

# Appendix R

## Preliminary Site Investigation







# **Douglas Partners**

*Geotechnics | Environment | Groundwater*

Report on  
Preliminary Site Investigation (Contamination)

Powerhouse Ultimo Renewal  
Powerhouse Ultimo, 500 Harris Street, Ultimo

Prepared for  
Department of Enterprise, Investment and Trade

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Integrated Practical Solutions



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## Executive Summary

This report presents the results of a Preliminary Site Investigation (Contamination) undertaken for the Powerhouse Ultimo Renewal project at the Powerhouse Ultimo, 500 Harris Street, Ultimo. The purpose of the investigation was to assess the contamination status of the site with respect to proposed land uses.

This investigation included soil sampling from 11 designated sampling points (Boreholes 101 to 109 and Boreholes 201 and 202). Boreholes 104 and 105 were designated as groundwater monitoring wells and were subject to one round of groundwater sampling. The investigation is considered to be preliminary in nature. Drawing 1, Appendix A shows the borehole locations. Fill was observed to be up to 5 m in depth, comprise various soil types, and commonly containing anthropogenic materials.

Soil samples were selected for analysis for potential contaminants including eight metals; total recoverable hydrocarbons (TRH); TRH with silica gel clean-up; benzene, toluene, ethylbenzene and xylenes (BTEX), polycyclic aromatic hydrocarbons (PAH); organochlorine pesticides (OCP); organophosphorus pesticides (OPP); polychlorinated biphenyls (PCB), phenols; volatile organic compounds (VOC); and asbestos. Groundwater samples from each groundwater monitoring well were analysed for potential contaminants including eight metals, TRH, BTEX, PAH, OCP, OPP, PCB, total phenols, ammonia and VOC.

The majority of the analytes were either not detected or found at low concentrations in the soil and groundwater samples analysed. Elevated concentrations of PAH, TRH, metals and OCP (above adopted assessment criteria) were identified in fill samples. In addition, an elevated concentration of ammonia in groundwater (above the adopted assessment criteria) was encountered at one sample location.

Overall, results indicate that remediation or management of contamination will be required for site development although the extent of remediation / management will likely depend on the layout of future site uses as well as the results of additional investigation.

The following is recommended for the Stage 2 detailed development application:

- A detailed site investigation (including soil and groundwater sampling).

The results of the detailed site investigation would be used to determine remediation or management requirements to address site contamination. On this basis, it is considered that the site can be made suitable for the proposed Powerhouse Ultimo Renewal development.



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## Report on Preliminary Site Investigation (Contamination)

### Powerhouse Ultimo Renewal

### Powerhouse Ultimo, 500 Harris Street, Ultimo

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## 1. Introduction

This report presents the results of a Preliminary Site Investigation (PSI) undertaken for the Powerhouse Ultimo Renewal project at the Powerhouse Ultimo, 500 Harris Street, Ultimo. This report was commissioned by the Department of Enterprise, Investment and Trade (ref: PROC 8007294 - 16) and was undertaken in accordance with Douglas Partners Pty Ltd (DP) proposal dated 23 February 2022 (ref: 86874.02.P.001.Rev1).

This report has been prepared to support a State Significant Development (SSD) Development Application (DA) for alterations and additions to Powerhouse Ultimo. It is understood that delivery of the Powerhouse Ultimo Renewal project will occur in stages, including a concept DA (Stage 1), an architectural design competition, and a detailed DA (Stage 2). Concept DA approval is being sought.

This report has been prepared to address item 18 of Planning Secretary's Environmental Assessment Requirements for the Powerhouse Ultimo Renewal project (application number SSD-32927319, issued on 18 January 2022), which states that *In accordance with SEPP 55, assess and quantify any soil and groundwater contamination and demonstrate that the site is suitable (or will be suitable, after remediation) for the development.* This report is the PSI listed as "documentation" under Condition 18.

The purpose of the PSI was to assess the contamination status of the site with respect to proposed land uses from the available soil and groundwater sampling (on the basis that untested parts of the site will be investigated for the Stage 2 detailed DA). The investigation was undertaken in conjunction with a geotechnical investigation by DP (ref: 86874.03.R.001.Rev3).

## 2. Scope of Works

The scope of works for the investigation was as follows:

- Review DP, *Desktop Study for Contamination, Powerhouse Museum Study of Alternate Uses, 500 Harris Street, Ultimo*, Prepared for CBRE Pty Ltd, September 2016 (ref: 85601.00.R.002.Rev0) [DP, 2016];
- Review Powerhouse Ultimo plans and Dial-Before-You-Dig plans to determine possible sampling locations;
- Set out a total of 11 borehole locations and scan each location for buried services using electromagnetic scanners;
- Collect soil samples at regular depths from each borehole drilled using a drilling rig and / or hand tools;
- Install groundwater monitoring wells at two boreholes;

- Screen all samples for volatile chemicals using a photoionisation detector (PID);
- Develop each groundwater monitoring well;
- Collect groundwater samples from each monitoring well using low flow techniques;
- Analysis of selected soil samples at a NATA accredited laboratory for various combinations of the following:
  - o Metals (arsenic, cadmium, chromium, copper, lead, mercury, nickel and zinc);
  - o Total recoverable hydrocarbons (TRH);
  - o TRH with silica gel clean-up;
  - o Benzene, toluene, ethylbenzene and xylenes (BTEX);
  - o Polycyclic aromatic hydrocarbons (PAH);
  - o Organochlorine pesticides (OCP);
  - o Organophosphorus pesticides (OPP);
  - o Polychlorinated biphenyls (PCB);
  - o Phenols (total);
  - o Volatile organic compounds (VOC);
  - o Asbestos (40 g soil sample);
  - o pH; and
  - o Cation exchange capacity (CEC).
- Analysis of two groundwater samples at a NATA accredited laboratory for metals, TRH, BTEX, PAH, OCP, OPP, PCB, total phenols, ammonia, VOC and hardness;
- Analysis of QA / QC samples including trip spikes, trip blanks and inter-laboratory replicate samples and intra-laboratory samples; and
- Preparation of this report.

### 3. Site Identification and Description

The site is defined by the following land parcels that are owned by The Trustees of the Museum of Applied Arts and Sciences:

- Lot 1, Deposited Plan 770031;
- Lot 1, Deposited Plan 781732;
- Lots 1 and 3, Deposited Plan 631345;
- Lot 3, Deposited Plan 216854; and
- Lot 37, Deposited Plan 822345.

Powerhouse Ultimo has a street address of 500 Harris Street, Ultimo. It is noted, however, that Lot 1 in Deposited Plan 770031 (the former post office building) has street address 494 Harris Street, Ultimo according to SIX Maps (NSW Government Spatial Services). Drawing 1, Appendix A shows the site location and approximate site boundary.



The site covers approximately 2.45 ha and adjoins William Henry Street (and Lot 1 Deposited Plan 801428) to the north; Harris Street, Macarthur Street and Omnibus Lane to the west, and Mary Ann Street to the south. The surrounding area is used for a mix of residential and commercial purposes. A light rail line and pedestrian access adjoins the eastern site boundary. The Ian Thorpe Aquatic Centre is present on the northern side of William Henry Street.

The Powerhouse Ultimo site comprises the following:

- The Harwood Building which is a large standalone structure at the south of the site that was formerly the Ultimo Tram Depot. The building has a basement, ground level and mezzanine. The basement is mainly used for storage of museum items. The building also has workshops, a laboratory and associated offices and amenities;
- Interlinked buildings at the north which include the Boiler House, Engine House, Turbine House, Switch House, Wran Building and North Annex. The buildings have seven levels (according to drawings provided by the client) and are used for museum exhibits and associated offices, education facilities and amenities. A courtyard is present at the north-west corner of the site (to the north of the Boiler House);
- The former Post Office which is a standalone building on the north-west corner of the site. A courtyard is present between the former Post Office and the Wran Building;
- The Grace Brothers Courtyard and Kiosk (café) at the east of the site;
- A gatehouse and vehicular access from Macarthur Street; and
- Parking areas located to the east and south of the Harwood Building.

It is understood that no substantive works or changes in use are proposed to the Harwood building located between Macarthur Street and Mary Ann Street.

## **4. Summary of Previous Contamination Report**

DP (2016) presents a desktop study for contamination which was undertaken to inform a study of future alternative uses for the site. Findings and identified potential sources of contamination presented in DP (2016) are summarised in the following subsections.

### **4.1 Regional Geology, Hydrogeology and Soils**

The Sydney 1:100 000 Geology Sheet indicates that the site is underlain by Hawkesbury Sandstone to the west and stream alluvial and estuarine sediment to the east as shown on Figure 1. Hawkesbury sandstone comprises medium to coarse grained quartz sandstone, very minor shale and laminite lenses. Stream alluvial and estuarine sediment comprises silty to peaty quartz sand, silt, and clay with ferruginous and humic cementation in places and common shell layers.



**Figure 1: Geology Sheet**

According to the Sydney 1:100 000 Soils Landscape Sheet, the site is located within the Gymea soil landscape where natural soils are formed from the erosion of rock.

The land surrounding the site has gentle slopes down to the east and north-east. Groundwater flow at the site is anticipated to be towards the north-east, in the general direction of Darling Harbour. Stormwater at the site is expected to enter the local stormwater system.

A search of the NSW Office of Water groundwater database revealed that there are no registered groundwater bores within 400 m of the site.

Based on published 1:25 000 Acid Sulfate Soil Risk mapping data (1994-1998), the site is an area of no known occurrences of acid sulfate soils. As shown on Figure 2, Disturbed Terrain is located approximately 40 m to the east of the site where the depth to acid sulfate soils is unknown and soil investigations are required to assess for acid sulfate soils.





**Figure 2: Acid Sulfate Soil Risk Mapping**

According to the Sydney Local Environment Plan 2012, Acid Sulfate Soils Map, the site is located in a 'Class 5' area. Acid sulfate soils are not typically found in Class 5 areas but are within 500 m of lands associated with a risk of acid sulfate soils (Class 1, 2, 3 or 4 areas). Works in a Class 5 area that are likely to lower the water table below 1 m AHD on adjacent Class 1, 2, 3 or 4 land will trigger the requirement for assessment and may require management.

## 4.2 Site History Information

### 4.2.1 Historical Aerial Photographs

Aerial photographs from 1930, 1943, 1955, 1970, 1982, 1991 and 2014 were reviewed to determine possible previous uses of the site and surrounding land.

The 1930 aerial photograph showed the power station at the northern part of the site (including the Boiler House, Engine House, Turbine House and Switch House) with two stacks at the southern part of the Boiler House and a separate stack at the north. The Post Office building was present at the western-most corner of the site. The tram depot building (with a saw-tooth roof) was present at the south of the site. An industrial / commercial building was present to the south of the Post Office building. Some of the eastern part of the site (adjoining Harris Street) and the southern end of the site appeared to have been undeveloped.

The 1943 aerial photograph showed that a building appeared to have been constructed to the south of the Boiler House since 1930. A commercial / industrial building had also been constructed at the eastern part of the site (at the corner of Harris Street and Macarthur Street). The land to the north of this building appears to have been used for a yard. A small structure was also present at the south-eastern corner of the site.

The 1970 aerial photograph showed that commercial / industrial buildings had been constructed at the western part of the site since 1955 (in the area that was previously used as a yard). The stack at the north of the site appeared to have been demolished for the realignment of the roadway to the north.

The 1991 aerial photograph showed that the site had been subject to significant change since 1982. New roofing was present on the Boiler House, Engine House, Turbine House and Switch House. The building to the south of the Boiler House had been demolished and replaced by a courtyard, walkways and small structures. Buildings at the eastern part of the site had been replaced by the Wran Building and the Forecourt.

The 2014 aerial photograph showed that the Kiosk had been constructed at the eastern part of the site since 1994.

#### **4.2.2 Historical Title Deeds**

A historical title deed search was conducted in order to provide an indication of possible previous site uses.

Between 1898 and 1964, Lot 3 Deposited Plan 216854 was owned by government entities (Minister for Public Works, Commissioner for Government Transport and Commissioner for Government Transport) presumably associated with the tramway and tram depot. The Trustees of the Museum of Applied Arts and Sciences were the registered owners from 1964.

Between 1924 and 1978, Lot 1 Deposited Plan 781732 was owned by commercial or industrial companies including Sydney Glass Company Limited, Hermann Haege Pty Limited, Carmichael Paper Agencies Pty Limited. The Trustees of the Museum of Applied Arts and Sciences were the registered owners from 1988.

Between 1923 and 1938, the majority of Lot 3 Deposited Plan 631345 was owned by Maurice Henry Newstead (Merchant). The majority of this Lot was then owned by Maize Products Proprietary Limited to 1967 and then Manassen Investments Pty Limited to 1980. The Trustees of the Museum of Applied Arts and Sciences were the registered owners from 1988.

Sydney Glass Company Limited owned the south-western part of Lot 1 Deposited Plan 631345 between 1924 and 1948, before being owned by Commissioner for Railways 1948 to 1983. From circa 1900 to 1983, the majority of Lot 1 Deposited Plan 631345 was owned by government entities (Minister for Public Works, Chief Commissioner for Railways and Tramways, Electricity Commission of New South Wales, Commissioner for Railways) presumably associated with power generation and the tramway. The Trustees of the Museum of Applied Arts and Sciences were the registered owners from 1988.

#### 4.2.3 Records for Storage of Hazardous Chemicals

A search of the SafeWork NSW database for the storage of hazardous chemicals at the site was requested. The search returned documents for record number 35/034632.

According to a licence amendment application to keep dangerous goods at 500 Harris Street, Ultimo, dated 20 June 2002, there were four dangerous goods storage depots including a roofed store (Depot 1), and three flammable liquids cabinets (Depots 2 to 4). A list of 14 other storage areas (identified as A, B, C, D, E, F, H, I, J, L, M, O, P and Q) was also provided.

According to the plan attached to the application, the roofed store (Depot 1) was located at the eastern side of the Harwood Building and was used to store petroleum spirit (typically 100 L), paint or paint thinners (typically 100 L), kerosene (typically 200 L), turpentine substitute (typically 160 L), toluene (typically 25 L), ethanol (typically 20 L) and nitrocellulose solution (typically 200 L). Depot 2 was located at the western part of the Boiler House basement and this cabinet was used to store paint (typically 160 L) and kerosene (typically 60 L). Depot 3 and 4 were located at the northern part of the Harwood Building and these cabinets were used to store paint (typically 160 L at each Depot), kerosene (typically 60 L at Depot 3) and turpentine substitute (typically 60 L at Depot 4).

Other storage areas identified at the Harwood Building (according to the plan attached to the application) were listed as:

- Flammable liquid cabinets (A, B, C, E, F, I);
- Cabinet (H);
- Roofed store (D);
- Gas cylinder store (J, L, M); and
- Roofed store (O).

The roofed store (O) was shown to be next to Depot 1 and was used for the storage of 7500 L of combustible oil.

Other storage areas identified at the Boiler House basement were listed as:

- Flammable liquid cabinet (P); and
- Roofed store (Q).

#### 4.2.4 Council Records

Selected documents and plans relating to 500 Harris Street, Ultimo were retrieved by Council for review. Relevant site history information obtained from the review was summarised as follows:

- According to a document stamped May 1980, it was noted that for 64 years, the Power House had generated electricity for the tramway and provided power for domestic and industrial purposes. The Power House had been vacant since 1963;
- An Interim Development Application, dated May 1981, was for major alterations and additions to the Ultimo Tram Depot and Power House Building;



- A Development Application, dated April 1987, was for major alterations and additions to the Ultimo Tram Depot and Power House Building; and
- A Draft Environmental Impact Statement (associated with the above-listed Development Application) for the northern part of the site concluded that no significant negative impacts on the local environment were associated with the proposed Power House Museum. The Darling Harbour Goods Yard and Darling Harbour Goods Railway were noted to be present at neighbouring land to the east of the site at that time.

#### 4.2.5 Previous Reports

According to Government Architect's Office (NSW), *Powerhouse: a study of the relocation of the Museum of Applied Arts and Sciences, in a development of the Ultimo Power Station and the Ultimo Tramway Depot, Sydney, 1978*:

- Site drilling was carried out to establish the nature of the foundation material and the level of hard stratum. Sandstone was located at an average of 4 m below the ground level on the region of the Boiler House, Turbine Hall and Coal Store Bunker. On the Tram Depot site, an investigation carried out in 1977 indicated hard stratum at an average 'level' of 3 m;
- The Ultimo Power Station was the original source of electricity for the Sydney tramway system. The original building dates from 1899, with later additions in 1916. A major reconstruction begun in 1927 and was completed in 1931. The station ceased operation in circa 1960 and was damaged during subsequent demolition, salvage operations, decay and vandalism. The power station consisted primarily of two adjacent halls, each approximately 85 m long by 25 m wide. The western hall (Turbine Hall) contained the engine room and turbine room. The eastern hall was originally the Boilerhouse and had one major floor positioned level with the old railway track diversion which ran into the building from the Darling Harbour Goods Yard lines. Towards the southern end of the Boilerhouse were two 5 m diameter masonry stacks which were then recently demolished to approximately roof level. Running down the centre of the Boilerhouse for almost the entire length of the hall were large steel coalhoppers. The main floor level was penetrated in a number of areas to a deep basement level below. An office annex was located at the northern end of the western hall. A switchhouse, approximately 60 m long by 20 m wide, was located at the southern end of the western wall;
- Between the power station and Harris Street a number of one and two storey commercial buildings, a petrol service station and the Ultimo Post Office were located;
- The Ultimo Tram Depot was a large single-level building for tramcar storage, constructed in 1910. The Small brick amenity areas were also present at the Darling Harbour railway boundary with a two-storey brick building on the Mary Ann Street frontage; and
- At 496-502 Harris Street (presumably the western part of the site, now part of 500 Harris Street), there was an office and warehouse building of three levels, including first floor offices, a ground floor warehouse area and basement. Adjoining this area were various older-style warehouses and workshops. The lease to Ampol at the southern section of the property was to expire in February 1979.

According to Architectural Projects Pty Ltd, *Conservation Management Plan for The Powerhouse Museum*, 2002:

- The location of the Powerhouse near the head of Darling Harbour gave it an adequate supply of salt water for condensers and proximity to the goods yards for coal supply and ash disposal;
- In 1980, the NSW government decided to develop the Power House and Tram Depot into the Museum. The Stage 1 work for the project saw the rebuilding and refurbishment of the Tram Depot. The Harwood building became a purpose-built museum structure. Photographic evidence indicated that nothing remained of the lines or tramway trenches at the Harwood building, although it was noted that a tiny section of track could be viewed from relatively inaccessible service areas;
- Stage 2 works included refurbishment of the power station and construction of The Wran Building (The Wran Building) which was opened in 1988; and
- The former Ultimo Post Office, built by 1901 was adapted to house a child care centre (at the time of preparing the conservation management plan).

Hibbs & Associates Pty Ltd, *Powerhouse Museum Asbestos Survey*, September 2014, provided a register and qualitative risk assessment of building materials containing asbestos. The survey did not include an assessment of potential contamination of soil from asbestos. It is noted that no friable asbestos was encountered during the inspection and no asbestos material (bonded asbestos) required short to medium term remedial works.

#### 4.3 Site Walkover Observations

The site was subject to a walkover by a DP Environmental Engineer on 16 August 2016. The walkover was limited to external areas and ground and basement level areas that were shown by museum maintenance staff at that time. Not all ground and basement level areas were accessed. Noteworthy observations are listed as follows:

- The remnant stack at the northern end of the site could be observed from Pier Street (above). Litter and (probably) soil were observed to partially fill the remnant stack;
- The pedestrian access way at the eastern site boundary had been improved and a remnant railway track was observed to align with the Kiosk;
- A sandstone cutting was observed at the exterior to a fire passage at the western side of Level 3 of the Wran Building;
- Brick structures, similar in appearance to fireplaces, were observed at lowest part of the north-east corner of the Boiler House. These may have been used for the collection of ash from chutes above. The base of these structures appeared to be concrete;
- Two stacks were present at the southern end of the Boiler House. The inside of the easternmost stack appeared to have a concrete base. The inside of the other stack was not observed;
- The basement (Level 1) of the former power station housed boilers, pumps and a series of plant rooms. This area appeared to be well kept with concrete floors generally absent of staining. Chemicals were kept in cabinets. Water was observed in open drains. A room was used for the storage of chlorine and was next to the seawater room which had a grid mesh floor;
- Soil and rubble were observed at the southern end of the Harwood building basement. A possible remnant tram track was also observed amongst the soil and rubble;

- Glass pipes (amongst other pipes) were observed at the perimeter of the Harwood building basement, presumably to drain wastewater (potentially containing corrosives) from the laboratory (to a location unknown to the maintenance worker at the time); and
- Roots from trees at the courtyard next to the Kiosk appeared to be causing damage to the courtyard surface. It was understood that a concrete slab exists beneath the soil in which the trees were planted.

