---
Nickel	7440-02-0	0.001	mg/L	<b>0.002</b>	---	---	---	---	---
Zinc	7440-66-6	0.005	mg/L	<0.005	---	---	---	---	---
<b>EG035F: Dissolved Mercury by FIMS</b>									
Mercury	7439-97-6	0.0001	mg/L	<0.0001	---	---	---	---	---
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons</b>									
Naphthalene	91-20-3	1.0	µg/L	<1.0	---	---	---	---	---
Acenaphthylene	208-96-8	1.0	µg/L	<1.0	---	---	---	---	---
Acenaphthene	83-32-9	1.0	µg/L	<1.0	---	---	---	---	---
Fluorene	86-73-7	1.0	µg/L	<1.0	---	---	---	---	---
Phenanthrene	85-01-8	1.0	µg/L	<1.0	---	---	---	---	---
Anthracene	120-12-7	1.0	µg/L	<1.0	---	---	---	---	---
Fluoranthene	206-44-0	1.0	µg/L	<1.0	---	---	---	---	---
Pyrene	129-00-0	1.0	µg/L	<1.0	---	---	---	---	---
Benz(a)anthracene	56-55-3	1.0	µg/L	<1.0	---	---	---	---	---
Chrysene	218-01-9	1.0	µg/L	<1.0	---	---	---	---	---
Benzo(b+j)fluoranthene	205-99-2 205-82-3	1.0	µg/L	<1.0	---	---	---	---	---
Benzo(k)fluoranthene	207-08-9	1.0	µg/L	<1.0	---	---	---	---	---
Benzo(a)pyrene	50-32-8	0.5	µg/L	<0.5	---	---	---	---	---
Indeno(1,2,3,cd)pyrene	193-39-5	1.0	µg/L	<1.0	---	---	---	---	---
Dibenz(a,h)anthracene	53-70-3	1.0	µg/L	<1.0	---	---	---	---	---
Benzo(g,h,i)perylene	191-24-2	1.0	µg/L	<1.0	---	---	---	---	---
^ Sum of polycyclic aromatic hydrocarbons	----	0.5	µg/L	<0.5	---	---	---	---	---



## Analytical Results

Sub-Matrix: WATER (Matrix: WATER)		Sample ID	TRIP1	TRIP2	---	---	---	---
		Sampling date / time	31-May-2024 00:00	31-May-2024 00:00	---	---	---	---
Compound	CAS Number	LOR	Unit	ES2418204-001	ES2418204-002	-----	-----	-----
				Result	Result	---	---	---
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons - Continued</b>								
^ Benzo(a)pyrene TEQ (zero)	---	0.5	µg/L	<0.5	---	---	---	---
<b>EP080/071: Total Petroleum Hydrocarbons</b>								
C6 - C9 Fraction	---	20	µg/L	<20	---	---	---	---
C10 - C14 Fraction	---	50	µg/L	<50	---	---	---	---
C15 - C28 Fraction	---	100	µg/L	<100	---	---	---	---
C29 - C36 Fraction	---	50	µg/L	<50	---	---	---	---
^ C10 - C36 Fraction (sum)	---	50	µg/L	<50	---	---	---	---
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions</b>								
C6 - C10 Fraction	C6_C10	20	µg/L	<20	---	---	---	---
^ C6 - C10 Fraction minus BTEX	C6_C10-BTEX (F1)	20	µg/L	<20	---	---	---	---
>C10 - C16 Fraction	---	100	µg/L	<100	---	---	---	---
>C16 - C34 Fraction	---	100	µg/L	<100	---	---	---	---
>C34 - C40 Fraction	---	100	µg/L	<100	---	---	---	---
^ >C10 - C40 Fraction (sum)	---	100	µg/L	<100	---	---	---	---
^ >C10 - C16 Fraction minus Naphthalene (F2)	---	100	µg/L	<100	---	---	---	---
<b>EP080: BTEXN</b>								
Benzene	71-43-2	1	µg/L	<1	---	---	---	---
Toluene	108-88-3	2	µg/L	<2	---	---	---	---
Ethylbenzene	100-41-4	2	µg/L	<2	---	---	---	---
meta- & para-Xylene	108-38-3 106-42-3	2	µg/L	<2	---	---	---	---
ortho-Xylene	95-47-6	2	µg/L	<2	---	---	---	---
^ Total Xylenes	---	2	µg/L	<2	---	---	---	---
^ Sum of BTEX	---	1	µg/L	<1	---	---	---	---
Naphthalene	91-20-3	5	µg/L	<5	---	---	---	---
<b>EP231A: Perfluoroalkyl Sulfonic Acids</b>								
Perfluorobutane sulfonic acid (PFBS)	375-73-5	0.02	µg/L	0.03	<0.02	---	---	---



## Analytical Results

Sub-Matrix: WATER (Matrix: WATER)			Sample ID	TRIP1	TRIP2	---	---	---
			Sampling date / time	31-May-2024 00:00	31-May-2024 00:00	---	---	---
Compound	CAS Number	LOR	Unit	ES2418204-001	ES2418204-002	-----	-----	-----
				Result	Result	---	---	---
<b>EP231A: Perfluoroalkyl Sulfonic Acids - Continued</b>								
Perfluorohexane sulfonic acid (PFHxS)	355-46-4	0.01	µg/L	<0.01	<0.01	---	---	---
Perfluorooctane sulfonic acid (PFOS)	1763-23-1	0.01	µg/L	<0.01	<0.01	---	---	---
<b>EP231B: Perfluoroalkyl Carboxylic Acids</b>								
Perfluorobutanoic acid (PFBA)	375-22-4	0.1	µg/L	<0.1	<0.1	---	---	---
Perfluoropentanoic acid (PFPeA)	2706-90-3	0.02	µg/L	<b>0.04</b>	<0.02	---	---	---
Perfluorohexanoic acid (PFHxA)	307-24-4	0.02	µg/L	<0.02	<0.02	---	---	---
Perfluoroheptanoic acid (PFHpA)	375-85-9	0.02	µg/L	<0.02	<0.02	---	---	---
Perfluorooctanoic acid (PFOA)	335-67-1	0.01	µg/L	<0.01	<0.01	---	---	---
<b>EP231D: (n:2) Fluorotelomer Sulfonic Acids</b>								
4:2 Fluorotelomer sulfonic acid (4:2 FTS)	757124-72-4	0.05	µg/L	<0.05	<0.05	---	---	---
6:2 Fluorotelomer sulfonic acid (6:2 FTS)	27619-97-2	0.05	µg/L	<0.05	<0.05	---	---	---
8:2 Fluorotelomer sulfonic acid (8:2 FTS)	39108-34-4	0.05	µg/L	<0.05	<0.05	---	---	---
10:2 Fluorotelomer sulfonic acid (10:2 FTS)	120226-60-0	0.05	µg/L	<0.05	<0.05	---	---	---
<b>EP231P: PFAS Sums</b>								
Sum of PFHxS and PFOS	355-46-4/1763-23-1	0.01	µg/L	<0.01	<0.01	---	---	---
Sum of PFAS (WA DER List)	---	0.01	µg/L	<b>0.07</b>	<0.01	---	---	---
<b>EP075(SIM)S: Phenolic Compound Surrogates</b>								
Phenol-d6	13127-88-3	1.0	%	<b>26.1</b>	---	---	---	---
2-Chlorophenol-D4	93951-73-6	1.0	%	<b>47.1</b>	---	---	---	---
2,4,6-Tribromophenol	118-79-6	1.0	%	<b>41.4</b>	---	---	---	---
<b>EP075(SIM)T: PAH Surrogates</b>								
2-Fluorobiphenyl	321-60-8	1.0	%	<b>59.3</b>	---	---	---	---
Anthracene-d10	1719-06-8	1.0	%	<b>58.8</b>	---	---	---	---
4-Terphenyl-d14	1718-51-0	1.0	%	<b>60.9</b>	---	---	---	---