It was understood at the time of the inspection that there are no fuel storage tanks at the site.

#### 4.4 Potential Sources of Contamination and Potential Contaminants

From a review of site information and the site walkover, identified potential sources of contamination and potential contaminants were summarised as in Table 1.

**Table 1: Potential Sources of Contamination and Potential Contaminants**

Potential Source of Contamination	Potential Contaminants	Comments
Coal ash from the previous power station operation.	Ash can contain metals and PAH.	Most of the ash is likely to have been removed and disposed off-site during the operation of the power station, however, some ash may have been used as filling or been placed (stockpiled) on surface soils during the power station's operation.
Former tram depot and railway use.	Total petroleum hydrocarbons (TPH), PAH, metals, phenolics (creosote), nitrates, ammonia and asbestos.	Asbestos may have been used in brake pads.
(Former) Switch House and other areas that housed electrical equipment.	PCB.	PCB are associated with transformers and capacitors.
Previous demolition of buildings.	Hazardous building materials including asbestos, lead and PCB.	Significant demolition occurred prior to the development of the museum, particularly at the western part of the site. Surface soils may have been impacted during demolition.
Imported contaminated filling to form the site.	Various possible such as TPH, PAH, metals, phenols, pesticides, asbestos and PCB.	The sandstone cutting and existing basement constructions indicated significant filling across much of the site is unlikely. Filling was considered more likely to be present at eastern parts of the site (where sandstone cuttings were not observed).



Potential Source of Contamination	Potential Contaminants	Comments
Previous petrol service station	TPH, VOC, lead, PAH and phenols.	The exact location of previous service station (present in 1978) is not known, but was adjacent Harris Street, to the south of the Post Office. A property at this part of the site was leased to Ampol. The lease was probably terminated in 1979. Potential TPH and VOC contamination may have dispersed since the service station ceased operation.
Previous general industrial use and workshops	Various possible such as TPH, VOC and metals.	Apart from the previous petrol service station, the western part of the site was previously used for (unknown) industrial purposes and workshops.
Chemicals including oils used by the museum (particularly at the Harwood Building which has workshops and a laboratory)	VOC and TPH	Contamination from chemicals and oils used in current operations is considered unlikely given that the museum appears to be well kept and maintained.

## 5. Preliminary Conceptual Site Model

A Conceptual Site Model (CSM) is a representation of site-related information regarding contamination sources, receptors and exposure pathways between those sources and receptors. The CSM provides the framework for identifying how the site became contaminated and how potential receptors may be exposed to contamination either in the present or the future i.e., it enables an assessment of the potential source - pathway - receptor linkages.

### 5.1 Potential Contamination Sources

Potential contamination sources at the site are listed in Section 4.4. No obvious potential off-site sources of contamination were identified in DP (2016).

### 5.2 Potential Receptors

Given that the proposed development layout is yet to be confirmed, potential future receptors to contamination cannot be clearly established, however, in general, potential receptors of contamination for the proposed development may include:

- (R1) Future site users (guests, workers and visitors);
- (R2) Construction workers (for the proposed development);
- (R3) Future maintenance workers (post-construction);

- (R4) Adjacent land users (including residents, pedestrians, workers and visitors);
- (R5) Groundwater;
- (R6) Surface water body (Darling Harbour);
- (R7) Terrestrial ecosystem; and
- (R8) In ground building structures.

### 5.3 Potential Pathways

Potential pathways for contamination to impact receptors include:

- (P1) Ingestion and dermal contact;
- (P2) Inhalation of dust;
- (P3) Inhalation of vapours;
- (P4) Surface water runoff (particularly during construction);
- (P5) Leaching of contaminants and vertical migration into groundwater;
- (P6) Lateral migration of groundwater;
- (P7) Contact of contaminated ground with terrestrial organism; and
- (P8) Contact of contaminated ground within ground structures.

### 5.4 Preliminary Conceptual Site Model

A 'source - pathway - receptor' approach has been used to assess the potential risks of harm being caused to human or environmental receptors from contamination sources via exposure pathways (complete pathways). The possible pathways between the above listed sources and receptors are provided in Table 2.

**Table 2: Preliminary Conceptual Site Model**

Source	Transport Pathway	Receptor
Contaminated ground from previous and current site use (see Table 1, Section 4.4 for list of potential sources)	P1 - Ingestion and dermal contact	R1 - Future site users
	P2 - Inhalation of dust	R2 - Construction workers
	P3 - Inhalation of vapours	R3 - Future maintenance workers
	P2 - Inhalation of dust	R4 - Adjacent site users
	P3 - Inhalation of vapours	R4 - Adjacent site users
	P5 - Leaching of contaminants and vertical migration into groundwater	R5 - Groundwater
	P4 - Surface water runoff	R6 - Surface water body
	P6 - Lateral migration of groundwater	R6 - Surface water body
	P7 - Contact of contaminated ground with terrestrial organisms	R7 - Terrestrial ecosystem
	P8 - Contact of contaminated ground within ground structures	R8 - In ground building structures

## 6. Field Work and QA / QC

### 6.1 Sample Locations and Rationale

The site covers approximately 2.45 ha. According to NSW EPA, *Sampling Design Guidelines*, 1995, a minimum of 35 systematic sampling points is recommended to characterise a site of this size. This investigation included 11 designated sampling points and, therefore, this investigation is considered to be preliminary in nature. Positioning of boreholes was limited by existing structures and site operations (i.e., for the museum). Boreholes 101 to 106, 201 and 202 were positioned for both geotechnical and contamination assessment purposes to provide coverage of external areas that were accessible to a drilling rig. To provide some further site coverage, Borehole 107 was positioned at the external area at the northern corner of the site (which was not accessible to a drilling rig) and Boreholes 108 and 109 were positioned in the basement of the Harwood Building.

Boreholes 104 and 105 were designated as groundwater monitoring wells as these were drilled to depths at which groundwater was likely to be encountered on the lower side of the site.

### 6.2 Soil Sampling Procedures

Soil sampling was undertaken on 22, 23 and 28 October 2019 (Boreholes 101 to 109); and 15 and 26 November 2019 (Boreholes 201 and 202).

Boreholes 101 to 106 and 202 were drilled using Bobcat-mounted drill rig and Borehole 201 was drilled using a truck-mounted drill rig. Soil samples of the fill were collected from the solid flight auger at regular depth intervals (at least one soil sample per metre depth). At least one sample of natural soil was collected from the solid flight auger where natural soil was encountered.

Boreholes 107, 108 and 109 were drilled using hand tools. Soil samples of fill were collected from the hand auger where fill was encountered.

All sampling data was recorded on DP's borehole logs provided in Appendix B which also has notes about this report. The general sampling procedure adopted for the collection of soil samples for chemical analysis was:

- Collect soil samples directly from the solid flight auger or hand auger using disposable gloves;
- Transfer samples into laboratory-prepared glass jars with Teflon-lined lids, filled to minimise the headspace within the sample jar, and capping immediately to minimise the loss of volatiles;
- Label sample containers with individual and unique identification, including project number, sample location and sample depth; and
- Place the glass jars into a cooled, insulated and sealed container for transport to the laboratory.

Replicate samples were collected in zip-lock bags for screening of VOC using a calibrated PID and asbestos analysis. Samples were also collected in zip-lock bags for asbestos analysis.

### 6.3 Groundwater Monitoring Well Installation and Development

Groundwater monitoring well construction details are provided on the borehole logs in Appendix B.

The wells were constructed of 50 mm diameter acid washed, Class 18, PVC casing and machine slotted well screen intervals. Joints were screw threaded, thereby avoiding the use of glues and solvents which may contaminate the groundwater. The wells were completed with a gravel pack extending above the well screen and then a bentonite plug. A Gatic cover and concrete was used at the ground surface to complete the installation.

Each groundwater monitoring well was developed (on 13 November 2019) by using a submersible pump. Prior to groundwater well development, an interface dip-meter was used to measure the groundwater level and check for light non-aqueous phase liquids (LNAPL).

### 6.4 Groundwater Sampling

Prior to groundwater sample collection, an interface dip-meter was used to measure the groundwater level and check for light non-aqueous phase liquids (LNAPL).

Groundwater sampling at Borehole 104 and 105 was undertaken (on 15 November 2019) using a low-flow geo-pump (peristaltic pump) and disposable tubing, following stabilisation (or near stabilisation) of field parameters. Field parameters were obtained using a calibrated water quality meter with probes placed inside a flow-through cell. The field parameters included temperature, dissolved oxygen (DO), electrical conductivity (EC), pH, oxidation reduction potential (redox) and turbidity. Disposable sampling



equipment (tubing) was used between collection of samples to avoid the need for decontamination. Samples were collected from the upper part of the water column in each well.

Samples were collected in laboratory prepared bottles and vials. The groundwater samples collected for metals testing were filtered in the field through a 45 µm membrane filter into nitric acid preserved bottles. Sample containers were labelled with individual and unique identification, including project number and sample number. Samples were placed in cooled, insulated and sealed containers for transport to the laboratory.

Groundwater field sheets were used to record groundwater depths, sample depths, field parameters, observations and samples collected. The groundwater field sheets are provided in Appendix C.

## 6.5 Quality Assurance and Quality Control

The field QC procedures for sampling were undertaken as prescribed in DP's *Field Procedures Manual* and included the use of trip spikes and trip blanks as well as the collection of blind replicate samples. The results of field QA / QC procedures as well as a discussion of data quality objectives (DQO) and data quality indicators (DQI) for the assessment are provided in Appendix D.

The analytical laboratories, accredited by NATA, are required to conduct in-house QA / QC procedures. These are normally incorporated into every analytical run and include reagent blanks, spike recovery, surrogate recovery and duplicate samples. These results are included in the laboratory certificates in Appendix E and results are discussed in Appendix D.

## 7. Site Assessment Criteria

### 7.1 Soil

The site assessment criteria applied to soils are informed by the preliminary CSM which identified receptors to potential contamination at the site. Analytical results were assessed (as a Tier 1 assessment) against the site assessment criteria comprising the investigation levels, screening levels and management limits of Schedule B1 of the National Environment Protection Council, *National Environment Protection (Assessment of Site Contamination) Measure* 1999, as amended 2013 (NEPC, 2013). The investigation levels, screening levels and management limits are applicable to generic land use settings and include consideration of, where relevant, the soil type and the depth of contamination. The investigation levels, screening levels and management limits are not intended to be used as clean up levels. Rather, they establish concentrations above which further appropriate investigation (e.g., Tier 2 assessment) should be undertaken. They are intentionally conservative and are based on a reasonable worst-case scenario.

The site assessment criteria for soil adopted for this investigation are listed in the following sub-sections.

### 7.1.1 Health Investigation Levels

Health investigation levels (HIL) are applicable to assessing health risk arising via all relevant pathways of exposure for a range of metals and organic substances. The HIL are generic to all soil types and apply generally to a depth of 3 m below the surface. From a contamination assessment perspective, 'information and education facilities' are considered to be commercial usage. However, as the project is at the conceptual stage, for the purpose of this assessment we have assumed a potential partial residential type land use (such as accommodation) incorporated into the Powerhouse Ultimo Renewal project. Therefore, HIL B (applicable to a residential land use scenario with minimal opportunities for soil access) and HIL D (applicable to a commercial or industrial land use scenario) have been adopted as site assessment criteria. The adopted HIL are shown in Table 3.

**Table 3: Health Investigation Levels**

Contaminant		HIL B Residential (mg/kg)	HIL D Commercial/Industrial (mg/kg)
Metals	Arsenic	500	3000
	Cadmium	150	900
	Chromium (VI)	500	3600
	Copper	30 000	240 000
	Lead	1200	1 500
	Mercury (inorganic)	120	80
	Nickel	1200	6 000
	Zinc	60 000	400 000
PAH	Benzo(a)pyrene TEQ <sup>1</sup>	4	40
	Total PAH	400	4000
Phenol	Pentachlorophenol	130	660
	Phenol	45 000	240 000
	Cresols	4 700	25 000
OCP	Aldrin + Dieldrin	10	45
	Chlordane	90	530
	DDT+DDE+DDD	600	3600
	Endosulfan	400	2000
	Endrin	20	100
	Heptachlor	10	50
	HCB	15	80
	Methoxychlor	500	2500
OPP	Chlorpyrifos	340	2000
Other Organics	PCB <sup>2</sup>	1	7

Notes: <sup>1</sup> sum of carcinogenic PAH

<sup>2</sup> non-dioxin-like PCB only

### 7.1.2 Health Screening Levels for Vapour Intrusion

Health screening levels (HSL) for vapour intrusion are applicable to selected petroleum compounds and fractions to assess the risk to human health via the inhalation pathway. HSL have been developed for different land uses, soil types and depths to contamination.

The proposed ground floor or basement use of the proposed development is likely to be for non-residential purposes, and, therefore, HSL D (for a commercial or industrial land use scenario) have been adopted as site assessment criteria. Given that ground-level residential use has not been ruled out at this stage, HSL B (for a residential land use scenario with minimal opportunities for soil access) have also been adopted as site assessment criteria (for comparative purposes). The adopted HSL shown in Table 4 are for contamination at a depth of 0 m to <1 m, and are the most conservative for sand, silt or clay soils given that various soil types are present at the site. Less conservative HSL may be appropriate depending on the contamination depth and actual soil type.

**Table 4: Health Screening Levels for Vapour Intrusion**

Contaminant		HSL B Residential (mg/kg)	HSL D Commercial /Industrial (mg/kg)
PAH	Naphthalene	3	NL
TRH	TPH C <sub>6</sub> -C <sub>10</sub> less BTEX	40	250
	TPH >C <sub>10</sub> -C <sub>16</sub> less Naphthalene	110	NL
BTEX	Benzene	0.5	3
	Toluene	160	NL
	Ethylbenzene	55	NL
	Xylenes	40	230

Note: The soil saturation concentration is defined as the soil concentration at which the porewater phase cannot dissolve any more of an individual chemical. The soil vapour that is in equilibrium with the porewater will be at its maximum. If the derived soil HSL exceeds C<sub>sat</sub>, a soil vapour source concentration for a petroleum mixture could not exceed a level that would result in the maximum allowable vapour risk for the given scenario. For these scenarios, no HSL is presented for these chemicals and the HSL is shown as 'not limiting' or 'NL'.

### 7.1.3 Ecological Investigation Levels

Ecological investigation levels (EIL) have been derived for selected metals and organic compounds and are applicable for assessing risk to terrestrial ecosystems (NEPC, 2013). EIL depend on specific soil physiochemical properties and land use scenarios and generally apply to the top 2 m of soil, which corresponds to the root zone and habitation zone of many species. The EIL is determined for a contaminant based on the sum of the ambient background concentration (ABC) and an added contaminant limit (ACL). The ABC of a contaminant is the soil concentration in a specific locality that is the sum of naturally occurring background levels and the contaminants levels that have been introduced from diffuse or non-point sources (e.g., motor vehicle emissions). The ACL is the added concentration (above the ABC) of a contaminant above which further appropriate investigation and evaluation of the impact on ecological values is required.

The EIL is calculated using the following formula:

$$\text{EIL} = \text{ABC} + \text{ACL},$$

The ABC is determined through direct measurement at an appropriate reference site or through the use of methods defined by Olszowy et al, *Trace element concentrations in soils from rural and urban areas of Australia*, Contaminated Sites monograph no. 4, South Australian Health Commission, Adelaide, Australia 1995 (Olszowy, 1995) or Hamon et al, *Geochemical indices allow estimation of heavy metal background concentrations in soils*, Global Biogeochemical Cycles, vol. 18, GB1014, (Hamon, 2004). ACL is based on soil characteristics.

An *Interactive (Excel) Calculation Spreadsheet* may be used for calculating site-specific EIL and has been provided in the ASC NEPM Toolbox available on the NEPC website.

The adopted EIL, derived from the *Interactive (Excel) Calculation Spreadsheet* are shown in Table 5.

The following input data and assumptions have been used to determine the adopted EIL:

- Given that all elements of the proposed development are yet to be confirmed, adopted EIL for urban residential and commercial / industrial land use scenarios is have been adopted;
- Given the site history, the contamination is considered as “aged” (>2 years);
- NSW is the state;
- The traffic volume is high;
- A pH of 9 has been use as an input value. This is the average of the three pH values obtained from testing of fill (see laboratory certificates 229301 and 229472, Appendix E, for test results);
- A CEC of 22 meq/100g has been adopted. This is the rounded average of the three CEC values obtained from testing of fill (see laboratory certificates 229301 and 229472, Appendix E, for test results);
- An organic carbon content of 1% has been adopted as a default value in the absence of site-specific data;
- An ABC of 160 mg/kg for lead has been adopted from Table 67 of Schedule B5c of NEPC (2013); and
- A clay content of 1% has been used as a conservative value in the absence of site-specific test results.

**Table 5: Ecological Investigation Levels**

<b>Contaminant</b>		<b>EIL Urban Residential (mg/kg)</b>	<b>EIL Commercial / Industrial (mg/kg)</b>
Metals	Arsenic	100	160
	Copper	240	340
	Nickel	290	490
	Chromium III	200	320
	Lead	1300	2000
	Zinc	870	1300
PAH	Naphthalene	170	370
OCP	DDT	180	640



It is noted that EIL do not apply at areas of the site that will have limited ecological value under the proposed development (such as at basements, building footprints, etc.).

#### 7.1.4 Ecological Screening Levels

Ecological screening levels (ESL) are used to assess the risk of selected hydrocarbon compounds to terrestrial ecosystems. ESL generally apply to the top 2 m of the soil profile as for EIL. ESL have been derived in NEPC (2013) for TPH fractions as well as BTEX and benzo(a)pyrene. Given that all elements of the proposed development are yet to be confirmed, adopted ESL for urban residential and commercial / industrial land use scenarios is have been adopted. The ESL shown in Table 6 are the more conservative for coarse and fine texture soils given that various soil types are present at the site.

**Table 6: Ecological Screening Levels**

Contaminant		ESL – Urban Residential (mg/kg)	ESL – Commercial / Industrial (mg/kg)
TRH	C <sub>6</sub> -C <sub>10</sub> (less BTEX)	180*	215*
	>C <sub>10</sub> -C <sub>16</sub>	120*	170*
	>C <sub>16</sub> -C <sub>34</sub>	300	1700
	>C <sub>34</sub> -C <sub>40</sub>	2800	3300
BTEX	Benzene	50	75
	Toluene	85	135
	Ethylbenzene	70	165
	Xylenes	105	180
PAH	Benzo(a)pyrene	0.7	1.4

Note: ESL are low reliability apart from those marked with \* which indicates that the ESL is of moderate reliability.

It is noted that ESL do not apply at areas of the site that will have limited ecological value under the proposed development (such as at basements, building footprints, etc.).

#### 7.1.5 Management Limits

In addition to appropriate consideration and application of the health-based and ecological assessment criteria, there are additional considerations which reflect the nature and properties of petroleum hydrocarbons, including:

- Formation of observable light non-aqueous phase liquid (LNAPL);
- Fire and explosion hazards; and
- Effects on buried infrastructure e.g., penetration of, or damage to, in-ground services.

'Management limits' to avoid or minimise these potential effects have been adopted in NEPC (2013) as Tier 1 guidance. Given that all elements of the proposed development are yet to be confirmed, management limits for a commercial / industrial land use scenario and an urban residential scenario have been adopted. The more conservative management limits for coarse and fine texture soils have been adopted given that various soil types are present at the site. Management limits may apply to any depth within the soil profile and are shown in Table 7.

**Table 7: Management Limits**

<b>Analyte</b>		<b>Management Limit Residential (mg/kg)</b>	<b>Management Limit Commercial / Industrial (mg/kg)</b>
TRH	C <sub>6</sub> – C <sub>10</sub>	700	700
	>C <sub>10</sub> -C <sub>16</sub>	1000	1000
	>C <sub>16</sub> -C <sub>34</sub>	2500	3500
	>C <sub>34</sub> -C <sub>40</sub>	10 000	10 000

### 7.1.6 Asbestos in Soil

A detailed asbestos assessment has not been undertaken. Visual observations for possible asbestos-containing materials (ACM) and the presence / absence of asbestos at a limit of reporting of 0.1 g/kg have been adopted as assessment criteria for this investigation as an initial screen.

## 7.2 Groundwater

The assessment criteria used for contamination in groundwater are based on the potential uses or risks posed by contaminated groundwater at or down-gradient of the site. The potential receptors (informed by the preliminary CSM) are considered to be:

- The surface water body ecosystem at Darling Harbour (a marine water ecosystem);
- Recreational water use at Darling Harbour; and
- Human receptors (site users and adjacent site users) via the vapour intrusion pathway.

The adopted assessment criteria for groundwater based on the above potential receptors are listed in the following sub-sections.