## Analytical Results

Sub-Matrix: WATER (Matrix: WATER)				Sample ID	TRIP1	TRIP2	---	---	---
				Sampling date / time	31-May-2024 00:00	31-May-2024 00:00	---	---	---
Compound	CAS Number	LOR	Unit	ES2418204-001	ES2418204-002	-----	-----	-----	
				Result	Result	---	---	---	
<b>EP080S: TPH(V)/BTEX Surrogates</b>									
1,2-Dichloroethane-D4	17060-07-0	2	%	101	---	---	---	---	---
Toluene-D8	2037-26-5	2	%	97.1	---	---	---	---	---
4-Bromofluorobenzene	460-00-4	2	%	93.3	---	---	---	---	---
<b>EP231S: PFAS Surrogate</b>									
13C4-PFOS	---	0.02	%	105	106	---	---	---	---
13C8-PFOA	---	0.02	%	101	104	---	---	---	---



## Surrogate Control Limits

Sub-Matrix: WATER		Recovery Limits (%)	
Compound	CAS Number	Low	High
<b>EP075(SIM)S: Phenolic Compound Surrogates</b>			
Phenol-d6	13127-88-3	10	44
2-Chlorophenol-D4	93951-73-6	14	94
2,4,6-Tribromophenol	118-79-6	17	125
<b>EP075(SIM)T: PAH Surrogates</b>			
2-Fluorobiphenyl	321-60-8	20	104
Anthracene-d10	1719-06-8	27	113
4-Terphenyl-d14	1718-51-0	32	112
<b>EP080S: TPH(V)/BTEX Surrogates</b>			
1,2-Dichloroethane-D4	17060-07-0	72	143
Toluene-D8	2037-26-5	75	131
4-Bromofluorobenzene	460-00-4	73	137
<b>EP231S: PFAS Surrogate</b>			
13C4-PFOS	----	60	120
13C8-PFOA	----	60	120



## QA/QC Compliance Assessment to assist with Quality Review

Work Order	: ES2418204	Page	: 1 of 6
Client	: REDITUS CONSULTING PTY LTD.	Laboratory	: Environmental Division Sydney
Contact	: Toby Scrivener	Telephone	: +61-2-8784 8555
Project	: 22148	Date Samples Received	: 03-Jun-2024
Site	:	Issue Date	: 12-Jun-2024
Sampler	: KYLE SIER	No. of samples received	: 2
Order number	: 22148	No. of samples analysed	: 2

This report is automatically generated by the ALS LIMS through interpretation of the ALS Quality Control Report and several Quality Assurance parameters measured by ALS. This automated reporting highlights any non-conformances, facilitates faster and more accurate data validation and is designed to assist internal expert and external Auditor review. Many components of this report contribute to the overall DQO assessment and reporting for guideline compliance.

Brief method summaries and references are also provided to assist in traceability.

### ***Summary of Outliers***

#### ***Outliers : Quality Control Samples***

This report highlights outliers flagged in the Quality Control (QC) Report.

- NO Method Blank value outliers occur.
- NO Duplicate outliers occur.
- NO Laboratory Control outliers occur.
- NO Matrix Spike outliers occur.
- For all regular sample matrices, where applicable to the methodology, NO surrogate recovery outliers occur.

#### ***Outliers : Analysis Holding Time Compliance***

- NO Analysis Holding Time Outliers exist.

#### ***Outliers : Frequency of Quality Control Samples***

- Quality Control Sample Frequency Outliers exist - please see following pages for full details.



### Outliers : Frequency of Quality Control Samples

Matrix: WATER

Quality Control Sample Type	Method	Count		Rate (%)		Quality Control Specification
		QC	Regular	Actual	Expected	
<b>Laboratory Duplicates (DUP)</b>						
PAH/Phenols (GC/MS - SIM)	EP075(SIM)	0	11	0.00	10.00	NEPM 2013 B3 & ALS QC Standard
Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS	EP231X	0	17	0.00	10.00	NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction	EP071	0	11	0.00	10.00	NEPM 2013 B3 & ALS QC Standard
<b>Matrix Spikes (MS)</b>						
PAH/Phenols (GC/MS - SIM)	EP075(SIM)	0	11	0.00	5.00	NEPM 2013 B3 & ALS QC Standard
Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS	EP231X	0	17	0.00	5.00	NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction	EP071	0	11	0.00	5.00	NEPM 2013 B3 & ALS QC Standard

### Analysis Holding Time Compliance

If samples are identified below as having been analysed or extracted outside of recommended holding times, this should be taken into consideration when interpreting results.

This report summarizes extraction / preparation and analysis times and compares each with ALS recommended holding times (referencing USEPA SW 846, APHA, AS and NEPM) based on the sample container provided. Dates reported represent first date of extraction or analysis and preclude subsequent dilutions and reruns. A listing of breaches (if any) is provided herein.

Holding time for leachate methods (e.g. TCLP) vary according to the analytes reported. Assessment compares the leach date with the shortest analyte holding time for the equivalent soil method. These are: organics 14 days, mercury 28 days & other metals 180 days. A recorded breach does not guarantee a breach for all non-volatile parameters.

Holding times for VOC in soils vary according to analytes of interest. Vinyl Chloride and Styrene holding time is 7 days; others 14 days. A recorded breach does not guarantee a breach for all VOC analytes and should be verified in case the reported breach is a false positive or Vinyl Chloride and Styrene are not key analytes of interest/concern.

Matrix: WATER

Evaluation: ✗ = Holding time breach ; ✓ = Within holding time.