### 7.2.1 Default Guideline Values

For the protection of surface water body ecosystems, freshwater default guideline values (DGV) recommended for a slightly to moderately disturbed ecosystem or, otherwise, an unknown level of species protection from the Australian and New Zealand Governments (ANZG), Australian and New Zealand Guidelines for Fresh and Marine Water Quality website (accessed on 8 March 2022) have been adopted. The DGV are shown in Table 8.

**Table 8: Default Guideline Values**

<b>Contaminant</b>		<b>Marine Water DGV (µg/L)</b>
Metals	Cadmium	0.7
	Chromium (III)	27
	Chromium (VI)	4.4
	Copper	1.3
	Lead	4.4
	Mercury	0.1
	Nickel	7
	Zinc	8
PAH	Anthracene	0.01
	Naphthalene	50
	Fluoranthene	1
	Benzo(a)pyrene	0.1
	Phenanthrene	0.6
BTEX & VOC	Benzene	500
	Toluene	180
	Ethylbenzene	80
	m-xylene	75
	o-xylene	350*
	p-xylene	200*
	Isopropylbenzene	30
	Tetrachloroethene	70
	Trichloroethene	330
	1,1-Dichloroethene	700
	Chloroethene (vinyl chloride)	100
	1,2,4-Trichlorobenzene	20
	1,2-Dichlorobenzene	160*
	1,3-Dichlorobenzene	260*
	1,4-Dichlorobenzene	60*
	1,1,2,2-Tetrachloroethane	400
	1,1,1-Trichloroethane	270
	1,1,2-Trichloroethane	1900
	1,2-Dichloroethane	1900
	Carbon tetrachloride	240
	Chloroform	370
	1,2-Dichloropropane	900
	1,3-Dichloropropane	1100
	Monochlorobenzene (chlorobenzene)	55
OCP	Aldrin	0.003
	Chlordane	0.001
	DDT	0.0004
	Dieldrin	0.01*
	Endosulfan	0.005
	Endrin	0.004
	Heptachlor	0.0004
	Methoxychlor	0.004

Contaminant		Marine Water DGV (µg/L)
OPP	Azinphos methyl	0.01*
	Chlorpyrifos	0.01
	Diazinon	0.01*
	Dimethoate	0.15
	Fenitrothion	0.001
	Malathion	0.05*
	Parathion	0.004*
PCB	Aroclor 1242	0.3*
	Aroclor 1254	0.01*
Non-metallic Inorganics	Ammonia (total) as N	5070 at pH 6.6**

Note: \* DGV for fresh water in the absence of marine water DGV.

\*\* DGV can be adjusted for pH. The DGV for a pH of 6.6 has been adopted based on field results.

Note that there are DGV for individual phenolic compounds which are not listed in Table 8.

### 7.2.2 Guidelines for Recreational Water

The National Health and Medical Research Council (NHMRC), *Guidelines for Managing Risks in Recreational Water*, 2008 adopts drinking water guideline values for managing risks in recreational waters. It is stated, however, that guideline values applicable for drinking water quality are based on a daily consumption of 2 L and when applying these values to recreational water exposure, consumption of 0.1 to 0.2 L per day should be taken into consideration. The adopted guidelines for the protection of recreational water shown in Table 9 are sourced from the National Health and Medical Research Council and National Resource Management Ministerial Council (NHMRC & NRMCC), *Australian Drinking Water Guidelines Paper 6 National Water Quality Management Strategy*, 2011, updated January 2022, with health-based drinking water guideline values multiplied by 10 to account for consumption for recreational water exposure.



**Table 9: Guidelines for Managing Risks to Users of Recreational Waters**

Contaminant		Guideline for Recreational Water (µg/L)	
		Health	Aesthetic
Metals	Arsenic (total)	100	-
	Cadmium	20	-
	Chromium (VI)	500	-
	Copper	20 000	1000
	Lead	100	-
	Mercury	10	-
	Nickel	200	-
	Zinc	-	3000
PAH	Benzo(a)pyrene	0.1	-
BTEX & VOC	Benzene	10	-
	Toluene	8000	25
	Ethylbenzene	3000	3
	Xylene (total)	6000	20
	Tetrachloroethene	500	-
	1,1-Dichloroethene	300	-
	1,2-Dichloroethene	600	-
	1,3-Dichloropropene	1000	-
	Chloroethene (vinyl chloride)	3	-
	Chlorobenzene	3000	10
	Trichlorobenzenes (total)	300	5
	1,2-Dichlorobenzene	15 000	1
	1,3-Dichlorobenzene	-	20
	1,4-Dichlorobenzene	400	0.3
	1,2-Dichloroethane	30	-
	Carbon tetrachloride	30	-
	Trihalomethanes (total)	2500	-
	Styrene	300	4
	Hexachlorobutadiene	7	-
OCP	Chlordane	20	-
	DDT	90	-
	Aldrin & Dieldrin (combined)	3	-
	Endosulfan	200	-
	Heptachlor & Heptachlor Epoxide	3	-
	Methoxychlor	3000	-
OPP	Azinphos methyl	300	-
	Chlorpyrifos	100	-
	Diazinon	40	-
	Dichlorovos	50	-
	Dimethoate	70	-
	Fenitrothion	70	-
	Malathion	700	-
	Parathion	200	-
	Methyl parathion	7	-
	Bromophos ethyl	100	-
	Ethion	40	-
Non-metallic Inorganics	Ammonia	-	500

Note that there are guidelines for individual phenolic compounds which are not listed in Table 9.

### 7.2.3 Health Screening Levels for Vapour Intrusion

The proposed ground floor or basement use of the proposed development is likely to be for non-residential purposes, and, therefore, HSL D (for a commercial or industrial land use scenario) have been adopted as site assessment criteria. Given that ground-level residential use has not been ruled out at this stage, HSL B (for a residential land use scenario with minimal opportunities for soil access) have also been adopted as site assessment criteria (for comparative purposes). The adopted HSL shown in Table 10 are for groundwater at a depth of 2 m to <4 m; and are the most conservative for sand, silt and clay. Less conservative HSL may be applicable where groundwater is present at the site at a depth of 4 m or greater, however, (less conservative) site specific HSL may need to be considered where groundwater is within 2 m of the lowest floor level of the proposed building(s) or groundwater is to contact the proposed basement.

**Table 10: Health Screening Levels for Vapour Intrusion**

Contaminant		HSL B Residential (µg/L)	HSL D Commercial / Industrial (µg/L)
PAH	Naphthalene	NL	NL
TRH	TRH C <sub>6</sub> -C <sub>10</sub> less BTEX	1000	6000
	TRH >C <sub>10</sub> -C <sub>16</sub> less Naphthalene	1000	NL
BTEX	Benzene	800	5000
	Toluene	NL	NL
	Ethylbenzene	NL	NL
	Xylenes	NL	NL

Note: The solubility limit is defined as the groundwater concentration at which the water cannot dissolve any more of an individual chemical based on a petroleum mixture. The soil vapour which is in equilibrium with the groundwater will be at its maximum. If the derived groundwater HSL exceeds the water solubility limit, a soil-vapour source concentration for a petroleum mixture could not exceed a level that would result in the maximum allowable vapour risk for a given scenario. For these scenarios no HSL is presented for these chemicals. These are denoted as not limiting 'NL'.

## 8. Field Work Results

### 8.1 Soil Observations and Field Results

Borehole logs are provided in Appendix B and should be referred to for detailed descriptions of soil and rock.

At Borehole 101, brown silty sand fill and red-brown sandstone boulder fill was underlain by concrete at depth of 0.55 m. The borehole was discontinued in this concrete at a depth of 0.78 m. The borehole was discontinued and not relocated nearby as the concrete may be associated with an unknown service or buried structure.

Borehole 102 was drilled through a surface layer of asphaltic concrete (0.05 m thick) then brown silty sand fill, grey sand fill and grey clayey sand fill to a depth of 1 m. Concrete fragments were observed in the fill at a depth of 0.3 m to 0.8 m. The borehole was discontinued at a depth of 1 m due to the presence of concrete.

Borehole 103 was drilled through a surface layer of asphaltic concrete (0.05 m thick) then grey-brown sand fill and red-brown sandstone boulder fill to a depth of 1.6 m. Building rubble (brick fragments) and steel rebars were observed at depths of 0.8 m and 1.1 - 1.2 m, respectively. The borehole was discontinued at a depth of 1.7 m in concrete which was encountered from a depth of 1.6 m. Two other attempts were made to drill through the fill, but these were unsuccessful due to the presence of buried obstructions at approximately 0.7 m depth.

Borehole 104 was drilled through a surface layer of concrete (0.1 m thick) which was underlain by fill to a depth of 4.5 m. Observed fill materials included brown sand, grey and brown clayey sand, black and grey coal wash gravel, grey and brown gravel and grey-brown sand. Coal wash was observed at depths of 1.5 - 1.6 m and 2.5 - 3.8 m. Building rubble (such as fragments of terracotta, brick, glass, ceramics and concrete) were observed at depths of 0.4 - 1.0 m and 2.5 - 4.5 m. Fill was underlain by alluvial grey silty clay and silty sand to a depth of 6.8 m, then residual grey mottled yellow-brown sandy clay and grey clay to a depth of 11.8 m. Yellow-brown sandstone was encountered from a depth of 11.8 m to 12.3 m. The borehole was discontinued in sandstone at a depth of 12.3 m.

Borehole 105 was drilled through a surface layer of concrete (0.23 m thick) which was underlain by fill to a depth of 3.5 m. Fill comprised yellow-brown, brown and grey sand and sandstone boulders / cobbles. Trace building rubble (fragments of brick, terracotta and ceramics) was observed at a depth of 1.4 m. Fill was underlain by alluvial grey-brown sand clay to a depth of 4 m, then residual yellow brown sandy clay to a depth of 4.5 m. Yellow- brown, grey and red-brown sandstone was encountered from a depth of 4.5 m to 10.5 m with siltstone beds at depths of 9.33 - 9.45 m and 9.55 - 9.61 m. The borehole was discontinued in sandstone at a depth of 10.5 m.

Borehole 106 was drilled through a surface layer of brick pavers (0.1 m thick) which was underlain by brown-grey sand fill to a depth of 1.5 m. This fill was underlain by residual yellow-brown and grey-yellow sandy clay to a depth of 3.5 m. Yellow, brown, yellow-brown and grey sandstone was encountered from a depth of 3.5 m to 10.06 m. The borehole was discontinued in sandstone at a depth of 10.06 m.

Borehole 107 was drilled through a concrete slab which was suspended over an apparent old storage space. The floor of the storage space was approximately 3.1 m below the ground surface. No soil samples were retrieved due to the significant void space.

Boreholes 108 and 109 were drilled through the concrete slab in the basement of the Harwood Building which was 0.1 to 0.15 m thick. The concrete slab was underlain by a thin layer of grey gravel fill (0.1 m to 0.15 m thick). Each borehole was discontinued on the underlying sandstone which may be sandstone boulders in fill.

Borehole 201 was drilled through brick pavers (0.05 m thick) which were underlain by a thin layer of yellow-brown sand fill (0.11 m thick) and then concrete (0.09 m thick). The concrete was underlain by brown sand fill which was observed to have a trace of coal at a depth of 0.5 m and tile at a depth of 1 m. This fill was underlain by red-brown sandy clay (possibly fill) at a depth of 1.3 - 1.5 m. Yellow-brown, grey, orange-brown and red-brown sandstone was encountered between depths of 1.5 m and 10 m. The borehole was discontinued in sandstone at a depth of 10 m.

Borehole 202 was drilled through a concrete slab (0.1 m thick) which was underlain by fill to a depth of 3.1 m. Observed fill materials included brown and grey clayey sand, grey gravelly sand and grey-brown silty sand. A hydrocarbon odour was associated with the fill at a depth of 1.2 - 2.0 m. Terracotta and ceramic fragments were observed in the fill at a depth of 2.3 m. Fill was underlain by (probably) alluvial (possibly residual) grey mottled yellow-brown silty clay to a depth of 5.7 m, then residual grey sandy clay to a depth of 5.7 m. Grey and yellow-brown sandstone was encountered from between depths of 5.7 m to 10 m. The borehole was discontinued in sandstone at a depth of 10 m.

Although asbestos containing materials were not observed in the fill, building rubble such as concrete, brick, tile, terracotta and ceramics (mentioned above) were identified in fill and asbestos contamination can be associated with building rubble in fill.

PID results were either less than or equal to 1 ppm indicating a low potential for the presence of volatile organic compounds.

## 8.2 Groundwater Observations and Field Results

Free groundwater was observed at a depth of 4 m whilst drilling at Borehole 104. No free groundwater was observed at the other boreholes whilst drilling. It is noted that the method of rock coring introduces tap water to the borehole which prevents making groundwater observations.

Well development and sampling field data is shown on the groundwater field sheets in Appendix C.

Approximately 100 L of water was removed from Borehole 104 for its development. For the development of Borehole 105, the well was dry after the removal of approximately 50 L of water. LNAPL was not identified from use of the dip-meter prior to development of each well.

Table 11 shows the measured groundwater levels at each monitoring well immediately prior to sampling. It is noted that the observed free groundwater whilst drilling at a depth of 4 m at Borehole 104 is considered likely to have been perched groundwater (i.e., groundwater in the sand fill, perched on the silty clay). The low groundwater depth at Borehole 104, as shown in Table 11, may reflect the collection of perched groundwater seeping into the well. As only two groundwater wells were installed, the direction of groundwater flow could not be determined from the groundwater measurements.

**Table 11: Groundwater Levels Prior to Sampling**

Borehole	Ground Surface Level (m AHD)	Groundwater Level (15 November 2019)	
		Depth (m bgl)	Water Level (m AHD)
104	5.8	6.42	-0.6
105	6.2	3.79	2.4

Notes: m bgl metres below ground level  
 m AHD metres Australian Height Datum



## 9. Laboratory Analytical Rationale and Results

### 9.1 Analytical Rationale

Soil samples were selected for analysis for potential contaminants based on field observations. The majority of selected samples for analysis were from fill as fill was considered to be a significant potential source of contamination at the site. Consequentially, multiple samples from boreholes with the greater fill depths were subject to analysis.

One soil sample from each borehole (except Borehole 109 where no soil samples were retrieved) was analysed for eight priority metals, TRH, BTEX, PAH, OCP, OPP, PCB, total phenols and asbestos to obtain data from each sample point (i.e., to provide site coverage). Four of these samples were analysed for VOC. Additional samples were analysed for a shorter suite of contaminants (typically eight priority metals, TRH, BTEX, PAH and asbestos). In particular:

- The sample from Borehole 102, depth 0.5 m, was analysed for asbestos given that concrete fragments were observed in the fill at this depth (and asbestos contamination may be associated with building rubble in fill);
- The sample from Borehole 103, depth 1 m, was analysed for asbestos given that building rubble (brick fragments) was observed in the fill at this depth;
- The samples from Borehole 104 at depths 0.5 m, 3 m and 4 m were analysed for asbestos given that building rubble (such as terracotta, brick, glass, ceramics and concrete) was observed in the fill at these depths;
- The samples from Borehole 104 at depths 1.5 m and 3 m were analysed for PAH given that coal wash was observed at these depths;
- Three samples of fill from Borehole 105 were analysed for asbestos given that building rubble (brick, terracotta and ceramics) were identified in the fill at this location;
- The sample from Borehole 201, depth 0.9 - 1.0 m, was analysed for PAH given the presence of coal in the fill at this location. This sample was analysed for asbestos given that tile was identified in the fill;
- The sample from Borehole 202, depth 1.2 - 13 m, was analysed for TRH, BTEX, PAH and VOC given that a hydrocarbon odour was associated with the fill at this depth; and
- The sample from Borehole 202, depth 2.0 - 1.2 m, was analysed for asbestos given the presence of terracotta and ceramic fragments at this depth.

Following review of initial laboratory results, the soil sample from Borehole 102, depth 0.5 m, was analysed for TRH C<sub>10</sub>-C<sub>40</sub> with silica gel clean-up given that a somewhat elevated concentration of TRH C<sub>10</sub>-C<sub>40</sub> was recorded in this sample.

Three fill samples from different sample locations (Borehole 102, depth 0.5 m; Borehole 104, depth 1.5 m; and Borehole 106, depth 1.0 m) were analysed for the soil parameters: pH and CEC.

A groundwater sample from each groundwater monitoring well was subject to analysis for a suite of potential contaminants including metals, TRH, BTEX, PAH, OCP, OPP, PCB, VOC, ammonia and phenols. Samples were also analysed for hardness.

## 9.2 Laboratory Results

Laboratory results are presented in the laboratory certificates in Appendix E.

Table 12 summarises laboratory results for contaminants in soil compared to the site assessment criteria for soil.

Table 13 summarises the laboratory results for contaminants in groundwater compared to the site assessment criteria.

[illegible]

Preliminary Site Investigation, Powerhouse Ultimo Renewal,  
 Powerhouse Ultimo, 500 Harris Street, Ultimo

86874.02 R.001 Rev0  
 May 2022

**Table 13: Summary of Results of Groundwater Analysis (All results in µg/L)**

[illegible]

Notes:	
BD1/20191115	is blind replicate of the sample from 105 <b>BOLD</b> Exceeds DGV
PQL	Practical Quantitation Limit
-	not defined/ not analysed/ not applicable

**Table 13 (continued): Summary of Results of Groundwater Analysis (All results in µg/L)**

Sample Identification (Borehole or Replicate)		Organochlorine Pesticides											Organophosphorus Pesticides												Polychlorinated Biphenyls			Phenols	Non-metallic Inorganics	
		Aldrin	Dieldrin	gamma-Chlordane	alpha-Chlordane	pp-DDT	Endosulfan I	Endosulfan II	Endrin	Heptachlor	Heptachlor Epoxide	Methoxychlor	Other OCP	Azinphos-methyl	Bromophos-ethyl	Chlorpyrifos	Diazinon	Dichlorovos	Dimethoate	Ethion	Fenitrothion	Malathion	Parathion	Methyl Parathion	Other OPP	Aroclor 1242	Aroclor 1254	Other PCB	Total Phenols	Ammonia (total) as N
MW104		<0.01	<0.01	<0.01	<0.01	<0.006	<0.01	<0.01	<0.01	<0.01	<0.01	<PQL	<0.02	<0.2	<0.01	<0.01	<0.2	<0.15	<0.2	<0.2	<0.05	<0.01	<0.2	<PQL	<0.1	<0.1	<PQL	<50	8300	
MW105		<0.01	<0.01	<0.01	<0.01	<0.006	<0.01	<0.01	<0.01	<0.01	<0.01	<PQL	<0.02	<0.2	<0.01	<0.01	<0.2	<0.15	<0.2	<0.2	<0.05	<0.01	<0.2	<PQL	<0.1	<0.1	<PQL	<50	220	
BD1/20191115		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
Assessment Criteria																														
Marine Water DGV		0.003	0.01	0.001		0.0004	0.005		0.004	0.0004	-	0.004	-	0.01	-	0.01	0.01	-	0.15	-	0.001	0.05	0.004	-	-	0.3	0.01	-	-	5070 at pH 6.6
Guidelines for Recreational Water	Health	3		20		90	200		-	3		3000	-	300	100	100	40	50	70	40	70	700	200	7	-	-	-	-	-	
	Aesthetic	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	500	
HSL B for Vapour Intrusion (depth 2 m to <4 m)		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
HSL D for Vapour Intrusion (depth 2 m to <4 m)		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	

Notes:

BD1/20191115 is blind replicate of the sample from 105

**BOLD** Exceeds DGV and guideline for recreational water

PQL Practical Quantitation Limit

- not defined/ not analysed/ not applicable



### 9.3 Soil Contaminants

Concentrations of arsenic, chromium, nickel and zinc were within the respective HIL B, HIL D, urban residential EIL, commercial/industrial EIL in analysed soil samples. Concentrations of cadmium and mercury were within HIL B and HIL D.

Concentrations of copper in soil were within HIL B and HIL D and the commercial/industrial EIL. Concentrations of copper were within the urban residential EIL (240 mg/kg) except for the fill sample from Borehole 104, depth 3 m (280 mg/kg). Statistical analysis on fill samples using ProUCL 5.1 indicates that this exceedance of the urban residential EIL is not significant given that the 95% Adjusted Gamma Upper Confidence Limit (UCL) of 104 mg/kg is less than the urban residential EIL and the standard deviation of 68.96 mg/kg is less than half of the urban residential EIL. The input values and output values are shown in Appendix F. Note that, to be conservative, the higher recorded concentration for replicate pairs was used as input values.