Method	Container / Client Sample ID(s)	Sample Date	Extraction / Preparation			Analysis		
			Date extracted	Due for extraction	Evaluation	Date analysed	Due for analysis	Evaluation
<b>EG020F: Dissolved Metals by ICP-MS</b>								
Clear Plastic Bottle - Nitric Acid; Filtered (EG020A-F) TRIP1		31-May-2024	---	---	---	07-Jun-2024	27-Nov-2024	✓
<b>EG035F: Dissolved Mercury by FIMS</b>								
Clear Plastic Bottle - Nitric Acid; Filtered (EG035F) TRIP1		31-May-2024	---	---	---	11-Jun-2024	28-Jun-2024	✓
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons</b>								
Amber Glass Bottle - Unpreserved (EP075(SIM)) TRIP1		31-May-2024	04-Jun-2024	07-Jun-2024	✓	06-Jun-2024	14-Jul-2024	✓
<b>EP080/071: Total Petroleum Hydrocarbons</b>								
Amber Glass Bottle - Unpreserved (EP071) TRIP1		31-May-2024	04-Jun-2024	07-Jun-2024	✓	07-Jun-2024	14-Jul-2024	✓
Clear glass VOC vial - HCl (EP080) TRIP1		31-May-2024	04-Jun-2024	14-Jun-2024	✓	04-Jun-2024	14-Jun-2024	✓
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions</b>								
Amber Glass Bottle - Unpreserved (EP071) TRIP1		31-May-2024	04-Jun-2024	07-Jun-2024	✓	07-Jun-2024	14-Jul-2024	✓
Clear glass VOC vial - HCl (EP080) TRIP1		31-May-2024	04-Jun-2024	14-Jun-2024	✓	04-Jun-2024	14-Jun-2024	✓



Matrix: WATER									Evaluation: ✗ = Holding time breach ; ✓ = Within holding time.					
Method	Container / Client Sample ID(s)	Sample Date	Extraction / Preparation			Analysis								
			Date extracted	Due for extraction	Evaluation	Date analysed	Due for analysis	Evaluation						
<b>EP080: BTEXN</b>														
Clear glass VOC vial - HCl (EP080)	TRIP1	31-May-2024	04-Jun-2024	14-Jun-2024	✓	04-Jun-2024	14-Jun-2024	✓						
<b>EP231A: Perfluoroalkyl Sulfonic Acids</b>														
HDPE (no PTFE) (EP231X)	TRIP1, TRIP2	31-May-2024	05-Jun-2024	27-Nov-2024	✓	11-Jun-2024	27-Nov-2024	✓						
<b>EP231B: Perfluoroalkyl Carboxylic Acids</b>														
HDPE (no PTFE) (EP231X)	TRIP1, TRIP2	31-May-2024	05-Jun-2024	27-Nov-2024	✓	11-Jun-2024	27-Nov-2024	✓						
<b>EP231D: (n:2) Fluorotelomer Sulfonic Acids</b>														
HDPE (no PTFE) (EP231X)	TRIP1, TRIP2	31-May-2024	05-Jun-2024	27-Nov-2024	✓	11-Jun-2024	27-Nov-2024	✓						
<b>EP231P: PFAS Sums</b>														
HDPE (no PTFE) (EP231X)	TRIP1, TRIP2	31-May-2024	05-Jun-2024	27-Nov-2024	✓	11-Jun-2024	27-Nov-2024	✓						

## Quality Control Parameter Frequency Compliance

The following report summarises the frequency of laboratory QC samples analysed within the analytical lot(s) in which the submitted sample(s) was(were) processed. Actual rate should be greater than or equal to the expected rate. A listing of breaches is provided in the Summary of Outliers.

Matrix: WATER

Evaluation: ✗ = Quality Control frequency not within specification ; ✓ = Quality Control frequency within specification.

Quality Control Sample Type	Analytical Methods	Method	Count		Rate (%)		Quality Control Specification
			QC	Regular	Actual	Expected	
<b>Laboratory Duplicates (DUP)</b>							
Dissolved Mercury by FIMS		EG035F	2	19	10.53	10.00	✓ NEPM 2013 B3 & ALS QC Standard
Dissolved Metals by ICP-MS - Suite A		EG020A-F	2	18	11.11	10.00	✓ NEPM 2013 B3 & ALS QC Standard
PAH/Phenols (GC/MS - SIM)		EP075(SIM)	0	11	0.00	10.00	✗ NEPM 2013 B3 & ALS QC Standard
Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS		EP231X	0	17	0.00	10.00	✗ NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction		EP071	0	11	0.00	10.00	✗ NEPM 2013 B3 & ALS QC Standard
TRH Volatiles/BTEX		EP080	2	13	15.38	10.00	✓ NEPM 2013 B3 & ALS QC Standard
<b>Laboratory Control Samples (LCS)</b>							
Dissolved Mercury by FIMS		EG035F	1	19	5.26	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Dissolved Metals by ICP-MS - Suite A		EG020A-F	1	18	5.56	5.00	✓ NEPM 2013 B3 & ALS QC Standard
PAH/Phenols (GC/MS - SIM)		EP075(SIM)	1	11	9.09	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS		EP231X	1	17	5.88	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction		EP071	1	11	9.09	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH Volatiles/BTEX		EP080	1	13	7.69	5.00	✓ NEPM 2013 B3 & ALS QC Standard
<b>Method Blanks (MB)</b>							
Dissolved Mercury by FIMS		EG035F	1	19	5.26	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Dissolved Metals by ICP-MS - Suite A		EG020A-F	1	18	5.56	5.00	✓ NEPM 2013 B3 & ALS QC Standard
PAH/Phenols (GC/MS - SIM)		EP075(SIM)	1	11	9.09	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS		EP231X	1	17	5.88	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction		EP071	1	11	9.09	5.00	✓ NEPM 2013 B3 & ALS QC Standard
TRH Volatiles/BTEX		EP080	1	13	7.69	5.00	✓ NEPM 2013 B3 & ALS QC Standard
<b>Matrix Spikes (MS)</b>							
Dissolved Mercury by FIMS		EG035F	1	19	5.26	5.00	✓ NEPM 2013 B3 & ALS QC Standard
Dissolved Metals by ICP-MS - Suite A		EG020A-F	1	18	5.56	5.00	✓ NEPM 2013 B3 & ALS QC Standard
PAH/Phenols (GC/MS - SIM)		EP075(SIM)	0	11	0.00	5.00	✗ NEPM 2013 B3 & ALS QC Standard
Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS		EP231X	0	17	0.00	5.00	✗ NEPM 2013 B3 & ALS QC Standard
TRH - Semivolatile Fraction		EP071	0	11	0.00	5.00	✗ NEPM 2013 B3 & ALS QC Standard
TRH Volatiles/BTEX		EP080	1	13	7.69	5.00	✓ NEPM 2013 B3 & ALS QC Standard



## Brief Method Summaries

The analytical procedures used by the Environmental Division have been developed from established internationally recognized procedures such as those published by the US EPA, APHA, AS and NEPM. In house developed procedures are employed in the absence of documented standards or by client request. The following report provides brief descriptions of the analytical procedures employed for results reported in the Certificate of Analysis. Sources from which ALS methods have been developed are provided within the Method Descriptions.