Concentrations of lead were within HIL D, urban residential EIL and commercial/industrial EIL. Concentrations of lead were within HIL B (1200 mg/kg) except for the replicate fill sample (BD26112019) from Borehole 202, depth 2 - 2.1 m (1300 mg/kg). Statistical analysis on fill samples using ProUCL 5.1 indicates that this exceedance of the urban residential EIL is not significant given that the 95% Chebyshev Upper Confidence Limit (UCL) of 607.6 mg/kg is less than the urban residential EIL and the standard deviation of 378.9 mg/kg is less than half of the urban residential EIL. The input values and output values are shown in Appendix F. Note that, to be conservative, the higher recorded concentration for replicate pairs was used as input values. It is also noted that the primary sample from Borehole 202, depth 2 - 2.1 m, had a concentration of 320 mg/kg which is well within HIL B.

Concentrations of benzo(a)pyrene TEQ were within HIL B (4 mg/kg) except for samples from Borehole 105, depth 2 m (11 mg/kg); Borehole 105, depth 3 m (7.9 mg/kg); Borehole 202, depth 1.2 - 1.3 m (420 mg/kg); and Borehole 202, depth 2 - 2.1 m (31 mg/kg). Concentrations of benzo(a)pyrene TEQ were within HIL D (40 mg/kg) except for sample from Borehole 202, depth 1.2 - 1.3 m (420 mg/kg). Concentrations of total PAH were within HIL B (400 mg/kg) and HIL D (4000 mg/kg) except for the sample from Borehole 202, depth 1.2 - 1.3 m (5700 mg/kg).

Concentrations of benzo(a)pyrene exceeded the urban residential ESL (0.7 mg/kg) in eight of nineteen analysed samples. Concentrations of benzo(a)pyrene exceeded the commercial/industrial ESL (1.4 mg/kg) in five of these samples. CRC CARE 2017, *Technical Report No. 39, Risk-based management and remediation guidance for benzo(a)pyrene*, 2017 provides a high reliability ecological guidelines for fresh benzo(a)pyrene of 33 mg/kg for public open space and 172 mg/kg for commercial/industrial sites (and the bioavailability and bio-accessibility of aged benzo(a)pyrene tends to be less than that of fresh benzo(a)pyrene which means that the ecological guideline is conservative for aged benzo(a)pyrene). (It is considered that the benzo(a)pyrene in fill is aged given the site history). The recorded concentrations of benzo(a)pyrene for all analysed samples were within the CRC CARE ecological guidelines for residential sites and commercial / industrial sites except for the sample from Borehole 202, depth 1.2 - 1.3 m (290 mg/kg). In addition to CRC Care remediation guidance for benzo(a)pyrene, the Canadian Council of Ministers of the Environment, *Canadian Soil Quality Guidelines for the Protection of Environmental and Human Health, Polycyclic Aromatic Hydrocarbons*, 2010 provides Environmental Soil Quality Guidelines (SQGE) of 20 mg/kg for residential sites and 72 mg/kg for commercial and industrial sites. The recorded concentrations of benzo(a)pyrene for all analysed samples are within the commercial / industrial SQGE except for the sample from Borehole 202,

depth 1.2 - 1.3 m. Concentrations were within the residential SQGE except for the samples from Borehole 202, depth 1.2 - 1.3 m and Borehole 202, depth 2.0 - 2.1 mg/kg (21 mg/kg).

Concentrations of naphthalene were within the urban residential EIL, commercial/industrial EIL and HSL D for vapour intrusion. Concentrations of naphthalene were within HSL B for vapour intrusion (3 mg/kg) except for samples from Borehole 105, depth 2 m (3.9 mg/kg); Borehole 105, depth 3 m (4 mg/kg); Borehole 202, depth 1.2 - 1.3 m (100 mg/kg and 73 mg/kg); Borehole 202, depth 2 - 2.1 m (8 mg/kg and 6 mg/kg for the primary samples and 26 mg/kg for the replicate sample). It is noted that samples from depths of 2 m and 3 m from Borehole 105 also had recorded naphthalene concentrations of 2 mg/kg and 1.9 mg/kg, respectively.

Concentrations of TRH C<sub>6</sub>-C<sub>10</sub> and BTEX were less than PQL in analysed samples, however, it is noted that the PQL for TRH C<sub>6</sub>-C<sub>10</sub> for the replicate sample BD26112019 was significantly raised due to matrix interference. The PQL of 200 mg/kg for TRH C<sub>6</sub>-C<sub>10</sub> in sample BD26112019 is greater than the HSL B for vapour intrusion and residential ESL. Apart from this, concentrations of TRH C<sub>6</sub>-C<sub>10</sub> and BTEX were within the respective HSL B for vapour intrusion, HSL D for vapour intrusion, residential ESL, commercial/industrial ESL, residential management limit and commercial / industrial management limit. Concentrations of VOC (in addition to BTEX) were less than the PQL in analysed samples.

Concentrations of TRH >C<sub>10</sub>-C<sub>16</sub> were less than the PQL except for samples from Borehole 102, depth 0.5 m (63 mg/kg); Borehole 202, depth 1.2 - 1.3 m (1100 mg/kg); and Borehole 202, depth 2.0 - 2.1 m (82 mg/kg). The concentration of TRH >C<sub>10</sub>-C<sub>16</sub> with silica gel clean-up in the sample from Borehole 102, depth 0.5 m, was less than the PQL. It is noted that the PQL for TRH >C<sub>10</sub>-C<sub>16</sub> for the replicate sample BD26112019 was significantly raised due to matrix interference. The PQL of 500 mg/kg for TRH >C<sub>10</sub>-C<sub>16</sub> in sample BD26112019 is greater than the HSL B for vapour intrusion, residential ESL, and commercial/industrial ESL. Concentrations of TRH >C<sub>10</sub>-C<sub>16</sub> (less naphthalene) were within the HSL D for vapour intrusion. Concentrations of TRH >C<sub>10</sub>-C<sub>16</sub> (less naphthalene) were within HSL B for vapour intrusion (110 mg/kg) except for the sample from Borehole 202, depth 1.2 - 1.3 m (1000 mg/kg). Concentrations of TRH >C<sub>10</sub>-C<sub>16</sub> were within residential ESL (120 mg/kg), commercial/industrial ESL (170 mg/kg), residential management limit (1000 mg/kg) and commercial / industrial management limit (1000 mg/kg) except for the sample from Borehole 202, depth 1.2 - 1.3 m.

Concentrations of TRH >C<sub>16</sub>-C<sub>34</sub> were within the urban residential ESL (300 mg/kg) except for the samples from Borehole 102, depth 0.5 m (640 mg/kg); Borehole 105, depth 2 m (390 mg/kg); Borehole 105, depth 3 m (340 mg/kg); Borehole 202, depth 1.2 - 1.3 m (8500 mg/kg); and Borehole 202, depth 2.0 - 2.1 m (780 mg/kg for the primary sample and 2300 mg/kg for the replicate sample). The concentration of TRH >C<sub>16</sub>-C<sub>34</sub> with silica gel clean-up for the sample from Borehole 102, depth 0.5 m (440 mg/kg) exceeded the urban residential ESL. Concentrations of TRH >C<sub>16</sub>-C<sub>34</sub> were within the industrial /commercial ESL (1700 mg/kg) except for the sample from Borehole 202, depth 1.2 - 1.3 m, and the replicate sample from Borehole 202, depth 2.0 - 2.1 m. Concentrations of TRH >C<sub>16</sub>-C<sub>34</sub> were within the urban residential management limit (2500 mg/kg) and commercial / industrial management limit (3500 mg/kg) except for the sample from Borehole 202, depth 1.2 - 1.3 m.

Concentrations of TRH >C<sub>34</sub>-C<sub>40</sub> were within the urban residential ESL, industrial / commercial ESL, urban residential management limit and commercial / industrial management limit.

Concentrations of aldrin and dieldrin were less than the PQL except for the samples from Borehole 108, depth 0.1 - 0.2 m, and Borehole 109, depth 0.15 - 0.2 m. The concentrations of aldrin (7.9 mg/kg) and dieldrin (15 mg/kg) in the sample from Borehole 108 were above HIL B (10 mg/kg) but below HIL D (45 mg/kg). The concentrations of aldrin (2.2 mg/kg) and dieldrin (4.1 mg/kg) in the sample from Borehole 109 were within HIL B and HIL D.

Concentrations of OCP other than aldrin and dieldrin were less than the PQL in analysed samples. Hence, concentrations of DDT were within the urban residential EIL and commercial / industrial EIL; and concentrations of DDT, DDE, DDD, chlordane, endosulfan, endrin, heptachlor, HCB and methoxychlor were within HIL B and HIL D.

Concentrations of PCB, OPP and total phenols were less than the PQL, and, hence, concentrations of total PCB, phenols and chlorpyrifos were within the respective HIL B and HIL D.

Asbestos was not detected at the laboratory limits of reporting in analysed samples.

#### **9.4 Groundwater Contaminants**

Concentrations of arsenic were below the PQL except in the primary sample from Borehole 105. The concentration (1 µg/L) in this sample was well within the guideline for recreational water. There is no DGV for marine water for arsenic. It is noted, however, that the recorded concentration for the primary sample from Borehole 105 is within the freshwater DGV for a slightly to moderately disturbed ecosystem which are 24 for arsenic III and 13 for arsenic V.

Concentrations of cadmium were below the PQL in all analysed samples.

Concentrations of chromium were below the PQL except in the primary sample from Borehole 105. The concentration (2 µg/L) in this sample was within the DGV and guideline for recreational water.

The copper concentration (1 µg/L) for the sample from Borehole 104 was within the DGV and guideline for recreational water. The copper concentrations in the primary sample (4 µg/L) and replicate sample (2 µg/L) from Borehole 105 were above the DGV (1.3 µg/L) but well within the guidelines for recreational water use. The recorded copper concentrations are considered to be consistent with typical background ranges for groundwater in Sydney.

Concentrations of lead were below the PQL except in the primary sample from Borehole 105. The concentration (5 µg/L) in this sample was marginally above the DGV (4.4 µg/L) but well within the guideline for recreational water. It is noted that the replicate sample from BH105 had a concentration (<1 µg/L) within the DGV. The recorded lead concentrations are considered to be consistent with typical background ranges for groundwater in Sydney.

Concentrations of mercury were below the PQL except in the primary sample from Borehole 105. The concentration (0.06 µg/L) in this sample was within the DGV and guideline for recreational water.

The nickel concentration in the sample from Borehole 104 was less than the PQL. The nickel concentrations in the primary sample (12 µg/L) and the replicate sample (11 µg/L) from BH105 were above the DGV (7 µg/L) but well within the guideline for recreational water. The recorded nickel concentrations are considered to be consistent with typical background ranges for groundwater in Sydney.

The recorded zinc concentrations (11 µg/L in the sample from Borehole 104, and 11 µg/L and 18 µg/L in the samples from Borehole 105) were above the DGV (8 µg/L) but well within the guideline for recreational water. The recorded zinc concentrations are considered to be consistent with typical background ranges for groundwater in Sydney.

Concentrations of TRH, BTEX, VOC, OCP, OPP, PCB and total phenols were less than the PQL in all analysed samples.

The concentration of total ammonia in the sample from Borehole 105 (220 µg/L as N) was well within the DGV and guideline for recreational use. The concentration of total ammonia (8300 µg/L as N) in the sample from Borehole 104 was above the DGV (5070 µg/L as N at pH 6.6) and the guideline for recreational water (500 µg/L).

## 10. Discussion

Given the presence of a hydrocarbon odour and the observed absence of coal or a similar substance, the primary source of the high concentrations of PAH and TRH in fill at Borehole 202 may be coal tar, although this is not confirmed. Coal tar is pre-classified as hazardous waste according to NSW EPA, *Waste Classification Guidelines*, 2014. It is noted that concentrations of PAH and TRH were below the PQL in samples from the two groundwater monitoring wells (Boreholes 104 and 105) which are inferred to be down-gradient of Borehole 202 which indicates that the PAH and TRH contaminated fill at Borehole 202 has not impacted groundwater.

Apart from the potential coal tar at Borehole 202 discussed above, elevated concentrations of metals (in particular, copper and lead), PAH and TRH in fill may be attributed to the nature of the fill which was observed to contain various soil types and commonly contained anthropogenic materials. Coal and coal wash (which has a component of coal) was observed in fill and may be a source of PAH which was recorded at concentrations above the PQL in the majority of fill samples. The results for groundwater samples indicate that the fill with elevated concentrations of metals, PAH and TRH has not impacted groundwater at the southern part of the site.

The two fill samples with aldrin and dieldrin concentrations above the PQL (from Boreholes 108 and 109) were from directly beneath the concrete slab in the basement of the Harwood Building which indicates that these pesticides have been applied beneath the slab for the purpose of pest control. These chemicals were not identified in the groundwater at Borehole 105 which is inferred to be down-gradient of the Harwood Building, which indicates that groundwater has not been impacted from the use of these pesticides.

The source of ammonia in groundwater at Borehole 105 is not known and could potentially be from off-site. It is noted that the samples were not filtered and, thus, the elevated concentration in the sample from BH104 could be from particulate matter (soil) rather than dissolved ammonia.

Overall, results indicate that remediation or management of contamination will be required for site development although the extent of remediation / management will likely depend on the layout of future site uses and the presence or absence of accessible soil (for landscaping).

Prior to establishing plans for remediation / management of contamination, a detailed site investigation (including soil and groundwater sampling) should be undertaken.

## 11. Conclusion

Limited intrusive investigations have revealed the presence of fill up to 5 m in depth. Fill comprised various soil types and commonly contained anthropogenic materials. Elevated concentrations of PAH, TRH, metals and OCP have been identified in fill. In addition, an elevated concentration of ammonia in groundwater was encountered at one sample location.

Overall, results indicate that remediation or management of contamination will be required for site development although the extent of remediation / management will likely depend on the layout of future site uses as well as the results of additional investigation.

The following is recommended for the Stage 2 detailed DA:

- A detailed site investigation (including soil and groundwater sampling).

The results of the detailed site investigation would be used to determine remediation or management requirements to address site contamination. On this basis, it is considered that the site can be made suitable for the proposed Powerhouse Ultimo Renewal development.

## 12. Limitations

Douglas Partners (DP) has prepared this report for the Powerhouse Ultimo Renewal project at 500 Harris Street, Ultimo in general accordance with DP's proposals dated 23 February 2022 and acceptance received from Department of Premier and Cabinet. This report is provided for the exclusive use of Department of Premier and Cabinet for this project only and for the purposes as described in the report. It should not be used by or relied upon for other projects or purposes on the same or other site or by a third party. Any party so relying upon this report beyond its exclusive use and purpose as stated above, and without the express written consent of DP, does so entirely at its own risk and without recourse to DP for any loss or damage. In preparing this report DP has necessarily relied upon information provided by the client and/or their agents.

The results provided in the report are indicative of the sub-surface conditions on the site only at the specific sampling and/or testing locations, and then only to the depths investigated and at the time the work was carried out. Sub-surface conditions can change abruptly due to variable geological processes and also as a result of human influences. Such changes may occur after DP's field testing has been completed.



DP's advice is based upon the conditions encountered during this investigation. The accuracy of the advice provided by DP in this report may be affected by undetected variations in ground conditions across the site between and beyond the sampling and/or testing locations. The advice may also be limited by budget constraints imposed by others or by site accessibility.

The assessment of atypical safety hazards arising from this advice is restricted to the (environmental) components set out in this report and based on known project conditions and stated design advice and assumptions. While some recommendations for safe controls may be provided, detailed 'safety in design' assessment is outside the current scope of this report and requires additional project data and assessment.

This report must be read in conjunction with all of the attached and should be kept in its entirety without separation of individual pages or sections. DP cannot be held responsible for interpretations or conclusions made by others unless they are supported by an expressed statement, interpretation, outcome or conclusion stated in this report.

This report, or sections from this report, should not be used as part of a specification for a project, without review and agreement by DP. This is because this report has been written as advice and opinion rather than instructions for construction.

Asbestos has not been detected by observation or by laboratory analysis, either on the surface of the site, or in filling materials at the test locations sampled and analysed. Building demolition materials, such as concrete, brick, terracotta and ceramics, were, however, located in previous below-ground fill, and these are considered as indicative of the possible presence of hazardous building materials (HBM), including asbestos.

Although the sampling plan adopted for this investigation is considered appropriate to achieve the stated project objectives, there are necessarily parts of the site that have not been sampled and analysed. This is either due to undetected variations in ground conditions or to budget constraints (as discussed above), or to parts of the site being inaccessible and not available for inspection/sampling. It is therefore considered possible that HBM, including asbestos, may be present in unobserved or untested parts of the site, between and beyond sampling locations, and hence no warranty can be given that asbestos is not present.

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**Douglas Partners Pty Ltd**

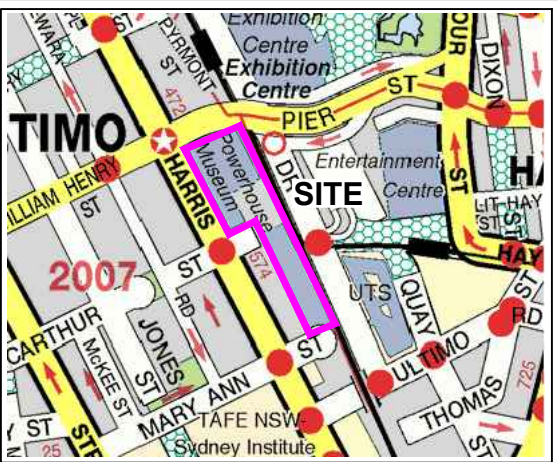
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## Appendix A

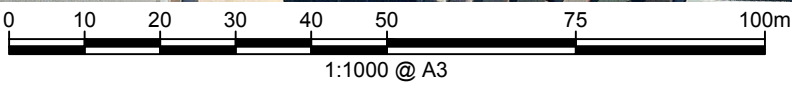
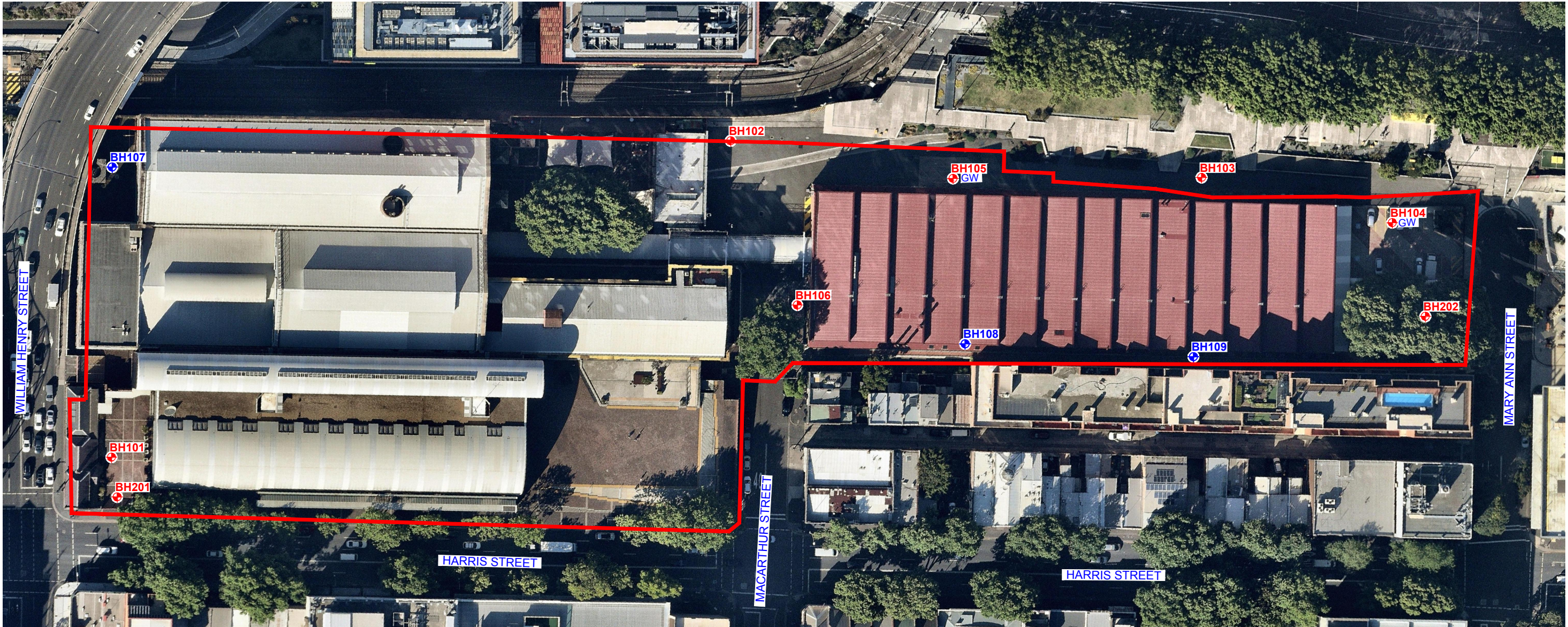
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Drawing



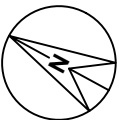


Locality Plan



NOTE:  
1: Base image from Nearmap.com  
(Dated 22.10.2019)  
2: Test locations are approximate only and are  
shown with reference to existing features.