Analytical Methods	Method	Matrix	Method Descriptions
Dissolved Metals by ICP-MS - Suite A	EG020A-F	WATER	In house: Referenced to APHA 3125; USEPA SW846 - 6020, ALS QWI-EN/EG020. Samples are 0.45µm filtered prior to analysis. The ICPMS technique utilizes a highly efficient argon plasma to ionize selected elements. Ions are then passed into a high vacuum mass spectrometer, which separates the analytes based on their distinct mass to charge ratios prior to their measurement by a discrete dynode ion detector.
Dissolved Mercury by FIMS	EG035F	WATER	In house: Referenced to APHA 3112 Hg - B (Flow-injection (SnCl <sub>2</sub> )(Cold Vapour generation) AAS) Samples are 0.45µm filtered prior to analysis. FIM-AAS is an automated flameless atomic absorption technique. A bromate/bromide reagent is used to oxidise any organic mercury compounds in the filtered sample. The ionic mercury is reduced online to atomic mercury vapour by SnCl <sub>2</sub> which is then purged into a heated quartz cell. Quantification is by comparing absorbance against a calibration curve. This method is compliant with NEPM Schedule B(3).
TRH - Semivolatile Fraction	EP071	WATER	In house: Referenced to USEPA SW 846 - 8015 The sample extract is analysed by Capillary GC/FID and quantification is by comparison against an established 5 point calibration curve of n-Alkane standards. This method is compliant with the QC requirements of NEPM Schedule B(3)
PAH/Phenols (GC/MS - SIM)	EP075(SIM)	WATER	In house: Referenced to USEPA SW 846 - 8270 Sample extracts are analysed by Capillary GC/MS in SIM Mode and quantification is by comparison against an established 5 point calibration curve. This method is compliant with NEPM Schedule B(3)
TRH Volatiles/BTEX	EP080	WATER	In house: Referenced to USEPA SW 846 - 8260 Water samples are directly purged prior to analysis by Capillary GC/MS and quantification is by comparison against an established 5 point calibration curve. Alternatively, a sample is equilibrated in a headspace vial and a portion of the headspace determined by GCMS analysis. This method is compliant with the QC requirements of NEPM Schedule B(3)
Per- and Polyfluoroalkyl Substances (PFAS) by LCMSMS	EP231X	WATER	In-house: Analysis of fresh and saline waters by Solid Phase Extraction (SPE) followed by LC-Electrospray-MS-MS, Negative Mode using MRM and internal standard quantitation. Isotopically labelled analogues of target analytes used as internal standards and surrogates are added to the sample container. The entire contents are transferred to a solid phase extraction (SPE) cartridge. The sample container is successively rinsed with aliquots of the elution solvent. The eluted extract is combined with an equal volume of reagent water and a portion is filtered for analysis. Method procedures and data quality objectives conform to US DoD QSM 5.4, table B-15 requirements.

Preparation Methods	Method	Matrix	Method Descriptions
Separatory Funnel Extraction of Liquids	ORG14	WATER	In house: Referenced to USEPA SW 846 - 3510 100 mL to 1L of sample is transferred to a separatory funnel and serially extracted three times using DCM for each extract. The resultant extracts are combined, dehydrated and concentrated for analysis. This method is compliant with NEPM Schedule B(3) . ALS default excludes sediment which may be resident in the container.
Volatiles Water Preparation	ORG16-W	WATER	A 5 mL aliquot or 5 mL of a diluted sample is added to a 40 mL VOC vial for purging.



<i>Preparation Methods</i>	<i>Method</i>	<i>Matrix</i>	<i>Method Descriptions</i>
Solid Phase Extraction (SPE) for PFAS in water	ORG72	WATER	In-house: Isotopically labelled analogues of target analytes used as internal standards and surrogates are added to the sample container. The entire contents are transferred to a solid phase extraction (SPE) cartridge. The sample container is successively rinsed with aliquots of the elution solvent. The eluted extract is combined with an equal volume of reagent water and a portion is filtered for analysis. Method procedures conform to US DoD QSM 5.3, table B-15 requirements.



## QUALITY CONTROL REPORT

Work Order	: ES2418204	Page	: 1 of 6
Client	: REDITUS CONSULTING PTY LTD.	Laboratory	: Environmental Division Sydney
Contact	: Toby Scrivener	Contact	: Customer Services ES
Address	: 1A/29-33 Waratah Street KIRRRAWEE	Address	: 277-289 Woodpark Road Smithfield NSW Australia 2164
Telephone	: ----	Telephone	: +61-2-8784 8555
Project	: 22148	Date Samples Received	: 03-Jun-2024
Order number	: 22148	Date Analysis Commenced	: 04-Jun-2024
C-O-C number	: ----	Issue Date	: 12-Jun-2024
Sampler	: KYLE SIER		
Site	:		
Quote number	: EN/333		
No. of samples received	: 2		
No. of samples analysed	: 2		



Accreditation No. 825  
Accredited for compliance with  
ISO/IEC 17025 - Testing

This report supersedes any previous report(s) with this reference. Results apply to the sample(s) as submitted, unless the sampling was conducted by ALS. This document shall not be reproduced, except in full.

This Quality Control Report contains the following information:

- Laboratory Duplicate (DUP) Report; Relative Percentage Difference (RPD) and Acceptance Limits
- Method Blank (MB) and Laboratory Control Spike (LCS) Report; Recovery and Acceptance Limits
- Matrix Spike (MS) Report; Recovery and Acceptance Limits

### Signatories

This document has been electronically signed by the authorized signatories below. Electronic signing is carried out in compliance with procedures specified in 21 CFR Part 11.

Signatories	Position	Accreditation Category
Ankit Joshi	Senior Chemist - Inorganics	Sydney Inorganics, Smithfield, NSW
Edwandy Fadjar	Organic Coordinator	Sydney Organics, Smithfield, NSW
Franco Lentini	LCMS Coordinator	Sydney Organics, Smithfield, NSW



## General Comments

The analytical procedures used by ALS have been developed from established internationally recognised procedures such as those published by the USEPA, APHA, AS and NEPM. In house developed procedures are fully validated and are often at the client request.