**LEGEND**  
• Borehole location  
• Hand auger bore  
GW Groundwater well





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## Appendix B

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Borehole Logs

About this Report

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 15.2 AHD  
**EASTING:** 333431  
**NORTHING:** 6249918  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH101  
**PROJECT No:** 86874.00  
**DATE:** 22/10/2019  
**SHEET 1 OF 1**

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing				Water	Well Construction Details	
				Type	Depth	Sample	Results & Comments			
15		FILL/Silty SAND: fine, brown-dark brown, trace fine mixed origin gravel, moist		A/E	0.1		PID<1			
0.4		FILL/SANDSTONE BOULDER: fine to medium, red-brown, dry		A/E*	0.5		PID<1			
0.55		CONCRETE: pale grey, aggregate diameter from 10-25mm, 8mm steel reinforcement								
0.78		Bore discontinued at 0.78m No further drilling after encountering concrete								
1										
14										
2										
13										
3										
12										
4										
11										

**RIG:** Bobcat **DRILLER:** JE **LOGGED:** RK **CASING:** HW to 0.5m, HQ to 0.5m  
**TYPE OF BORING:** Solid Flight Auger (TC-bit) to 0.55m, NMLC to 0.78m  
**WATER OBSERVATIONS:** No free groundwater observed during augering  
**REMARKS:** \*replicate BD1\_20191022

SAMPLING & IN SITU TESTING LEGEND					
A	Auger sample	G	Gas sample	PID	Photo ionisation detector (ppm)
B	Bulk sample	P	Piston sample	PL(A)	Point load axial test Is(50) (MPa)
BLK	Block sample	U	Tube sample (x mm dia.)	PL(D)	Point load diametral test Is(50) (MPa)
C	Core drilling	W	Water sample	pp	Pocket penetrometer (kPa)
D	Disturbed sample	W	Water seep	S	Standard penetration test
E	Environmental sample	W	Water level	V	Shear vane (kPa)



BORE: 101

PROJECT: ULTIMO

OCTOBER 2019



Project No: 86874.00  
BH ID: BH 101  
Depth: 0.55 - 0.78 m  
Core Box No.: 1



Powerhouse Museum  
86874.00 BH101 Start at 0.55m 22/10/19



End of hole at  
0.78m

0.55 - 0.78m

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 6.1 AHD  
**EASTING:** 333572  
**NORTHING:** 6249819  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH102  
**PROJECT No:** 86874.00  
**DATE:** 22/10/2019  
**SHEET 1 OF 1**

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing				Water	Well Construction Details	
				Type	Depth	Sample	Results & Comments			
6	0.05	ASPHALTIC CONCRETE: dark grey-black		A/E	0.1		PID<1			
		FILL/Silty SAND: fine, dark brown-brown with fine to coarse mixed origin gravel, dry-moist								
	0.3	FILL/SAND: fine to coarse, dark grey with concrete fragments (20-70mm diameter) and fine to coarse mixed origin gravel, moist		A/E	0.5		PID<1			
	0.8	FILL/Clayey SAND: fine to coarse, low plasticity, dark grey with fine to coarse mixed origin gravel, moist								
1	1.0	Bore discontinued at 1.0m No further drilling after encountering concrete		A/E S	1.0		PID<1 1/20mm refusal HB			
6										
2										
3										
4										

**RIG:** Bobcat

**DRILLER:** JE

**LOGGED:** RK

**CASING:** none

**TYPE OF BORING:** Solid Flight Auger (TC-bit) to 1.00m

**WATER OBSERVATIONS:** No free groundwater observed during augering

**REMARKS:** HB = SPT hammer bouncing

## SAMPLING & IN SITU TESTING LEGEND

A	Auger sample	G	Gas sample	PID	Photo ionisation detector (ppm)
B	Bulk sample	P	Piston sample	PL(A)	Point load axial test Is(50) (MPa)
BLK	Block sample	U	Tube sample (x mm dia.)	PL(D)	Point load diametral test Is(50) (MPa)
C	Core drilling	W	Water sample	pp	Pocket penetrometer (kPa)
D	Disturbed sample	>	Water seep	S	Standard penetration test
E	Environmental sample	≡	Water level	V	Shear vane (kPa)

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 6.3 AHD  
**EASTING:** 333618  
**NORTHING:** 6249712  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH103  
**PROJECT No:** 86874.00  
**DATE:** 22/10/2019  
**SHEET 1 OF 1**

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing				Water	Well Construction Details	
				Type	Depth	Sample	Results & Comments			
6	0.05	ASPHALTIC CONCRETE: black-dark grey		A/E	0.1		PID<1		1	
		FILL/SAND: fine to coarse, dark grey-dark brown with fine to coarse mixed origin gravel, dry-moist								
	0.4	FILL/SANDSTONE BOULDERS: fine to medium, red-brown, with fine to coarse mixed origin gravel, moist		A/E	0.5		PID<1			
		0.80m: with building rubble (brick fragments)		A	0.8					
		1.10-1.20m: steel rebars (x2) in sidewall 1.20-1.30m: dimensioned sandstone block		A/E* S	1.0		PID<1 3/20mm refusal HB			
	1.6		E	1.5		PID<1				
	1.7	CONCRETE: pale grey								
	1.7	Bore discontinued at 1.7m No further drilling after encountering concrete								
	2								2	
	3								3	
	4								4	
	5									
	6									
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	98									
	99									
	100									

**RIG:** Bobcat

**DRILLER: JE**

**LOGGED: RK**

**CASING:** none

**TYPE OF BORING:** Solid Flight Auger (TC-bit) to 1.70m

**WATER OBSERVATIONS:** No free groundwater observed during augering

**REMARKS:** \*replicate BD2\_20191022, borehole location moved three times, about 0.5m apart, first two attempts refused on steel at a depth of 0.8m, HB = SPT hammer bouncing

SAMPLING & IN SITU TESTING LEGEND			
A	Auger sample	G	Gas sample
B	Bulk sample	P	Piston sample
BLK	Block sample	U	Tube sample (x mm dia.)
C	Core drilling	W	Water sample
D	Disturbed sample	W	Water seep
E	Environmental sample	W	Water level
		PID	Photo ionisation detector (ppm)
		PL(A)	Point load axial test Is(50) (MPa)
		PL(D)	Point load diametral test Is(50) (MPa)
		pp	Pocket penetrometer (kPa)
		S	Standard penetration test
		V	Shear vane (kPa)



# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 5.8 AHD  
**EASTING:** 333630  
**NORTHING:** 6249665  
**DIP/AZIMUTH:** 90°/-

**BORE No:** BH104  
**PROJECT No:** 86874.00  
**DATE:** 23/10/2019  
**SHEET 1 OF 2**

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing				Water	Well Construction Details	
				Type	Depth	Sample	Results & Comments			
	0.1	CONCRETE: pale grey, aggregate diameter from 15-50mm		A/E	0.1		PID<1		Flush Gatic Cover	
	0.4	FILL/SAND: medium to coarse, dark brown-brown, with fine to medium mixed origin gravel, moist-wet (wet from diatube)		A/E*	0.5		PID<1		Gravel	
	1.0	FILL/Clayey SAND: fine to medium, low plasticity, pale grey, trace building rubble (fragments of terracotta, brick and glass), moist		A/E S	1.0		PID<1 4,5,7 N = 12		Blank	
	1.5	FILL/CLAY: low plasticity, brown-dark brown with fine to medium clayey sand and fine to coarse mixed origin gravel, w<PL		A/E	1.5		PID<1		Bentonite	
	1.6	FILL/COAL WASH: fine to coarse coal wash gravel, black-dark grey with fine to coarse sand, moist		A/E	2.0		PID<1			
	2.5	FILL/GRAVEL: fine to coarse mixed origin, dark grey and brown with igneous cobbles and fine to coarse sand, moist		E S	2.5		1,2,5 N = 7			
	3.0	FILL/SAND: fine to coarse, dark grey-dark brown, trace clay, coal wash, fine to coarse mixed origin gravel and building rubble (fragments of terracotta, brick, ceramics and concrete), moist		E	3.0		PID<1			
	3.5			E	3.5		PID<1			
	3.8	FILL/SAND: fine to medium, dark grey-dark brown, trace building rubble (fragments of terracotta and ceramics), moist		E S	4.0		PID<1 5,6,8 N = 14			
	4.0	4.00m: becoming wet		E	4.5		PID<1			
	4.5	Silty CLAY CH: high plasticity, dark grey with fine sand, w<PL, firm, alluvial		A/E	5.0		PID<1			
	5.5	Silty SAND SP: fine to medium, dark grey with clay, wet, loose, alluvial		S	5.5		2,3,6 N = 9			
	6.8	Sandy CLAY CL: low plasticity, fine, pale grey mottled yellow-brown, w<PL, very stiff, residual		S	7.0		8,11,12 N = 23		Gravel	
	8.2	CLAY CH: high plasticity, dark grey-grey with fine sand, w<PL, very stiff, residual		S	8.5		5,10,14 N = 24		Slotted Pipe	
	9.6	Sandy CLAY Cl: low plasticity, fine to medium, pale grey, w<PL, very stiff, residual		S	10.0		7,11,12			

**RIG:** Bobcat

**DRILLER:** JE

**LOGGED:** RK

**CASING:** HW to 6.0m

**TYPE OF BORING:** Diatube to 0.10m, Solid Flight Auger (TC-bit) to 6.00m, wash boring to 12.30m

**WATER OBSERVATIONS:** Free groundwater encountered at 4.00m

**REMARKS:** \*replicate BD3\_20191023, HB = SPT hammer bouncing

## SAMPLING & IN SITU TESTING LEGEND

A	Auger sample	G	Gas sample	PID	Photo ionisation detector (ppm)
B	Bulk sample	P	Piston sample	PL(A)	Point load axial test Is(50) (MPa)
BLK	Block sample	U	Tube sample (x mm dia.)	PL(D)	Point load diametral test Is(50) (MPa)
C	Core drilling	W	Water sample	pp	Pocket penetrometer (kPa)
D	Disturbed sample	>	Water seep	sp	Standard penetration test
E	Environmental sample	≡	Water level	V	Shear vane (kPa)



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# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 5.8 AHD  
**EASTING:** 333630  
**NORTHING:** 6249665  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH104  
**PROJECT No:** 86874.00  
**DATE:** 23/10/2019  
**SHEET 2 OF 2**

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing				Water	Well Construction Details	
				Type	Depth	Sample	Results & Comments			
		Sandy CLAY Cl: low plasticity, fine to medium, pale grey, w<PL, very stiff, residual ( <i>continued</i> )					N = 23			
	11									
	11.8									
	12	SANDSTONE: fine to medium, yellow-brown, inferred low-medium strength		S	12.1		7.5/50mm refusal HB		End Cap	
	12.3	Bore discontinued at 12.3m Groundwater monitoring well installed								
	13									
	14									
	15									
	16									
	17									
	18									
	19									

**RIG:** Bobcat

**DRILLER:** JE

**LOGGED:** RK

**CASING:** HW to 6.0m

**TYPE OF BORING:** Diatube to 0.10m, Solid Flight Auger (TC-bit) to 6.00m, wash boring to 12.30m

**WATER OBSERVATIONS:** Free groundwater encountered at 4.00m

**REMARKS:** \*replicate BD3\_20191023, HB = SPT hammer bouncing

## SAMPLING & IN SITU TESTING LEGEND

A	Auger sample	G	Gas sample	PID	Photo ionisation detector (ppm)
B	Bulk sample	P	Piston sample	PL(A)	Point load axial test Is(50) (MPa)
BLK	Block sample	U	Tube sample (x mm dia.)	PL(D)	Point load diametral test Is(50) (MPa)
C	Core drilling	W	Water sample	pp	Pocket penetrometer (kPa)
D	Disturbed sample	>	Water seep	S	Standard penetration test
E	Environmental sample	≡	Water level	V	Shear vane (kPa)

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 6.2 AHD  
**EASTING:** 333589  
**NORTHING:** 6249766  
**DIP/AZIMUTH:** 90°/-

**BORE No:** BH105  
**PROJECT No:** 86874.00  
**DATE:** 28/10/2019  
**SHEET 1 OF 2**

RL	Depth (m)	Description of Strata	Degree of Weathering						Graphic Log	Rock Strength					Water	Fracture Spacing (m)	Discontinuities		Sampling & In Situ Testing			
			EW	HW	MW	SW	FS	FR		Ex Low	Very Low	Low	Medium	High			Very High	Ex High	B - Bedding S - Shear	J - Joint F - Fault	Type	Core Rec. %
6	0.23	CONCRETE: pale grey, aggregate size from 15-45mm, 8mm reinforcing steel																A/E			PID<1	
		FILL/ SAND: fine to coarse, yellow brown, with fine to coarse mixed origin gravel, moist (wet at top from diatube)																A/E			PID<1	
1		0.7-.9m: sandstone boulders																S			2.4,4 N = 8	
5		0.9m: colour change to dark brown dark grey, trace fine to coarse mixed origin gravel																E			PID<1	
		1.4m: trace building rubble (fragments of brick, terracotta and ceramics)																E*			PID<1	
2		2.0m: with low plasticity clay, trace sandstone cobbles																E			PID<1	
4																		S			1.2,5 N = 7	
3																		A/E			PID<1	
3	3.5	Sandy CLAY CL: low plasticity, fine to medium, dark grey dark brown, w<PL, soft to firm, alluvial																E			PID<1	
4	4.0	Sandy CLAY CL-CI: low to medium plasticity, fine to medium, yellow brown, w<PL, stiff, residual																E			PID<1	
2																		S			1.4,11 N = 15	
4.5		SANDSTONE: medium to coarse, yellow-brown, iron stained, low strength, moderately weathered, Hawkesbury Sandstone																			PL(A) = 0.2	
5	5.0																				4.55m: J 40°, cln, pl, vr 4.8m: CORE LOSS: 200mm 5.11m: CORE LOSS: 270mm	
5.38		SANDSTONE: medium to coarse, pale grey and yellow-brown some red-brown, iron stained, indistinctly bedded, medium then high strength, slightly weathered, slightly fractured, Hawkesbury Sandstone																C	82	80	PL(A) = 0.5	
6																					5.62m: J 35°, cln, pl 5.85m: J 65°, cln, pl 5.92m: B 10°, cln, pl	
																					6.59m: B 10°, cln, pl, ro 6.84m: Cs, h 20mm 7.04m: B 5°, cln, pl, ro	
7																					PL(A) = 0.9	
																					PL(A) = 1	
8	8.0	SANDSTONE: as above																			7.75m: J 45°, cln, pl, ro	
																					8.25m: J 30°, cln, pl, ro 8.47m: B 10°, cln, pl, ro 8.64m: B h, cln, pl, ro	
8.62		SANDSTONE: medium to coarse, pale grey, indistinctly bedded, high strength, fresh, slightly fractured, Hawkesbury Sandstone																C	100	93	PL(A) = 1.4	
9																						
9.33		9.33-9.45m: siltstone bed																			9.33m: Cs h 35mm	
		9.55-9.61m: siltstone bed																			PL(A) = 0.8	

**RIG:** Bobcat  
**DRILLER:** JE  
**LOGGED:** RK  
**CASING:** HW to 4.0m, HQ to 4.5m

**TYPE OF BORING:** Diatube to 0.23m, Solid Flight Auger (TC-bit) to 4.00m, wash boring to 4.50m, NMLC coring to 10.15m

**WATER OBSERVATIONS:** No free groundwater observed during augering

**REMARKS:** \*replicate BD4\_20191028

SAMPLING & IN SITU TESTING LEGEND			
A	Auger sample	G	Gas sample
B	Bulk sample	P	Piston sample
BLK	Block sample	U	Tube sample (x mm dia.)
C	Core drilling	W	Water sample
D	Disturbed sample	>	Water seep
E	Environmental sample	≡	Water level
		PID	Photo ionisation detector (ppm)
		PL(A)	Point load axial test Is(50) (MPa)
		PL(D)	Point load diametral test Is(50) (MPa)
		pp	Pocket penetrometer (kPa)
		S	Standard penetration test
		V	Shear vane (kPa)



# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo  
**SURFACE LEVEL:** 6.2 AHD  
**EASTING:** 333589  
**NORTHING:** 6249766  
**DIP/AZIMUTH:** 90°/--  
**BORE No:** BH105  
**PROJECT No:** 86874.00  
**DATE:** 28/10/2019  
**SHEET 2 OF 2**

RL	Depth (m)	Description of Strata	Degree of Weathering					Graphic Log	Rock Strength					Water	Fracture Spacing (m)				Discontinuities		Sampling & In Situ Testing																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																	
			EW	HW	MW	SW	FS		FR	Ex Low	Very Low	Low	Medium		High	Very High	Ex High	0.01	0.05	0.10	0.50	1.00	B - Bedding S - Shear	J - Joint F - Fault	Type	Core Rec. %	RQD %	Test Results & Comments																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																										
-4	10.15	SANDSTONE: fine to medium, pale grey, indistinctly bedded, high strength with very low to low strength beds, fresh, Hawkesbury Sandstone <i>(continued)</i> Bore discontinued at 10.15m Target depth reached																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																				

**RIG:** Bobcat  
**DRILLER:** JE  
**LOGGED:** RK  
**CASING:** HW to 4.0m, HQ to 4.5m  
**TYPE OF BORING:** Diatube to 0.23m, Solid Flight Auger (TC-bit) to 4.00m, wash boring to 4.50m, NMLC coring to 10.15m  
**WATER OBSERVATIONS:** No free groundwater observed during augering  
**REMARKS:** \*replicate BD4\_20191028

SAMPLING & IN SITU TESTING LEGEND			
A	Auger sample	G	Gas sample
B	Bulk sample	P	Piston sample
BLK	Block sample	U	Tube sample (x mm dia.)
C	Core drilling	W	Water sample
D	Disturbed sample	W	Water seep
E	Environmental sample	W	Water level
		PID	Photo ionisation detector (ppm)
		PL(A)	Point load axial test Is(50) (MPa)
		PL(D)	Point load diametral test Is(50) (MPa)
		pp	Pocket penetrometer (kPa)
		S	Standard penetration test
		V	Shear vane (kPa)

BORE: 105

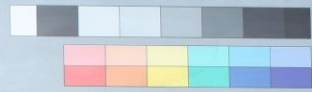
PROJECT: ULTIMO

OCTOBER 2019



**Douglas Partners**  
Geotechnics | Environment | Groundwater

Project No: 86874.00  
BH ID: BH 105  
Depth: 4.50 - 9.00 m  
Core Box No.: 1



4.50 - 9.00m

BORE: 105

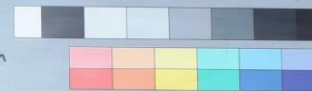
PROJECT: ULTIMO

OCTOBER 2019



**Douglas Partners**  
Geotechnics | Environment | Groundwater

Project No: 86874.00  
BH ID: BH 105  
Depth: 9.00 - 10.15 m  
Core Box No.: 2



9.00 - 10.15m

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 6.2 AHD  
**EASTING:** 333589  
**NORTHING:** 6249766  
**DIP/AZIMUTH:** 90°/-

**BORE No:** BH105  
**PROJECT No:** 86874.00  
**DATE:** 28/10/2019  
**SHEET 1 OF 2**

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing				Water	Well Construction Details	
				Type	Depth	Sample	Results & Comments			
6	0.23	CONCRETE: pale grey, aggregate size from 15-45mm, 8mm reinforcing steel		A/E	0.3		PID<1		Flush Gatic Cover	
		FILL/ SAND: fine to coarse, yellow brown, with fine to coarse mixed origin gravel, moist (wet at top from diatube)		A/E	0.5		PID<1			
		0.7-.9m: sandstone boulders								
1		0.9m: colour change to dark brown dark grey, trace fine to coarse mixed origin gravel		A/E	1.0		PID<1		1	Gravel
				S			2,4,4 N = 8			
		1.4m: trace building rubble (fragments of brick, terracotta and ceramics)		E	1.45		PID<1			
					1.5					
2		2.0m: with low plasticity clay, trace sandstone cobbles		E*	2.0		PID<1		2	Blank
				E	2.5		PID<1 1,2,5 N = 7			
				S						
3				A/E	2.95		PID<1		3	
					3.0					
3.5		Sandy CLAY CL: low plasticity, fine to medium, dark grey dark brown, w<PL, soft to firm, alluvial		E	3.5		PID<1			
4	4.0	Sandy CLAY CL-CI: low to medium plasticity, fine to medium, yellow brown, w<PL, stiff, residual		E	4.0		PID<1 1,4,11 N = 15		4	
				S						
4.5		SANDSTONE: medium to coarse, yellow-brown, iron stained, low strength, moderately weathered, Hawkesbury Sandstone			4.45		PL(A) = 0.2			
					4.5					
					4.55					
5	5.0								5	
5.38		SANDSTONE: medium to coarse, pale grey and yellow-brown some red-brown, iron stained, indistinctly bedded, medium then high strength, slightly weathered, slightly fractured, Hawkesbury Sandstone		C	5.5		PL(A) = 0.5			
6									6	
					6.6		PL(A) = 0.9			Gravel
7					7.11				7	Slotted Pipe
					7.5		PL(A) = 1			
8	8.0	SANDSTONE: as above							8	
				C	8.5		PL(A) = 1.4			
8.62		SANDSTONE: medium to coarse, pale grey, indistinctly bedded, high strength, fresh, slightly fractured, Hawkesbury Sandstone							9	
9										
9.33		SANDSTONE: fine to medium, pale grey, indistinctly bedded, high strength with very low to low strength beds, fresh, Hawkesbury Sandstone			9.45		PL(A) = 0.8			
		9.33-9.45m: siltstone bed			10.0		PL(A) = 1.3			

**RIG:** Bobcat

**DRILLER:** JE

**LOGGED:** RK

**CASING:** HW to 4.0m, HQ to 4.5m

**TYPE OF BORING:** Diatube to 0.23m, Solid Flight Auger (TC-bit) to 4.00m, wash boring to 4.50m, NMLC coring to 10.15m

**WATER OBSERVATIONS:** No free groundwater observed during augering

**REMARKS:** \*replicate BD4\_20191028

## SAMPLING & IN SITU TESTING LEGEND



A	Auger sample	G	Gas sample	PID	Photo ionisation detector (ppm)
B	Bulk sample	P	Piston sample	PL(A)	Point load axial test Is(50) (MPa)
BLK	Block sample	U	Tube sample (x mm dia.)	PL(D)	Point load diametral test Is(50) (MPa)
C	Core drilling	W	Water sample	pp	Pocket penetrometer (kPa)
D	Disturbed sample	W	Water seep	S	Standard penetration test
E	Environmental sample	W	Water level	V	Shear vane (kPa)

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street,  
 Ultimo

**SURFACE LEVEL:** 6.2 AHD  
**EASTING:** 333589  
**NORTHING:** 6249766  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH105  
**PROJECT No:** 86874.00  
**DATE:** 28/10/2019  
**SHEET 2 OF 2**

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing			Water	Well Construction Details	
				Type	Depth	Sample		Results & Comments	
-4	10.15	9.55-9.61m: siltstone bed		C	10.15			End Cap	
		Bore discontinued at 10.15m Target depth reached							
-5									
-6									
-7									
-8									
-9									
-10									
-11									
-12									
-13									
-14									
-15									
-16									
-17									
-18									
-19									
-20									

**RIG:** Bobcat

**DRILLER: JE**

**LOGGED: RK**

**CASING:** HW to 4.0m, HQ to 4.5m

**TYPE OF BORING:** Diatube to 0.23m, Solid Flight Auger (TC-bit) to 4.00m, wash boring to 4.50m, NMLC coring to 10.15m

**WATER OBSERVATIONS:** No free groundwater observed during augering

REMARKS: \*replicate BD4\_20191028

SAMPLING & IN SITU TESTING LEGEND			
A	Auger sample	G	Gas sample
B	Bulk sample	P	Piston sample
BLK	Block sample	U	Tube sample (x mm dia.)
C	Core drilling	W	Water sample
D	Disturbed sample	W	Water seep
E	Environmental sample	W	Water level
		PID	Photo ionisation detector (ppm)
		PL(A)	Point load axial test ls(50) (MPa)
		PL(D)	Point load diametral test ls(50) (MPa)
		pp	Pocket penetrometer (kPa)
		S	Standard penetration test
		V	Shear vane (kPa)



# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 7.0 AHD  
**EASTING:** 333544  
**NORTHING:** 6249786  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH106  
**PROJECT No:** 86874.00  
**DATE:** 28/10/2019  
**SHEET 1 OF 2**

RL	Depth (m)	Description of Strata	Degree of Weathering EW HW MW SW FS FR	Graphic Log	Rock Strength Ex Low Very Low Low Medium High Very High Ex High	Water	Fracture Spacing (m) 0.01 0.05 0.10 0.50 1.00	Discontinuities B - Bedding J - Joint S - Shear F - Fault		Sampling & In Situ Testing				Test Results & Comments
										Type	Core Rec. %	RQD %		
	0.1	BRICKS PAVERS								A/E				PID<1
		FILL/ SAND: fine to coarse grained, dark brown-dark grey, trace fine to coarse mixed origin gravel, moist								A/E				PID<1
	1									A/E				PID<1
	1.5	Sandy CLAY CL-Cl: low to medium plasticity, fine to medium grained, yellow brown, w<PL, firm to stiff, residual								A/E				PID<1
	2									A/E*				PID<1
	2.5	Sandy CLAY CL-Cl: low to medium plasticity, fine to medium grained, pale grey -yellow, w<PL, stiff, residual								S				4,4,6 N = 10
	3													
	3.5	SANDSTONE: medium to coarse grained, yellow and pale-brown, inferred low-medium strength												
	4.0	SANDSTONE: medium to coarse grained, pale grey and yellow-brown, indistinctly bedded to massive, medium to high strength, slightly weathered, Hawkesbury Sandstone												PL(A) = 1
	5													
	5.35	SANDSTONE: medium to coarse grained, pale grey, indistinctly bedded, high strength, fresh, Hawkesbury Sandstone								C	100	92		PL(A) = 1
	6													PL(A) = 1.9
	7													
	8													PL(A) = 1.8
	9													PL(A) = 2.1
														PL(A) = 2.8

**RIG:** Bobcat  
**DRILLER:** JE  
**LOGGED:** RK  
**CASING:** HW to 2.5, HQ to 4.0m

**TYPE OF BORING:** Hand auger to 1.3m, Solid Flight Auger (TC-bit) to 2.5m, wash boring to 4.0m, NMLC coring to 10.06m

**WATER OBSERVATIONS:** No free groundwater observed during augering

**REMARKS:** \*replicate BD5-20191028

SAMPLING & IN SITU TESTING LEGEND			
A Auger sample	G Gas sample	PID Photo ionisation detector (ppm)	
B Bulk sample	P Piston sample	PL(A) Point load axial test Is(50) (MPa)	
BLK Block sample	U Tube sample (x mm dia.)	PL(D) Point load diametral test Is(50) (MPa)	
C Core drilling	W Water sample	pp Pocket penetrometer (kPa)	
D Disturbed sample	> Water seep	S Standard penetration test	
E Environmental sample	≡ Water level	V Shear vane (kPa)	

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 7.0 AHD  
**EASTING:** 333544  
**NORTHING:** 6249786  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH106  
**PROJECT No:** 86874.00  
**DATE:** 28/10/2019  
**SHEET 2 OF 2**

[illegible]

**RIG:** Bobcat

**DRILLER: JE**

LOGGED: RK

**CASING:** HW to 2.5, HQ to 4.0m

**TYPE OF BORING:** Hand auger to 1.3m, Solid Flight Auger (TC-bit) to 2.5m, wash boring to 4.0m, NMLC coring to 10.06m

**WATER OBSERVATIONS:** No free groundwater observed during augering

REMARKS: \*replicate BD5-20191028

SAMPLING & IN SITU TESTING LEGEND			
A	Auger sample	G	Gas sample
B	Bulk sample	P	Piston sample
BLK	Block sample	U	Tube sample (x mm dia.)
C	Core drilling	W	Water sample
D	Disturbed sample	W	Water seep
E	Environmental sample	W	Water level
		PID	Photo ionisation detector (ppm)
		PL(A)	Point load axial test (s(50) (MPa)
		PL(D)	Point load diametral test (s(50) (MPa)
		pp	Pepp penetrometer (kPa)
		S	Standard penetration test
		V	Shear vane (kPa)





BORE: 106

PROJECT: ULTIMO

OCTOBER 2019



**Douglas Partners**  
Geotechnics | Environment | Groundwater

Project No: 86874.00

BH ID: BH 106

Depth: 4.00 - 8.00 m

Core Box No.: 1



Powerhouse Museum 86874.00 BH106 Start at 4.00m 28/10/19



4.00 - 8.00m

BORE: 106

PROJECT: ULTIMO

OCTOBER 2019



**Douglas Partners**  
Geotechnics | Environment | Groundwater

Project No: 86874.00

BH ID: BH 106

Depth: 8.00 - 10.06 m

Core Box No.: 2



8.00 - 10.06m

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo  
**SURFACE LEVEL:** 5.8 AHD  
**EASTING:** 333494  
**NORTHING:** 6249446  
**DIP/AZIMUTH:** 90°/--  
**BORE No:** BH107  
**PROJECT No:** 86874.00  
**DATE:** 22/10/2019  
**SHEET** 1 OF 1

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing				Water	Well Construction Details	
				Type	Depth	Sample	Results & Comments			
	0.3	CONCRETE: suspended slab	△							
		VOID: probable disused storage space	△							
	1									
	2									
	3									
	3.1	Bore discontinued at 3.1m Test Bore discontinued due to substantial void space								
	4									

**RIG:** Hand tools  
**DRILLER:** TG  
**LOGGED:** TG  
**CASING:** Uncased  
**TYPE OF BORING:** Diatube to 0.3m  
**WATER OBSERVATIONS:** No free groundwater observed during augering  
**REMARKS:** Level interpolated from LTS Lockley drawing 50686 001DT, 30/05/19



SAMPLING & IN SITU TESTING LEGEND					
A	Auger sample	G	Gas sample	PID	Photo ionisation detector (ppm)
B	Bulk sample	P	Piston sample	PL(A)	Point load axial test Is(50) (MPa)
BLK	Block sample	U	Tube sample (x mm dia.)	PL(D)	Point load diametral test Is(50) (MPa)
C	Core drilling	W	Water sample	pp	Pocket penetrometer (kPa)
D	Disturbed sample	W	Water seep	S	Standard penetration test
E	Environmental sample	W	Water level	V	Shear vane (kPa)

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 3.5 AHD  
**EASTING:**  
**NORTHING:**  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH108  
**PROJECT No:** 86874.00  
**DATE:** 22/10/2019  
**SHEET 1 OF 1**

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing				Water	Well Construction Details	
				Type	Depth	Sample	Results & Comments			
	0.1	CONCRETE			0.1					
	0.25	FILL/ GRAVEL: dense, grey angular igneous gravel with trace sand		E*	0.2		PID<1			
		Bore discontinued at 0.25m refusal on sandstone or sandstone boulder in fill								

**RIG:** Hand tools

**DRILLER: TG**

LOGGED: TG

**CASING:** Uncased

**TYPE OF BORING:** Diatube to 0.1m; Hand auger to 0.25m

**WATER OBSERVATIONS:** No free groundwater observed during augering



**REMARKS:** \*replicate BDA/2019022, Level from drawing titled PLAN BASEMENT LEVEL, May 2005

SAMPLING & IN SITU TESTING LEGEND			
A	Auger sample	G	Gas sample
B	Bulk sample	P	Piston sample
BLK	Block sample	U <sub>s</sub>	Tube sample (x mm dia.)
C	Core drilling	W	Water sample
D	Disturbed sample	W <sub>s</sub>	Water seep
E	Environmental sample	W <sub>l</sub>	Water level
		PID	Photo ionisation detector (ppm)
		PL(A)	Point load axial test Is(50) (MPa)
		PL(D)	Point load diametral test Is(50) (MPa)
		pp	Pocket penetrometer (kPa)
		S	Standard penetration test
		V	Shear vane (kPa)



# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo  
**SURFACE LEVEL:** 3.5 AHD  
**EASTING:**  
**NORTHING:**  
**DIP/AZIMUTH:** 90°/--  
**BORE No:** BH109  
**PROJECT No:** 86874.00  
**DATE:** 22/10/2019  
**SHEET** 1 OF 1

RL	Depth (m)	Description of Strata	Graphic Log	Sampling & In Situ Testing				Water	Well Construction Details	
				Type	Depth	Sample	Results & Comments			
	0.15	CONCRETE								
	0.25	FILL/ GRAVEL: grey angular igneous gravel with trace sand								
		Bore discontinued at 0.25m refusal on sandstone or sandstone boulder in fill								
	1									
	2									
	3									
	4									

**RIG:** Hand tools  
**DRILLER:** TG  
**LOGGED:** TG  
**CASING:** Uncased  
**TYPE OF BORING:** Diatube to 0.15m; Hand auger to 0.25m  
**WATER OBSERVATIONS:** No free groundwater observed during augering  
**REMARKS:** Level from drawing titled PLAN BASEMENT LEVEL, May 2005

SAMPLING & IN SITU TESTING LEGEND					
A	Auger sample	G	Gas sample	PID	Photo ionisation detector (ppm)
B	Bulk sample	P	Piston sample	PL(A)	Point load axial test Is(50) (MPa)
BLK	Block sample	U	Tube sample (x mm dia.)	PL(D)	Point load diametral test Is(50) (MPa)
C	Core drilling	W	Water sample	pp	Pocket penetrometer (kPa)
D	Disturbed sample	W	Water seep	S	Standard penetration test
E	Environmental sample	W	Water level	V	Shear vane (kPa)

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street,  
 Ultimo

**SURFACE LEVEL:** 15.6 AHD  
**EASTING:** 333423  
**NORTHING:** 6249912  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH201  
**PROJECT No:** 86874.00  
**DATE:** 15/11/2019  
**SHEET** 1 OF 1

[illegible]

Bore discontinued at 10.0m

**RIG:** Scout Target depth reached

**DRILLER: JE**

**LOGGED: LS**

**CASING:** HW to 1.8m

**TYPE OF BORING:** Diatube 0.4m, Solid Flight Auger (TC-bit) to 1.8m, NMLC coring to 10.0m

**WATER OBSERVATIONS:** No free groundwater observed during augering

REMARKS: \*BD20191115-1

SAMPLING & IN SITU TESTING LEGEND			
A	Auger sample	G	Gas sample
B	Bulk sample	P	Piston sample
BLK	Block sample	U	Tube sample (x mm dia.)
C	Core drilling	W	Water sample
D	Disturbed sample	W	Water seep
E	Environmental sample	W	Water level
		PID	Photo ionisation detector (ppm)
		PL(A)	Point load axial test Is(50) (MPa)
		PL(D)	Point load diametral test Is(50) (MPa)
		pp	Pocket penetrometer (kPa)
		S	Standard penetration test
		V	Shear vane (kPa)



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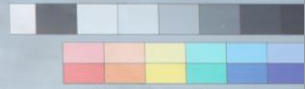
BORE: 201

PROJECT: ULTIMO

OCTOBER 2019



Project No: 86874.00  
BH ID: BH201  
Depth: 1.8-6.0m  
Core Box No.: 10F2.



1.80 – 6.00m

BORE: 201

PROJECT: ULTIMO

OCTOBER 2019



Project No: 86874.00  
BH ID: BH201  
Depth: 6.0-10.0m  
Core Box No.: 20F2.



6.00 – 10.00m



# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 5.4 AHD  
**EASTING:** 333613  
**NORTHING:** 6249647  
**DIP/AZIMUTH:** 90°/-

**BORE No:** BH202  
**PROJECT No:** 86874.00  
**DATE:** 26/11/2019  
**SHEET 1 OF 2**

RL	Depth (m)	Description of Strata	Degree of Weathering				Graphic Log	Rock Strength					Water	Fracture Spacing (m)	Discontinuities		Sampling & In Situ Testing				
			EW	HW	MW	SW		FS	FR	Ex Low	Very Low	Low			Medium	High	Very High	Ex High	B - Bedding S - Shear	J - Joint F - Fault	Type
5 1 4 2 3 2 4 5 0	0.1	CONCRETE: pale grey, aggregate 10-40mm, 8mm diameter steel reinforcement																A/E			PID<1
		FILL/Clayey SAND: fine to medium, pale brown-brown, low plasticity clay, with fine sandstone gravel, trace silt, dry																A/E			PID<1
	0.8																	A/E			PID<1
	1.2	FILL/Gravelly SAND: fine to coarse, grey-dark grey, fine to coarse igneous gravel with clay, moist																S A/E			PID<1 24/120 refusal HB PID<1
		FILL/Clayey SAND: fine to coarse, dark grey, low plasticity clay, with fine igneous gravel and silt, moist, slight hydrocarbon odour																A/E*			PID=1
	2.0																	E			PID<1
		FILL/Silty SAND: fine to medium, dark grey-dark brown, with fine sandstone gravel, moist																S			3,6,14 N = 20
		2.30m: with terracotta and ceramic fragments and clay																			PID<1
	3.1																	E			PID<1
		Silty CLAY CH: high plasticity, grey mottled yellow-brown, trace fine ironstone gravel, w<PL, stiff, appears alluvial (possibly residual)																A/E			PID<1
4 1 5 0 6 -1 -2 -3 -4																		S			5,11/50 refusal HB
	5.0	Sandy CLAY CL: low plasticity, pale grey, fine to medium sand, w<PL, hard, residual																			PL(A) = 1.1
	5.7																	C	100	88	PL(A) = 1.8
		SANDSTONE: medium to coarse, pale grey, indistinctly bedded, high strength, fresh, slightly fractured, Hawkesbury Sandstone																			
		6.03m: becoming yellow-brown and pale grey, slightly weathered																			
9 -4																					
	9.22m: carbonaceous bed (5mm thick)																				
	9.48	SANDSTONE: fine to medium, pale grey-pale brown, distinct carbonaceous laminations at 0-5°																			
	10.0																				

**RIG:** Scout 1 **DRILLER:** JE **LOGGED:** RK **CASING:** HW to 4.3m, HQ to 5.5m

**TYPE OF BORING:** Diatube to 0.1m, Solid Flight Auger (TC-bit) to 4.5m, wash boring to 5.7m, NMLC to 10.0m

**WATER OBSERVATIONS:** No free groundwater observed during augering

**REMARKS:** \*BD26112019, HB = SPT hammer bouncing

SAMPLING & IN SITU TESTING LEGEND			
A Auger sample	G Gas sample	PID Photo ionisation detector (ppm)	
B Bulk sample	P Piston sample	PL(A) Point load axial test Is(50) (MPa)	
BLK Block sample	U Tube sample (x mm dia.)	PL(D) Point load diametral test Is(50) (MPa)	
C Core drilling	W Water sample	pp Pocket penetrometer (kPa)	
D Disturbed sample	> Water seep	S Standard penetration test	
E Environmental sample	≡ Water level	V Shear vane (kPa)	

# BOREHOLE LOG

**CLIENT:** Creat NSW, Department of Premier and Cabinet  
**PROJECT:** Ultimo Creative Industries Precinct  
**LOCATION:** Powerhouse Museum, 500 Harris Street, Ultimo

**SURFACE LEVEL:** 5.4 AHD  
**EASTING:** 333613  
**NORTHING:** 6249647  
**DIP/AZIMUTH:** 90°/--

**BORE No:** BH202  
**PROJECT No:** 86874.00  
**DATE:** 26/11/2019  
**SHEET 2 OF 2**

RL	Depth (m)	Description of Strata	Degree of Weathering					Graphic Log	Rock Strength					Water	Fracture Spacing (m)	Discontinuities		Sampling & In Situ Testing			
			EW	HW	MW	SW	FS		FR	Ex Low	Very Low	Low	Medium			High	Very High	Ex High	B - Bedding S - Shear	J - Joint F - Fault	Type
	-5	<div>very low strength, highly weathered, Hawkesbury Sandstone</div> <div>9.70m: becoming indistinctly bedded, low strength</div> <div>Bore discontinued at 10.0m</div> <div>Target depth reached</div>																			
	-11																				
	-6																				
	-12																				
	-7																				
	-13																				
	-8																				
	-14																				
	-9																				
	-10																				
	-16																				
	-11																				
	-17																				
	-12																				
	-18																				
	-13																				
	-19																				
	-14																				

**RIG:** Scout 1      **DRILLER:** JE      **LOGGED:** RK      **CASING:** HW to 4.3m, HQ to 5.5m  
**TYPE OF BORING:** Diatube to 0.1m, Solid Flight Auger (TC-bit) to 4.5m, wash boring to 5.7m, NMLC to 10.0m  
**WATER OBSERVATIONS:** No free groundwater observed during augering  
**REMARKS:** \*BD26112019, HB = SPT hammer bouncing

SAMPLING & IN SITU TESTING LEGEND			
A	Auger sample	G	Gas sample
B	Bulk sample	P	Piston sample
BLK	Block sample	U	Tube sample (x mm dia.)
C	Core drilling	W	Water sample
D	Disturbed sample	>	Water seep
E	Environmental sample	≡	Water level
		PID	Photo ionisation detector (ppm)
		PL(A)	Point load axial test Is(50) (MPa)
		PL(D)	Point load diametral test Is(50) (MPa)
		pp	Pocket penetrometer (kPa)
		S	Standard penetration test
		V	Shear vane (kPa)

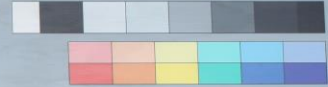
BORE: 202

PROJECT: ULTIMO

NOVEMBER 2019



Project No: 86874.00  
BH ID: BH202  
Depth: 5.70-10.00  
Core Box No.: 1/1



Ultimo 86874.00 BH202 Start at 5.70m



5.70 - 10.00m



## Sampling

Sampling is carried out during drilling or test pitting to allow engineering examination (and laboratory testing where required) of the soil or rock.