Where moisture determination has been performed, results are reported on a dry weight basis.

Where a reported less than (<) result is higher than the LOR, this may be due to primary sample extract/digestate dilution and/or insufficient sample for analysis. Where the LOR of a reported result differs from standard LOR, this may be due to high moisture content, insufficient sample (reduced weight employed) or matrix interference.

Key : Anonymous = Refers to samples which are not specifically part of this work order but formed part of the QC process lot

CAS Number = CAS registry number from database maintained by Chemical Abstracts Services. The Chemical Abstracts Service is a division of the American Chemical Society.

LOR = Limit of reporting

RPD = Relative Percentage Difference

# = Indicates failed QC

## Laboratory Duplicate (DUP) Report

The quality control term Laboratory Duplicate refers to a randomly selected intralaboratory split. Laboratory duplicates provide information regarding method precision and sample heterogeneity. The permitted ranges for the Relative Percent Deviation (RPD) of Laboratory Duplicates are specified in ALS Method QWI-EN/38 and are dependent on the magnitude of results in comparison to the level of reporting: Result < 10 times LOR: No Limit; Result between 10 and 20 times LOR: 0% - 50%; Result > 20 times LOR: 0% - 20%.

Sub-Matrix: WATER

Laboratory sample ID		Sample ID	Method: Compound	CAS Number	LOR	Unit	Original Result	Duplicate Result	RPD (%)	Acceptable RPD (%)
<b>EG020F: Dissolved Metals by ICP-MS (QC Lot: 5842140)</b>										
ES2418202-001	Anonymous	EG020A-F: Cadmium	7440-43-9	0.0001	mg/L	<0.0001	<0.0001	0.0	No Limit	
		EG020A-F: Arsenic	7440-38-2	0.001	mg/L	<0.001	<0.001	0.0	No Limit	
		EG020A-F: Chromium	7440-47-3	0.001	mg/L	<0.001	<0.001	0.0	No Limit	
		EG020A-F: Copper	7440-50-8	0.001	mg/L	<0.001	<0.001	0.0	No Limit	
		EG020A-F: Lead	7439-92-1	0.001	mg/L	<0.001	<0.001	0.0	No Limit	
		EG020A-F: Nickel	7440-02-0	0.001	mg/L	0.014	0.010	32.2	0% - 50%	
		EG020A-F: Zinc	7440-66-6	0.005	mg/L	0.007	<0.005	36.9	No Limit	
ES2418260-002	Anonymous	EG020A-F: Cadmium	7440-43-9	0.0001	mg/L	<0.0001	<0.0001	0.0	No Limit	
		EG020A-F: Arsenic	7440-38-2	0.001	mg/L	<0.001	<0.001	0.0	No Limit	
		EG020A-F: Chromium	7440-47-3	0.001	mg/L	<0.001	<0.001	0.0	No Limit	
		EG020A-F: Copper	7440-50-8	0.001	mg/L	<0.001	<0.001	0.0	No Limit	
		EG020A-F: Lead	7439-92-1	0.001	mg/L	<0.001	<0.001	0.0	No Limit	
		EG020A-F: Nickel	7440-02-0	0.001	mg/L	<0.001	<0.001	0.0	No Limit	
		EG020A-F: Zinc	7440-66-6	0.005	mg/L	<0.005	<0.005	0.0	No Limit	
<b>EG035F: Dissolved Mercury by FIMS (QC Lot: 5842139)</b>										
ES2418246-001	Anonymous	EG035F: Mercury	7439-97-6	0.0001	mg/L	<0.0001	<0.0001	0.0	No Limit	
ES2418288-003	Anonymous	EG035F: Mercury	7439-97-6	0.0001	mg/L	<0.0001	<0.0001	0.0	No Limit	
<b>EP080/071: Total Petroleum Hydrocarbons (QC Lot: 5833350)</b>										
WN2406736-002	Anonymous	EP080: C6 - C9 Fraction	---	20	µg/L	<20	<20	0.0	No Limit	
ES2418166-001	Anonymous	EP080: C6 - C9 Fraction	---	20	µg/L	<20	<20	0.0	No Limit	
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5833350)</b>										
WN2406736-002	Anonymous	EP080: C6 - C10 Fraction	C6_C10	20	µg/L	<20	<20	0.0	No Limit	



**Sub-Matrix: WATER**

Laboratory Duplicate (DUP) Report									
Laboratory sample ID	Sample ID	Method: Compound	CAS Number	LOR	Unit	Original Result	Duplicate Result	RPD (%)	Acceptable RPD (%)
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5833350) - continued</b>									
ES2418166-001	Anonymous	EP080: C6 - C10 Fraction	C6_C10	20	µg/L	<20	<20	0.0	No Limit
<b>EP080: BTEXN (QC Lot: 5833350)</b>									
WN2406736-002	Anonymous	EP080: Benzene	71-43-2	1	µg/L	<1	<1	0.0	No Limit
		EP080: Toluene	108-88-3	2	µg/L	<2	<2	0.0	No Limit
		EP080: Ethylbenzene	100-41-4	2	µg/L	<2	<2	0.0	No Limit
		EP080: meta- & para-Xylene	108-38-3 106-42-3	2	µg/L	<2	<2	0.0	No Limit
		EP080: ortho-Xylene	95-47-6	2	µg/L	<2	<2	0.0	No Limit
		EP080: Naphthalene	91-20-3	5	µg/L	<5	<5	0.0	No Limit
ES2418166-001	Anonymous	EP080: Benzene	71-43-2	1	µg/L	<1	<1	0.0	No Limit
		EP080: Toluene	108-88-3	2	µg/L	<2	<2	0.0	No Limit
		EP080: Ethylbenzene	100-41-4	2	µg/L	<2	<2	0.0	No Limit
		EP080: meta- & para-Xylene	108-38-3 106-42-3	2	µg/L	<2	<2	0.0	No Limit
		EP080: ortho-Xylene	95-47-6	2	µg/L	<2	<2	0.0	No Limit
		EP080: Naphthalene	91-20-3	5	µg/L	<5	<5	0.0	No Limit



## Method Blank (MB) and Laboratory Control Sample (LCS) Report

The quality control term Method / Laboratory Blank refers to an analyte free matrix to which all reagents are added in the same volumes or proportions as used in standard sample preparation. The purpose of this QC parameter is to monitor potential laboratory contamination. The quality control term Laboratory Control Sample (LCS) refers to a certified reference material, or a known interference free matrix spiked with target analytes. The purpose of this QC parameter is to monitor method precision and accuracy independent of sample matrix. Dynamic Recovery Limits are based on statistical evaluation of processed LCS.

Sub-Matrix: WATER

Method: Compound	CAS Number	LOR	Unit	Result	Method Blank (MB) Report	Laboratory Control Spike (LCS) Report		
					Spike Concentration	Spike Recovery (%)	Acceptable Limits (%)	
							Low	High
<b>EG020F: Dissolved Metals by ICP-MS (QC Lot: 5842140)</b>								
EG020A-F: Arsenic	7440-38-2	0.001	mg/L	<0.001	0.1 mg/L	88.7	85.0	114
EG020A-F: Cadmium	7440-43-9	0.0001	mg/L	<0.0001	0.1 mg/L	91.6	84.0	110
EG020A-F: Chromium	7440-47-3	0.001	mg/L	<0.001	0.1 mg/L	87.3	85.0	111
EG020A-F: Copper	7440-50-8	0.001	mg/L	<0.001	0.1 mg/L	89.2	81.0	111
EG020A-F: Lead	7439-92-1	0.001	mg/L	<0.001	0.1 mg/L	88.2	83.0	111
EG020A-F: Nickel	7440-02-0	0.001	mg/L	<0.001	0.1 mg/L	85.7	82.0	112
EG020A-F: Zinc	7440-66-6	0.005	mg/L	<0.005	0.1 mg/L	86.5	81.0	117
<b>EG035F: Dissolved Mercury by FIMS (QC Lot: 5842139)</b>								
EG035F: Mercury	7439-97-6	0.0001	mg/L	<0.0001	0.01 mg/L	96.7	83.0	105
<b>EP075(SIM)B: Polynuclear Aromatic Hydrocarbons (QC Lot: 5833287)</b>								
EP075(SIM): Naphthalene	91-20-3	1	µg/L	<1.0	5 µg/L	68.9	50.0	94.0
EP075(SIM): Acenaphthylene	208-96-8	1	µg/L	<1.0	5 µg/L	75.4	63.6	114
EP075(SIM): Acenaphthene	83-32-9	1	µg/L	<1.0	5 µg/L	74.3	62.2	113
EP075(SIM): Fluorene	86-73-7	1	µg/L	<1.0	5 µg/L	75.6	63.9	115
EP075(SIM): Phenanthrene	85-01-8	1	µg/L	<1.0	5 µg/L	72.2	62.6	116
EP075(SIM): Anthracene	120-12-7	1	µg/L	<1.0	5 µg/L	73.2	64.3	116
EP075(SIM): Fluoranthene	206-44-0	1	µg/L	<1.0	5 µg/L	68.1	63.6	118
EP075(SIM): Pyrene	129-00-0	1	µg/L	<1.0	5 µg/L	71.3	63.1	118
EP075(SIM): Benz(a)anthracene	56-55-3	1	µg/L	<1.0	5 µg/L	72.8	64.1	117
EP075(SIM): Chrysene	218-01-9	1	µg/L	<1.0	5 µg/L	77.3	62.5	116
EP075(SIM): Benzo(b+j)fluoranthene	205-99-2	1	µg/L	<1.0	5 µg/L	67.4	61.7	119
	205-82-3							
EP075(SIM): Benzo(k)fluoranthene	207-08-9	1	µg/L	<1.0	5 µg/L	80.1	63.0	115
EP075(SIM): Benzo(a)pyrene	50-32-8	0.5	µg/L	<0.5	5 µg/L	72.7	63.3	117
EP075(SIM): Indeno(1,2,3,cd)pyrene	193-39-5	1	µg/L	<1.0	5 µg/L	71.1	59.9	118
EP075(SIM): Dibenz(a,h)anthracene	53-70-3	1	µg/L	<1.0	5 µg/L	73.4	61.2	117
EP075(SIM): Benzo(g,h,i)perylene	191-24-2	1	µg/L	<1.0	5 µg/L	71.0	59.1	118
<b>EP080/071: Total Petroleum Hydrocarbons (QC Lot: 5833285)</b>								
EP071: C10 - C14 Fraction	----	50	µg/L	<50	400 µg/L	63.6	53.7	97.0
EP071: C15 - C28 Fraction	----	100	µg/L	<100	600 µg/L	107	63.3	107
EP071: C29 - C36 Fraction	----	50	µg/L	<50	400 µg/L	103	58.3	120



Sub-Matrix: WATER				Method Blank (MB) Report	Laboratory Control Spike (LCS) Report				
					Spike Concentration	Spike Recovery (%)	Acceptable Limits (%)		
Method: Compound	CAS Number	LOR	Unit		Result		LCS	Low	High
<b>EP080/071: Total Petroleum Hydrocarbons (QC Lot: 5833350)</b>									
EP080: C6 - C9 Fraction	---	20	µg/L	<20	260 µg/L	93.3	75.0	127	
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5833285)</b>									
EP071: >C10 - C16 Fraction	---	100	µg/L	<100	500 µg/L	69.3	53.9	95.5	
EP071: >C16 - C34 Fraction	---	100	µg/L	<100	700 µg/L	105	57.8	110	
EP071: >C34 - C40 Fraction	---	100	µg/L	<100	300 µg/L	101	50.5	115	
<b>EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QC Lot: 5833350)</b>									
EP080: C6 - C10 Fraction	C6_C10	20	µg/L	<20	310 µg/L	90.9	75.0	127	
<b>EP080: BTEXN (QC Lot: 5833350)</b>									
EP080: Benzene	71-43-2	1	µg/L	<1	10 µg/L	94.4	68.3	119	
EP080: Toluene	108-88-3	2	µg/L	<2	10 µg/L	93.1	73.5	120	
EP080: Ethylbenzene	100-41-4	2	µg/L	<2	10 µg/L	91.1	73.8	122	
EP080: meta- & para-Xylene	108-38-3 106-42-3	2	µg/L	<2	10 µg/L	89.5	73.0	122	
EP080: ortho-Xylene	95-47-6	2	µg/L	<2	10 µg/L	87.9	76.4	123	
EP080: Naphthalene	91-20-3	5	µg/L	<5	10 µg/L	108	75.5	124	
<b>EP231A: Perfluoroalkyl Sulfonic Acids (QC Lot: 5836674)</b>									
EP231X: Perfluorobutane sulfonic acid (PFBS)	375-73-5	0.02	µg/L	<0.02	0.25 µg/L	92.2	72.0	130	
EP231X: Perfluorohexane sulfonic acid (PFHxS)	355-46-4	0.01	µg/L	<0.01	0.25 µg/L	91.9	68.0	131	
EP231X: Perfluorooctane sulfonic acid (PFOS)	1763-23-1	0.01	µg/L	<0.01	0.25 µg/L	99.6	65.0	140	
<b>EP231B: Perfluoroalkyl Carboxylic Acids (QC Lot: 5836674)</b>									
EP231X: Perfluorobutanoic acid (PFBA)	375-22-4	0.1	µg/L	<0.1	1.25 µg/L	102	73.0	129	
EP231X: Perfluoropentanoic acid (PFPeA)	2706-90-3	0.02	µg/L	<0.02	0.25 µg/L	101	72.0	129	
EP231X: Perfluorohexanoic acid (PFHxA)	307-24-4	0.02	µg/L	<0.02	0.25 µg/L	102	72.0	129	
EP231X: Perfluoroheptanoic acid (PFHpA)	375-85-9	0.02	µg/L	<0.02	0.25 µg/L	102	72.0	130	
EP231X: Perfluorooctanoic acid (PFOA)	335-67-1	0.01	µg/L	<0.01	0.25 µg/L	90.2	71.0	133	
<b>EP231D: (n:2) Fluorotelomer Sulfonic Acids (QC Lot: 5836674)</b>									
EP231X: 4:2 Fluorotelomer sulfonic acid (4:2 FTS)	757124-72-4	0.05	µg/L	<0.05	0.25 µg/L	85.9	63.0	143	
EP231X: 6:2 Fluorotelomer sulfonic acid (6:2 FTS)	27619-97-2	0.05	µg/L	<0.05	0.25 µg/L	86.5	64.0	140	
EP231X: 8:2 Fluorotelomer sulfonic acid (8:2 FTS)	39108-34-4	0.05	µg/L	<0.05	0.25 µg/L	82.7	67.0	138	
EP231X: 10:2 Fluorotelomer sulfonic acid (10:2 FTS)	120226-60-0	0.05	µg/L	<0.05	0.25 µg/L	87.4	71.4	144	