Disturbed samples taken during drilling provide information on colour, type, inclusions and, depending upon the degree of disturbance, some information on strength and structure.

Undisturbed samples are taken by pushing a thin-walled sample tube into the soil and withdrawing it to obtain a sample of the soil in a relatively undisturbed state. Such samples yield information on structure and strength, and are necessary for laboratory determination of shear strength and compressibility. Undisturbed sampling is generally effective only in cohesive soils.

## Test Pits

Test pits are usually excavated with a backhoe or an excavator, allowing close examination of the in-situ soil if it is safe to enter into the pit. The depth of excavation is limited to about 3 m for a backhoe and up to 6 m for a large excavator. A potential disadvantage of this investigation method is the larger area of disturbance to the site.

## Large Diameter Augers

Boreholes can be drilled using a rotating plate or short spiral auger, generally 300 mm or larger in diameter commonly mounted on a standard piling rig. The cuttings are returned to the surface at intervals (generally not more than 0.5 m) and are disturbed but usually unchanged in moisture content. Identification of soil strata is generally much more reliable than with continuous spiral flight augers, and is usually supplemented by occasional undisturbed tube samples.

## Continuous Spiral Flight Augers

The borehole is advanced using 90-115 mm diameter continuous spiral flight augers which are withdrawn at intervals to allow sampling or in-situ testing. This is a relatively economical means of drilling in clays and sands above the water table. Samples are returned to the surface, or may be collected after withdrawal of the auger flights, but they are disturbed and may be mixed with soils from the sides of the hole. Information from the drilling (as distinct from specific sampling by SPTs or undisturbed samples) is of relatively low

reliability, due to the remoulding, possible mixing or softening of samples by groundwater.

## Non-core Rotary Drilling

The borehole is advanced using a rotary bit, with water or drilling mud being pumped down the drill rods and returned up the annulus, carrying the drill cuttings. Only major changes in stratification can be determined from the cuttings, together with some information from the rate of penetration. Where drilling mud is used this can mask the cuttings and reliable identification is only possible from separate sampling such as SPTs.

## Continuous Core Drilling

A continuous core sample can be obtained using a diamond tipped core barrel, usually with a 50 mm internal diameter. Provided full core recovery is achieved (which is not always possible in weak rocks and granular soils), this technique provides a very reliable method of investigation.

## Standard Penetration Tests

Standard penetration tests (SPT) are used as a means of estimating the density or strength of soils and also of obtaining a relatively undisturbed sample. The test procedure is described in Australian Standard 1289, Methods of Testing Soils for Engineering Purposes - Test 6.3.1.

The test is carried out in a borehole by driving a 50 mm diameter split sample tube under the impact of a 63 kg hammer with a free fall of 760 mm. It is normal for the tube to be driven in three successive 150 mm increments and the 'N' value is taken as the number of blows for the last 300 mm. In dense sands, very hard clays or weak rock, the full 450 mm penetration may not be practicable and the test is discontinued.

The test results are reported in the following form.

- In the case where full penetration is obtained with successive blow counts for each 150 mm of, say, 4, 6 and 7 as:  
4,6,7  
N=13
- In the case where the test is discontinued before the full penetration depth, say after 15 blows for the first 150 mm and 30 blows for the next 40 mm as:  
15, 30/40 mm

# *Sampling Methods*

The results of the SPT tests can be related empirically to the engineering properties of the soils.

## **Dynamic Cone Penetrometer Tests / Perth Sand Penetrometer Tests**

Dynamic penetrometer tests (DCP or PSP) are carried out by driving a steel rod into the ground using a standard weight of hammer falling a specified distance. As the rod penetrates the soil the number of blows required to penetrate each successive 150 mm depth are recorded. Normally there is a depth limitation of 1.2 m, but this may be extended in certain conditions by the use of extension rods. Two types of penetrometer are commonly used.

- Perth sand penetrometer - a 16 mm diameter flat ended rod is driven using a 9 kg hammer dropping 600 mm (AS 1289, Test 6.3.3). This test was developed for testing the density of sands and is mainly used in granular soils and filling.
- Cone penetrometer - a 16 mm diameter rod with a 20 mm diameter cone end is driven using a 9 kg hammer dropping 510 mm (AS 1289, Test 6.3.2). This test was developed initially for pavement subgrade investigations, and correlations of the test results with California Bearing Ratio have been published by various road authorities.



## Description and Classification Methods

The methods of description and classification of soils and rocks used in this report are generally based on Australian Standard AS1726:2017, Geotechnical Site Investigations. In general, the descriptions include strength or density, colour, structure, soil or rock type and inclusions.

## Soil Types

Soil types are described according to the predominant particle size, qualified by the grading of other particles present:

Type	Particle size (mm)
Boulder	>200
Cobble	63 - 200
Gravel	2.36 - 63
Sand	0.075 - 2.36
Silt	0.002 - 0.075
Clay	<0.002

The sand and gravel sizes can be further subdivided as follows:

Type	Particle size (mm)
Coarse gravel	19 - 63
Medium gravel	6.7 - 19
Fine gravel	2.36 - 6.7
Coarse sand	0.6 - 2.36
Medium sand	0.21 - 0.6
Fine sand	0.075 - 0.21

Definitions of grading terms used are:

- Well graded - a good representation of all particle sizes
- Poorly graded - an excess or deficiency of particular sizes within the specified range
- Uniformly graded - an excess of a particular particle size
- Gap graded - a deficiency of a particular particle size with the range

The proportions of secondary constituents of soils are described as follows:

In fine grained soils (>35% fines)

Term	Proportion of sand or gravel	Example
And	Specify	Clay (60%) and Sand (40%)
Adjective	>30%	Sandy Clay
With	15 - 30%	Clay with sand
Trace	0 - 15%	Clay with trace sand

In coarse grained soils (>65% coarse)

- with clays or silts

Term	Proportion of fines	Example
And	Specify	Sand (70%) and Clay (30%)
Adjective	>12%	Clayey Sand
With	5 - 12%	Sand with clay
Trace	0 - 5%	Sand with trace clay

In coarse grained soils (>65% coarse)

- with coarser fraction

Term	Proportion of coarser fraction	Example
And	Specify	Sand (60%) and Gravel (40%)
Adjective	>30%	Gravelly Sand
With	15 - 30%	Sand with gravel
Trace	0 - 15%	Sand with trace gravel

The presence of cobbles and boulders shall be specifically noted by beginning the description with 'Mix of Soil and Cobbles/Boulders' with the word order indicating the dominant first and the proportion of cobbles and boulders described together.



# Soil Descriptions

## Cohesive Soils

Cohesive soils, such as clays, are classified on the basis of undrained shear strength. The strength may be measured by laboratory testing, or estimated by field tests or engineering examination. The strength terms are defined as follows:

Description	Abbreviation	Undrained shear strength (kPa)
Very soft	VS	<12
Soft	S	12 - 25
Firm	F	25 - 50
Stiff	St	50 - 100
Very stiff	VSt	100 - 200
Hard	H	>200
Friable	Fr	-

## Cohesionless Soils

Cohesionless soils, such as clean sands, are classified on the basis of relative density, generally from the results of standard penetration tests (SPT), cone penetration tests (CPT) or dynamic penetrometers (PSP). The relative density terms are given below:

Relative Density	Abbreviation	Density Index (%)
Very loose	VL	<15
Loose	L	15-35
Medium dense	MD	35-65
Dense	D	65-85
Very dense	VD	>85

## Soil Origin

It is often difficult to accurately determine the origin of a soil. Soils can generally be classified as:

- Residual soil - derived from in-situ weathering of the underlying rock;
- Extremely weathered material – formed from in-situ weathering of geological formations. Has soil strength but retains the structure or fabric of the parent rock;
- Alluvial soil – deposited by streams and rivers;

- Estuarine soil – deposited in coastal estuaries;
- Marine soil – deposited in a marine environment;
- Lacustrine soil – deposited in freshwater lakes;
- Aeolian soil – carried and deposited by wind;
- Colluvial soil – soil and rock debris transported down slopes by gravity;
- Topsoil – mantle of surface soil, often with high levels of organic material.
- Fill – any material which has been moved by man.

## Moisture Condition – Coarse Grained Soils

For coarse grained soils the moisture condition should be described by appearance and feel using the following terms:

- Dry (D) Non-cohesive and free-running.
- Moist (M) Soil feels cool, darkened in colour.  
Soil tends to stick together.  
Sand forms weak ball but breaks easily.
- Wet (W) Soil feels cool, darkened in colour.  
Soil tends to stick together, free water forms when handling.

## Moisture Condition – Fine Grained Soils

For fine grained soils the assessment of moisture content is relative to their plastic limit or liquid limit, as follows:

- 'Moist, dry of plastic limit' or 'w < PL' (i.e. hard and friable or powdery).
- 'Moist, near plastic limit' or 'w ≈ PL' (i.e. soil can be moulded at moisture content approximately equal to the plastic limit).
- 'Moist, wet of plastic limit' or 'w > PL' (i.e. soils usually weakened and free water forms on the hands when handling).
- 'Wet' or 'w ≈ LL' (i.e. near the liquid limit).
- 'Wet' or 'w > LL' (i.e. wet of the liquid limit).



## Rock Strength

Rock strength is defined by the Unconfined Compressive Strength and it refers to the strength of the rock substance and not the strength of the overall rock mass, which may be considerably weaker due to defects.

The Point Load Strength Index  $Is_{(50)}$  is commonly used to provide an estimate of the rock strength and site specific correlations should be developed to allow UCS values to be determined. The point load strength test procedure is described by Australian Standard AS4133.4.1-2007. The terms used to describe rock strength are as follows:

Strength Term	Abbreviation	Unconfined Compressive Strength MPa	Point Load Index * $Is_{(50)}$ MPa
Very low	VL	0.6 - 2	0.03 - 0.1
Low	L	2 - 6	0.1 - 0.3
Medium	M	6 - 20	0.3 - 1.0
High	H	20 - 60	1 - 3
Very high	VH	60 - 200	3 - 10
Extremely high	EH	>200	>10

\* Assumes a ratio of 20:1 for UCS to  $Is_{(50)}$ . It should be noted that the UCS to  $Is_{(50)}$  ratio varies significantly for different rock types and specific ratios should be determined for each site.

## Degree of Weathering

The degree of weathering of rock is classified as follows:

Term	Abbreviation	Description
Residual Soil	RS	Material is weathered to such an extent that it has soil properties. Mass structure and material texture and fabric of original rock are no longer visible, but the soil has not been significantly transported.
Extremely weathered	XW	Material is weathered to such an extent that it has soil properties. Mass structure and material texture and fabric of original rock are still visible
Highly weathered	HW	The whole of the rock material is discoloured, usually by iron staining or bleaching to the extent that the colour of the original rock is not recognisable. Rock strength is significantly changed by weathering. Some primary minerals have weathered to clay minerals. Porosity may be increased by leaching, or may be decreased due to deposition of weathering products in pores.
Moderately weathered	MW	The whole of the rock material is discoloured, usually by iron staining or bleaching to the extent that the colour of the original rock is not recognisable, but shows little or no change of strength from fresh rock.
Slightly weathered	SW	Rock is partially discoloured with staining or bleaching along joints but shows little or no change of strength from fresh rock.
Fresh	FR	No signs of decomposition or staining.
<i>Note: If HW and MW cannot be differentiated use DW (see below)</i>		
Distinctly weathered	DW	Rock strength usually changed by weathering. The rock may be highly discoloured, usually by iron staining. Porosity may be increased by leaching or may be decreased due to deposition of weathered products in pores.

# Rock Descriptions

## Degree of Fracturing

The following classification applies to the spacing of natural fractures in diamond drill cores. It includes bedding plane partings, joints and other defects, but excludes drilling breaks.

Term	Description
Fragmented	Fragments of <20 mm
Highly Fractured	Core lengths of 20-40 mm with occasional fragments
Fractured	Core lengths of 30-100 mm with occasional shorter and longer sections
Slightly Fractured	Core lengths of 300 mm or longer with occasional sections of 100-300 mm
Unbroken	Core contains very few fractures

## Rock Quality Designation

The quality of the cored rock can be measured using the Rock Quality Designation (RQD) index, defined as:

$$\text{RQD \%} = \frac{\text{cumulative length of 'sound' core sections} \geq 100 \text{ mm long}}{\text{total drilled length of section being assessed}}$$

where 'sound' rock is assessed to be rock of low strength or stronger. The RQD applies only to natural fractures. If the core is broken by drilling or handling (i.e. drilling breaks) then the broken pieces are fitted back together and are not included in the calculation of RQD.

## Stratification Spacing

For sedimentary rocks the following terms may be used to describe the spacing of bedding partings:

Term	Separation of Stratification Planes
Thinly laminated	< 6 mm
Laminated	6 mm to 20 mm
Very thinly bedded	20 mm to 60 mm
Thinly bedded	60 mm to 0.2 m
Medium bedded	0.2 m to 0.6 m
Thickly bedded	0.6 m to 2 m
Very thickly bedded	> 2 m

# Symbols & Abbreviations

## Douglas Partners



### Introduction

These notes summarise abbreviations commonly used on borehole logs and test pit reports.

### Drilling or Excavation Methods

C	Core drilling
R	Rotary drilling
SFA	Spiral flight augers
NMLC	Diamond core - 52 mm dia
NQ	Diamond core - 47 mm dia
HQ	Diamond core - 63 mm dia
PQ	Diamond core - 81 mm dia

### Water

▷	Water seep
▽	Water level

### Sampling and Testing

A	Auger sample
B	Bulk sample
D	Disturbed sample
E	Environmental sample
U <sub>50</sub>	Undisturbed tube sample (50mm)
W	Water sample
pp	Pocket penetrometer (kPa)
PID	Photo ionisation detector
PL	Point load strength Is(50) MPa
S	Standard Penetration Test
V	Shear vane (kPa)

### Description of Defects in Rock

The abbreviated descriptions of the defects should be in the following order: Depth, Type, Orientation, Coating, Shape, Roughness and Other. Drilling and handling breaks are not usually included on the logs.

### Defect Type

B	Bedding plane
Cs	Clay seam
Cv	Cleavage
Cz	Crushed zone
Ds	Decomposed seam
F	Fault
J	Joint
Lam	Lamination
Pt	Parting
Sz	Sheared Zone
V	Vein

### Orientation

The inclination of defects is always measured from the perpendicular to the core axis.

h	horizontal
v	vertical
sh	sub-horizontal
sv	sub-vertical

### Coating or Infilling Term

cln	clean
co	coating
he	healed
inf	infilled
stn	stained
ti	tight
vn	veneer

### Coating Descriptor

ca	calcite
cbs	carbonaceous
cly	clay
fe	iron oxide
mn	manganese
slt	silty

### Shape

cu	curved
ir	irregular
pl	planar
st	stepped
un	undulating

### Roughness

po	polished
ro	rough
sl	slickensided
sm	smooth
vr	very rough

### Other

fg	fragmented
bnd	band
qtz	quartz

# Symbols & Abbreviations

## Graphic Symbols for Soil and Rock

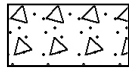
### General



Asphalt



Road base



Concrete



Filling

### Soils



Topsoil



Peat



Clay



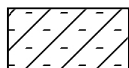
Silty clay



Sandy clay



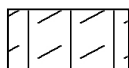
Gravelly clay



Shaly clay



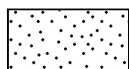
Silt



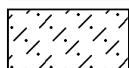
Clayey silt



Sandy silt



Sand



Clayey sand



Silty sand



Gravel



Sandy gravel



Cobbles, boulders



Talus

### Sedimentary Rocks



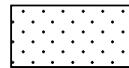
Boulder conglomerate



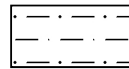
Conglomerate



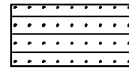
Conglomeratic sandstone



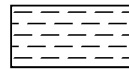
Sandstone



Siltstone



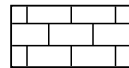
Laminite



Mudstone, claystone, shale

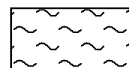


Coal

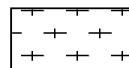


Limestone

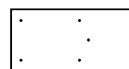
### Metamorphic Rocks



Slate, phyllite, schist

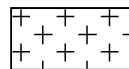


Gneiss

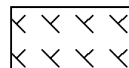


Quartzite

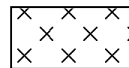
### Igneous Rocks



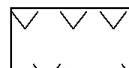
Granite



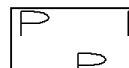
Dolerite, basalt, andesite



Dacite, epidote



Tuff, breccia



Porphyry



# About this Report

# Douglas Partners



## Introduction

These notes have been provided to amplify DP's report in regard to classification methods, field procedures and the comments section. Not all are necessarily relevant to all reports.

DP's reports are based on information gained from limited subsurface excavations and sampling, supplemented by knowledge of local geology and experience. For this reason, they must be regarded as interpretive rather than factual documents, limited to some extent by the scope of information on which they rely.

## Copyright

This report is the property of Douglas Partners Pty Ltd. The report may only be used for the purpose for which it was commissioned and in accordance with the Conditions of Engagement for the commission supplied at the time of proposal. Unauthorised use of this report in any form whatsoever is prohibited.

## Borehole and Test Pit Logs

The borehole and test pit logs presented in this report are an engineering and/or geological interpretation of the subsurface conditions, and their reliability will depend to some extent on frequency of sampling and the method of drilling or excavation. Ideally, continuous undisturbed sampling or core drilling will provide the most reliable assessment, but this is not always practicable or possible to justify on economic grounds. In any case the boreholes and test pits represent only a very small sample of the total subsurface profile.

Interpretation of the information and its application to design and construction should therefore take into account the spacing of boreholes or pits, the frequency of sampling, and the possibility of other than 'straight line' variations between the test locations.

## Groundwater

Where groundwater levels are measured in boreholes there are several potential problems, namely:

- In low permeability soils groundwater may enter the hole very slowly or perhaps not at all during the time the hole is left open;

- A localised, perched water table may lead to an erroneous indication of the true water table;
- Water table levels will vary from time to time with seasons or recent weather changes. They may not be the same at the time of construction as are indicated in the report; and
- The use of water or mud as a drilling fluid will mask any groundwater inflow. Water has to be blown out of the hole and drilling mud must first be washed out of the hole if water measurements are to be made.

More reliable measurements can be made by installing standpipes which are read at intervals over several days, or perhaps weeks for low permeability soils. Piezometers, sealed in a particular stratum, may be advisable in low permeability soils or where there may be interference from a perched water table.

## Reports

The report has been prepared by qualified personnel, is based on the information obtained from field and laboratory testing, and has been undertaken to current engineering standards of interpretation and analysis. Where the report has been prepared for a specific design proposal, the information and interpretation may not be relevant if the design proposal is changed. If this happens, DP will be pleased to review the report and the sufficiency of the investigation work.

Every care is taken with the report as it relates to interpretation of subsurface conditions, discussion of geotechnical and environmental aspects, and recommendations or suggestions for design and construction. However, DP cannot always anticipate or assume responsibility for:

- Unexpected variations in ground conditions. The potential for this will depend partly on borehole or pit spacing and sampling frequency;
- Changes in policy or interpretations of policy by statutory authorities; or
- The actions of contractors responding to commercial pressures.

If these occur, DP will be pleased to assist with investigations or advice to resolve the matter.

# *About this Report*

## **Site Anomalies**

In the event that conditions encountered on site during construction appear to vary from those which were expected from the information contained in the report, DP requests that it be immediately notified. Most problems are much more readily resolved when conditions are exposed rather than at some later stage, well after the event.

## **Information for Contractual Purposes**

Where information obtained from this report is provided for tendering purposes, it is recommended that all information, including the written report and discussion, be made available. In circumstances where the discussion or comments section is not relevant to the contractual situation, it may be appropriate to prepare a specially edited document. DP would be pleased to assist in this regard and/or to make additional report copies available for contract purposes at a nominal charge.

## **Site Inspection**

The company will always be pleased to provide engineering inspection services for geotechnical and environmental aspects of work to which this report is related. This could range from a site visit to confirm that conditions exposed are as expected, to full time engineering presence on site.