### Matrix Spike (MS) Report

The quality control term Matrix Spike (MS) refers to an intralaboratory split sample spiked with a representative set of target analytes. The purpose of this QC parameter is to monitor potential matrix effects on analyte recoveries. Static Recovery Limits as per laboratory Data Quality Objectives (DQOs). Ideal recovery ranges stated may be waived in the event of sample matrix interference.



Sub-Matrix: WATER

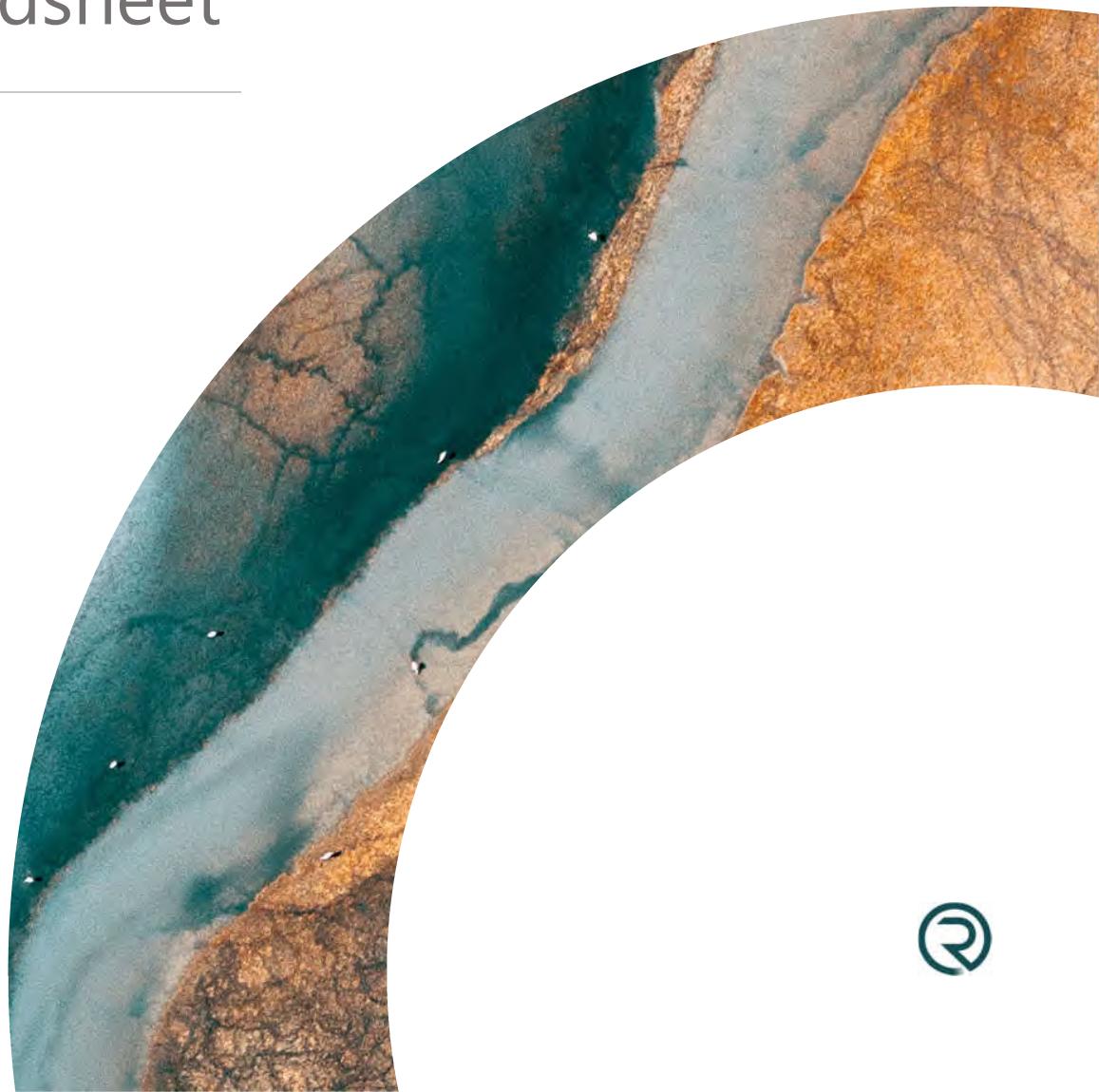
				Matrix Spike (MS) Report								
Laboratory sample ID	Sample ID	Method: Compound	CAS Number	Spike	Spike Recovery(%)	Acceptable Limits (%)						
EG020F: Dissolved Metals by ICP-MS (QCLot: 5842140)				Concentration	MS	Low	High					
ES2418204-001	TRIP1	EG020A-F: Arsenic	7440-38-2	1 mg/L	83.2	70.0	130					
		EG020A-F: Cadmium	7440-43-9	0.25 mg/L	88.6	70.0	130					
		EG020A-F: Chromium	7440-47-3	1 mg/L	83.7	70.0	130					
		EG020A-F: Copper	7440-50-8	1 mg/L	87.2	70.0	130					
		EG020A-F: Lead	7439-92-1	1 mg/L	76.9	70.0	130					
		EG020A-F: Nickel	7440-02-0	1 mg/L	90.2	70.0	130					
		EG020A-F: Zinc	7440-66-6	1 mg/L	87.7	70.0	130					
EG035F: Dissolved Mercury by FIMS (QCLot: 5842139)				7439-97-6	0.01 mg/L	84.0	70.0	130				
EP080/071: Total Petroleum Hydrocarbons (QCLot: 5833350)				---	325 µg/L	73.3	70.0	130				
EP080/071: Total Recoverable Hydrocarbons - NEPM 2013 Fractions (QCLot: 5833350)				EP080: C6 - C10 Fraction	375 µg/L	71.0	70.0	130				
EP080: BTEXN (QCLot: 5833350)				ES2418166-001	Anonymous	EP080: Benzene	71-43-2	25 µg/L	90.4	70.0	130	
				EP080: Toluene	108-88-3	25 µg/L	87.2	70.0	130			
				EP080: Ethylbenzene	100-41-4	25 µg/L	87.4	70.0	130			
				EP080: meta- & para-Xylene	108-38-3 106-42-3	25 µg/L	86.9	70.0	130			
				EP080: ortho-Xylene	95-47-6	25 µg/L	89.1	70.0	130			
				EP080: Naphthalene	91-20-3	25 µg/L	105	70.0	130			

# G

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## EIL Calculation Spreadsheet

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<b>Inputs</b>
Select contaminant from list below
As
Below needed to calculate fresh and aged ACLs
Below needed to calculate fresh and aged ABCs
or for fresh ABCs only
or for aged ABCs only

<b>Outputs</b>		
Land use	Arsenic generic EILs (mg contaminant/kg dry soil)	
	Fresh	Aged
National parks and areas of high conservation value	20	40
Urban residential and open public spaces	50	100
Commercial and industrial	80	160

Inputs
Select contaminant from list below <b>Cr_III</b>
Below needed to calculate fresh and aged ACLs
Enter % clay (values from 0 to 100%) 12.28
Below needed to calculate fresh and aged ABCs
Measured background concentration (mg/kg). Leave blank if no measured value
or for fresh ABCs only Enter iron content (aqua regia method) (values from 0 to 50%) to obtain estimate of background concentration 7
or for aged ABCs only
Enter State (or closest State) NSW
Enter traffic volume (high or low) low

Outputs		
Land use	Cr III soil-specific EILs (mg contaminant/kg dry soil)	
	Fresh	Aged
National parks and areas of high conservation value	130	150
Urban residential and open public spaces	250	430
Commercial and industrial	360	720

Inputs	
Select contaminant from list below	
Cu	
Below needed to calculate fresh and aged ACLs	
Enter cation exchange capacity (silver thiourea method) (values from 0 to 100 cmolc/kg dwt)	
3	
Enter soil pH (calcium chloride method) (values from 1 to 14)	
4.65	
Enter organic carbon content (%OC) (values from 0 to 50%)	
0.285	
Below needed to calculate fresh and aged ABCs	
Measured background concentration (mg/kg). Leave blank if no measured value	
or for fresh ABCs only	
Enter iron content (aqua regia method) (values from 0 to 50%) to obtain estimate of background concentration	
7	
or for aged ABCs only	
Enter State (or closest State)	
NSW	
Enter traffic volume (high or low)	
low	

Outputs		
Land use	Cu soil-specific EILs (mg contaminant/kg dry soil)	
	Fresh	Aged
National parks and areas of high conservation value	25	25
Urban residential and open public spaces	30	40
Commercial and industrial	35	45

<b>Inputs</b>
Select contaminant from list below
DDT
Below needed to calculate fresh and aged ACLs
Below needed to calculate fresh and aged ABCs
or for fresh ABCs only
or for aged ABCs only

<b>Outputs</b>		
Land use	DDT generic EILs (mg contaminant/kg dry soil)	
	Fresh	Aged
National parks and areas of high conservation value	3	3
Urban residential and open public spaces	180	180
Commercial and industrial	640	640

<b>Inputs</b>
Select contaminant from list below
Naphthalene
Below needed to calculate fresh and aged ACLs
Below needed to calculate fresh and aged ABCs
or for fresh ABCs only
or for aged ABCs only

<b>Outputs</b>		
Land use	Naphthalene generic EILs (mg contaminant/kg dry soil)	
	Fresh	Aged
National parks and areas of high conservation value	10	10
Urban residential and open public spaces	170	170
Commercial and industrial	370	370

Inputs	
<b>Select contaminant from list below</b>	
Ni	
<b>Below needed to calculate fresh and aged ACLs</b>	
Enter cation exchange capacity (silver thiourea method) (values from 0 to 100 cmolc/kg dwt)	
3	
<b>Below needed to calculate fresh and aged ABCs</b>	
Measured background concentration (mg/kg). Leave blank if no measured value	
or for fresh ABCs only	
Enter iron content (aqua regia method) (values from 0 to 50%) to obtain estimate of background concentration	
7	
<b>or for aged ABCs only</b>	
Enter State (or closest State)	
NSW	
Enter traffic volume (high or low)	
low	

Outputs		
Land use	Ni soil-specific EILs (mg contaminant/kg dry soil)	
	Fresh	Aged
National parks and areas of high conservation value	25	7
Urban residential and open public spaces	30	15
Commercial and industrial	35	20

<b>Inputs</b>
Select contaminant from list below
Pb
Below needed to calculate fresh and aged ACLs
Below needed to calculate fresh and aged ABCs
or for fresh ABCs only
or for aged ABCs only

<b>Outputs</b>		
Land use	Lead generic ELLs (mg contaminant/kg dry soil)	
	Fresh	Aged
National parks and areas of high conservation value	110	470
Urban residential and open public spaces	270	1100
Commercial and industrial	440	1800

Inputs	
<b>Select contaminant from list below</b>	
Zn	
<b>Below needed to calculate fresh and aged ACLs</b>	
Enter cation exchange capacity (silver thiourea method) (values from 0 to 100 cmolc/kg dwt)	
3	
Enter soil pH (calcium chloride method) (values from 1 to 14)	
4.65	
<b>Below needed to calculate fresh and aged ABCs</b>	
Measured background concentration (mg/kg). Leave blank if no measured value	
or for fresh ABCs only	
Enter iron content (aqua regia method) (values from 0 to 50%) to obtain estimate of background concentration	
7	
or for aged ABCs only	
Enter State (or closest State)	
NSW	
Enter traffic volume (high or low)	
low	

Outputs		
Land use	Zn soil-specific EILs (mg contaminant/kg dry soil)	
	Fresh	Aged
National parks and areas of high conservation value	40	95
Urban residential and open public spaces	60	150
Commercial and industrial	80	190



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