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## **Appendix C**

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Groundwater Field Sheets

## Groundwater Field Sheet

### Project and Bore Installation Details

Bore / Standpipe ID:	MW104
Project Name:	Powerhouse Museum
Project Number:	86874.01
Site Location:	Ultimo
Bore GPS Co-ord:	
Installation Date:	
GW Level (during drilling):	- m bgl
Well Depth:	m bgl
Screened Interval:	m bgl
Contaminants/Comments:	-

Bore Volume = casing volume + filter pack volume  

$$= \pi h_1 d_1^2 / 4 + n(\pi h_2 d_1^2 / 4 - \pi h_2 d_2^2 / 4)$$
  
 Where:  $\pi = 3.14$   
 $n$  = porosity (0.3 for most filter pack material)  
 $h_1$  = height of water column  
 $d_1$  = diameter of annulus  
 $h_2$  = length of filter pack  
 $d_2$  = diameter of casing  
 Bore Vol Normally:  $7.2 * h$

### Bore Development Details

Date/Time:	13/11/19 11:30am
Purged By:	CL
GW Level (pre-purge):	6.0 m bgl
GW Level (post-purge):	9.69 m bgl
PSH observed:	Yes / No ( interface / visual ). Thickness if observed:
Observed Well Depth:	11.89 m bgl
Estimated Bore Volume:	41 L
Total Volume Purged:	(target: no drill mud, min 3 well vol. or dry ) 6 vols ~100L
Equipment:	Twister pump, Interface meter

### Micropurge and Sampling Details

Date/Time:	15/11/19 11:30am
Sampled By:	CL
Weather Conditions:	Sunny
GW Level (pre-purge):	6.62 m bgl
GW Level (post sample):	7.89 m bgl
PSH observed:	Yes / No ( interface / visual ). Thickness if observed:
Observed Well Depth:	11.7 m bgl
Estimated Bore Volume:	38 L
Total Volume Purged:	27.4 L
Equipment:	Low flow pump, TDS meter, Interface meter, Boulder

### Water Quality Parameters

Time / Volume	Temp (°C)	DO (mg/L)	EC (µS or mS/cm)	pH	Turbidity	Redox (mV)
<b>Stabilisation Criteria (3 readings)</b>	<b>0.1 °C</b>	<b>+/- 0.3 mg/L</b>	<b>+/- 3%</b>	<b>+/- 0.1</b>	<b>+/- 10%</b>	<b>+/- 10 mV</b>
11:40	38.5	6.30	1467	6.62	200	-53
11:41	37.5	4.09	1533	6.60	200	-56
11:42	26.3	2.47	1626	6.58	153	-68
11:43	25.4	2.40	1643	6.57	157.1	-70
11:44	24.6	2.33	1664	6.56	149	-73
Additional Readings Following stabilisation:	DO % Sat	SPC	TDS			

### Sample Details

Sampling Depth (rationale):	7.5 m bgl,
Sample Appearance (e.g. colour, siltiness, odour):	N/A
Sample ID:	MW104
QA/QC Samples:	N/A
Sampling Containers and filtration:	1 x Amber 2 x vials, 3 x plastic
Comments / Observations:	



## Groundwater Field Sheet

### Project and Bore Installation Details

Bore / Standpipe ID:	MW105
Project Name:	Powerhouse Museum
Project Number:	86571-0
Site Location:	Ultimo
Bore GPS Co-ord:	
Installation Date:	
GW Level (during drilling):	- m bgl
Well Depth:	m bgl
Screened Interval:	m bgl
Contaminants/Comments:	-

Bore Volume = casing volume + filter pack volume  

$$= \pi h_1 d_1^2 / 4 + n(\pi h_2 d_1^2 / 4 - \pi h_2 d_2^2 / 4)$$
  
 Where:  $\pi = 3.14$   
 $n$  = porosity (0.3 for most filter pack material)  
 $h_1$  = height of water column  
 $d_1$  = diameter of annulus  
 $h_2$  = length of filter pack  
 $d_2$  = diameter of casing  
 Bore Vol Normally:  $7.2 * h$

### Bore Development Details

Date/Time:	13/11/19 10:30 am
Purged By:	CL
GW Level (pre-purge):	3.86 m bgl
GW Level (post-purge):	4.0 m bgl
PSH observed:	Yes / <input checked="" type="radio"/> No ( interface / visual ). Thickness if observed:
Observed Well Depth:	10.2 m bgl
Estimated Bore Volume:	47.8 L
Total Volume Purged:	(target: no drill mud, min 3 well vol. or dry ) full dry ~ 50L
Equipment:	twister pump, Interface meter

### Micropurge and Sampling Details

Date/Time:	15/11/19 10:30 am
Sampled By:	CL
Weather Conditions:	Sunny
GW Level (pre-purge):	3.79 m bgl
GW Level (post sample):	4.0 m bgl
PSH observed:	Yes / <input checked="" type="radio"/> No ( interface / visual ). Thickness if observed:
Observed Well Depth:	10 m bgl
Estimated Bore Volume:	44.7 L
Total Volume Purged:	3.6 L
Equipment:	Lowflow pump, Interface meter, TPS meter, Boiler

### Water Quality Parameters

Time / Volume	Temp (°C)	DO (mg/L)	EC (µS or mS/cm)	pH	Turbidity	Redox (mV)
<b>Stabilisation Criteria (3 readings)</b>	<b>0.1 °C</b>	<b>+/- 0.3 mg/L</b>	<b>+/- 3%</b>	<b>+/- 0.1</b>	<b>+/- 10%</b>	<b>+/- 10 mV</b>
10:43	26.3	1.11	1161	6.33	34.8	187
10:44	24.2	1.57	1174	6.36	33.3	183
10:45	22.8	1.35	1175	6.37	34.3	180
10:46	22.5	1.19	1179	6.37	35.0	175
10:47	22.3	1.08	1172	6.36	36.8	170
Additional Readings Following stabilisation:	DO % Sat	SPC	TDS			

### Sample Details

Sampling Depth (rationale):	5.1 m bgl, mid-column
Sample Appearance (e.g. colour, siltiness, odour):	N/A
Sample ID:	BD1/20191115 MW105
QA/QC Samples:	
Sampling Containers and filtration:	3x vials, 1x Amber, <del>1x plastic</del> 3x plastic
Comments / Observations:	



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## **Appendix D**

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QA / QC Report

## QA / QC PROCEDURES AND RESULTS

### Q1. Data Quality Objectives

The contamination investigation was prepared with reference to the seven step data quality objective (DQO) process which is provided in Appendix B, Schedule B2 of the *National Environment Protection (Assessment of Site Contamination) Measure* 1999 as amended 2013 (NEPC, 2013). The DQO process is outlined as follows:

- Stating the Problem;
- Identifying the Decision;
- Identifying Inputs to the Decision;
- Defining the Boundary of the Assessment;
- Developing a Decision Rule;
- Specifying Acceptable Limits on Decision Errors; and
- Optimising the Design for Obtaining Data.

The DQOs have been addressed within the report as shown in Table Q1.

**Table Q1: Data Quality Objectives**

Data Quality Objective	Report Section where Addressed
State the Problem	S1 Introduction
Identify the Decision	S10 Discussion S11 Conclusion
Identify Inputs to the Decision	S1 Introduction S3 Site Identification and Description S4 Summary of Previous Contamination Report S5 Preliminary Conceptual Site Model S8 Field Work Results S9 Laboratory Analytical Rational and Results
Define the Boundary of the Assessment	S3 Site Identification and Description
Develop a Decision Rule	S7 Site Assessment Criteria
Specify Acceptable Limits on Decision Errors	S6 Field Work and QA/QC
Optimise the Design for Obtaining Data	S2 Scope of Works S6 Field Work and QA/QC

## Q2. FIELD AND LABORATORY QUALITY CONTROL

The field and laboratory QC procedures and results are described below. Field work procedures are documented in Section 6. Laboratory certificates are provided in Appendix E.

### Q2.1 Trip Spike and Blank

During the course of soil sampling, two sets of soil trip spikes and soil trip blanks were taken into the field and subject to the same conditions as the soil samples that were collected in jars. Similarly, one water trip blank and water trip spike was taken into the field and subject to the same conditions as the groundwater samples.

The results of analysis of the trip spike and trip blank is shown in Tables Q2 and Q3. Trip spike recoveries between 60% to 140% and trip blank results of less than the practical quantitation limits were achieved.

**Table Q2: Trip Spike Results (%)**

Sample	Media	Benzene	Toluene	Ethylbenzene	m+p-Xylene	o-Xylene
229301-12 TS	Soil	113	108	109	116	125
230944-4 TS	Water	103	108	100	95	101
231640-3 TS	Soil	103	103	107	106	106

**Table Q3 Trip Blank Results**

Sample	Media	Benzene	Toluene	Ethylbenzene	m+p-Xylene	o-Xylene
229301-13 TB	Soil	<0.2 mg/kg	<0.5 mg/kg	<1 mg/kg	<2 mg/kg	<1 mg/kg
230944-5 TB	Water	<1 µg/L	<1 µg/L	<1 µg/L	<2 µg/L	<1 µg/L
231640-4 TB	Soil	<0.2 mg/kg	<0.5 mg/kg	<1 mg/kg	<2 mg/kg	<1 mg/kg

## Q2.2 Intra-laboratory Analysis

An intra-laboratory replicate was analysed as an internal check of the reproducibility within the primary laboratory (Envirolab Services) and as a measure of consistency of sampling techniques. The comparative results of analysis between the primary samples and intra-laboratory replicate samples are summarised in Tables Q4 to Q6.

**Table Q4: Relative Percentage Difference for Intra-laboratory Replicate BD2\_20191022**

Contaminant	Sample BH103 / 1m	Sample BD2_20191022		
	Concentration (mg/kg)	Concentration (mg/kg)	Difference (mg/kg)	RPD (%)
Arsenic	4	5	1	22
Cadmium	<0.4	<0.4	0	0
Chromium	36	36	0	0
Copper	21	17	4	21
Lead	42	47	5	11
Mercury	<0.1	<0.1	0	0
Nickel	16	14	2	13
Zinc	69	62	7	11
TRH C <sub>6</sub> - C <sub>10</sub>	<25	<25	0	0
TRH >C <sub>10</sub> -C <sub>16</sub>	<50	<50	0	0
TRH >C <sub>16</sub> -C <sub>34</sub>	<100	<100	0	0
TRH >C <sub>34</sub> -C <sub>40</sub>	<100	<100	0	0
Benzene	<0.2	<0.2	0	0
Toluene	<0.5	<0.5	0	0
Ethylbenzene	<1	<1	0	0
Total Xylenes	<3	<3	0	0
Naphthalene	<1	<1	0	0

**Table Q5: Relative Percentage Difference for Intra-laboratory Replicate BD3\_20191023**

Contaminant	Sample BH104 / 0.5m	Sample BD3_20191023		
	Concentration (mg/kg)	Concentration (mg/kg)	Difference (mg/kg)	RPD (%)
Arsenic	<4	<4	0	0
Cadmium	<0.4	<0.4	0	0
Chromium	4	8	4	<b>67</b>
Copper	3	4	1	29
Lead	36	33	3	9
Mercury	<0.1	<0.1	0	0
Nickel	<1	1	0	0
Zinc	21	22	1	5
TRH C <sub>6</sub> - C <sub>10</sub>	<25	<25	0	0
TRH >C <sub>10</sub> -C <sub>16</sub>	<50	<50	0	0
TRH >C <sub>16</sub> -C <sub>34</sub>	<100	<100	0	0
TRH >C <sub>34</sub> -C <sub>40</sub>	<100	<100	0	0
Benzene	<0.2	<0.2	0	0
Toluene	<0.5	<0.5	0	0
Ethylbenzene	<1	<1	0	0
Total Xylenes	<3	<3	0	0
Naphthalene	<1	<1	0	0



**Table Q6: Relative Percentage Difference for Intra-laboratory Replicate BD1/20191115**

Contaminant	Sample MW105	Sample BD1/20191115		
	Concentration (µg/L)	Concentration (µg/L)	Difference (µg/L)	RPD (%)
Arsenic	1	<1	0	0
Cadmium	<0.1	<0.1	0	0
Chromium	2	<1	1	<b>50</b>
Copper	4	2	2	<b>67</b>
Lead	5	<1	4	<b>133</b>
Mercury	0.06	<0.05	0.01	18
Nickel	12	11	1	9
Zinc	18	11	7	<b>48</b>
TRH C <sub>6</sub> - C <sub>10</sub>	<10	<10	0	0
TRH >C <sub>10</sub> -C <sub>16</sub>	<50	<50	0	0
TRH >C <sub>16</sub> -C <sub>34</sub>	<100	<100	0	0
TRH >C <sub>34</sub> -C <sub>40</sub>	<100	<100	0	0
Benzene	<1	<1	0	0
Toluene	<1	<1	0	0
Ethylbenzene	<1	<1	0	0
o-Xylene	<1	<1	0	0
m+p xylene	<2	<2	0	0
Naphthalene	<1	<1	0	0

The calculated relative percentage difference (RPD) values were less than 50% for organics. A number of calculated RPD values were greater than 30% for inorganics (metals) as shown in bold, however, these are not considered to be significant because the actual concentration differences between samples is small indicating little variation in concentrations between replicate pairs. Overall, the intra-laboratory replicate comparisons indicate that the sampling techniques were generally consistent and repeatable.

### Q2.3 Inter-laboratory Analysis

Inter-laboratory replicates were conducted as a check of the reproducibility of results between the primary laboratory (Envirolab Services) and the secondary laboratory (Eurofins) and as a measure of consistency of sampling techniques. The comparative results of analysis between the primary and inter-laboratory replicate samples are summarised in Tables Q7 and Q8.

**Table Q7: Relative Percentage Difference for Inter-laboratory Replicate BDA/20191022**

Contaminant	Sample BH108 / 0.1-0.2 m	Sample BDA/20191022		
	Concentration (mg/kg)	Concentration (mg/kg)	Difference (mg/kg)	RPD (%)
Arsenic	<4	<2	0	0
Cadmium	<0.4	<0.4	0	0
Chromium	34	67	33	<b>65</b>
Copper	10	22	12	<b>75</b>
Lead	6	14	8	<b>80</b>
Mercury	<0.1	<0.1	0	0
Nickel	37	67	30	<b>58</b>
Zinc	20	31	11	<b>43</b>
TRH C <sub>6</sub> - C <sub>10</sub>	<25	<20	0	0
TRH >C <sub>10</sub> -C <sub>16</sub>	<50	<50	0	0
TRH >C <sub>16</sub> -C <sub>34</sub>	<100	<100	0	0
TRH >C <sub>34</sub> -C <sub>40</sub>	<100	<100	0	0
Benzene	<0.2	<0.1	0	0
Toluene	<0.5	<0.1	0	0
Ethylbenzene	<1	<0.1	0	0
Total Xylenes	<3	<0.3	0	0
Naphthalene	<1	<0.5	0	0

**Table Q8: Relative Percentage Difference for Inter-laboratory Replicate BD26112019**

Contaminant	Sample 202 / 2.0-2.1	Sample BD26112019		
	Concentration (mg/kg)	Concentration (mg/kg)	Difference (mg/kg)	RPD (%)
Arsenic	4	8	4	<b>67</b>
Cadmium	<0.4	<0.4	0	0
Chromium	30	45	15	<b>40</b>
Copper	70	120	50	<b>52</b>
Lead	320	1300	980	<b>121</b>
Mercury	1.4	3.9	2.5	<b>94</b>
Nickel	36	40	4	<b>11</b>
Zinc	120	240	120	<b>67</b>
TRH C <sub>6</sub> - C <sub>10</sub>	<25	<200	0	0
TRH >C <sub>10</sub> -C <sub>16</sub>	82	<500	0	0
TRH >C <sub>16</sub> -C <sub>34</sub>	780	2300	1520	<b>99</b>
TRH >C <sub>34</sub> -C <sub>40</sub>	<100	<1000	0	0
Benzene	<0.2	<0.5	0	0
Toluene	<0.5	<0.5	0	0
Ethylbenzene	<1	<0.5	0	0
Total Xylenes	<3	<1.5	0	0
Naphthalene	6	26	20	<b>163</b>

A number of calculated RPD values were above 50% for organics and 30% for inorganics (metals) as shown in bold. These are not considered to be significant because:

- In many cases, the actual concentration differences (in mg/kg) between samples is small;
- The replicate pairs are from fill soils which is heterogeneous in nature; and
- Soil replicates, rather than homogenised soil duplicates, were used to minimise the risk of possible volatile loss, hence variability can be expected.

Overall, the inter-laboratory replicate comparisons indicate that the sampling techniques were generally consistent and repeatable and the sampling handling and analytical methods for the two laboratories are comparable.

## Q2.4 Laboratory QC

A summary of laboratory QC is shown in Table Q9.

**Table Q9: Laboratory QC**

Item	Frequency	Acceptance Criteria	Achievement
Analytical laboratories used		NATA accreditation	Yes
Holding times		In accordance with NEPC (2013) which references various Australian and international standards	Yes
Laboratory / Reagent Blanks	1 per lab batch	<PQL	Yes
Laboratory duplicates	10% primary samples	Laboratory specific	Yes
Matrix Spikes	1 per lab batch	70-130% recovery (inorganics); 60-140% (organics); 10-140% (SVOC, speciated phenols)	Generally yes (see comments below)
Surrogate Spikes	organics by GC	70-130% recovery (inorganics); 60-140% (organics); 10-140% (SVOC, speciated phenols)	Generally yes (see comments below)
Control Samples	1 per lab batch	70-130% recovery (inorganics); 60-140% (organics); 10-140% (SVOC, speciated phenols)	Yes

Where laboratory QA / QC results were unable to be achieved by the laboratory, report comments were made by the laboratory. These included:

- Percent recovery in the matrix spike for sample 231640-1 was not possible to report due to the inhomogeneous nature of the element/s in the sample/s. However an acceptable recovery was obtained for the laboratory control sample (LCS).
- For TRH C<sub>10</sub>-C<sub>40</sub>, percent recovery for the surrogate and matrix spike was not possible to report as the high concentration of analytes in sample 231640-1 caused interference.
- For OCP, OPP and PCB, the PQL was raised due to interferences from analytes (other than those being tested) in sample 231640-1. Percent recovery for the matrix spike was not possible to report due to interference from analytes (other than those being tested) in sample 231640-1.
- For sample S19-No38471, RPD results for arsenic, copper and zinc were outside the acceptable range, however, the RPD reported passes Eurofins Environment Testing's QC - Acceptance Criteria.
- The limits of reporting were raised due to matrix interference in sample S19-No38471.

Overall, laboratory QA / QC results indicate that laboratory results are usable for this assessment.

### Q3. Data Quality Indicators

The reliability of field procedures and analytical results was assessed against the following data quality indicators (DQIs):

- Completeness - a measure of the amount of usable data from a data collection activity;
- Comparability - the confidence (qualitative) that data may be considered to be equivalent for each sampling and analytical event;
- Representativeness - the confidence (qualitative) of data representativeness of media present on-site;
- Precision - a measure of variability or reproducibility of data; and
- Accuracy - a measure of closeness of the data to the 'true' value.

The DQIs were assessed as outlined in the following Table Q10.

**Table Q10: Data Quality Indicators**

<b>Data Quality Indicator</b>	<b>Method(s) of Achievement</b>
Completeness	Preparation of borehole logs, sample location plan and chain of custody records; Preparation of field groundwater sampling sheets; Laboratory sample receipt information received confirming receipt of samples intact and appropriateness of the chain of custody; Samples analysed for identified potential contaminants; Completion of chain of custody (COC) documentation; NATA accredited laboratory results certificates provided by the laboratory; Satisfactory frequency and results for field and laboratory quality control (QC) samples.
Comparability	Using appropriate techniques for sample recovery, storage and transportation, which were the same for the duration of the project; Experienced samplers used; Use of NATA registered laboratories, with test methods the same or similar between laboratories; Satisfactory results for field and laboratory QC samples.
Representativeness	Target media sampled; Sample numbers recovered and analysed are considered to be representative of the target media and complying with DQOs; Samples were extracted and analysed within holding times; Samples were analysed in accordance with the COC.

<b>Data Quality Indicator</b>	<b>Method(s) of Achievement</b>
Precision	Field staff followed standard operating procedures; Acceptable RPD between original samples and replicates; Satisfactory results for all other field and laboratory QC samples.
Accuracy	Field staff followed standard operating procedures; Satisfactory results for all field and laboratory QC samples.

Based on the above, it is considered that the DQIs have been complied with. As such, it is concluded that the field and laboratory test data obtained are reliable and useable for this assessment.



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## Appendix E

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Laboratory Certificates

Chain of Custody

## CERTIFICATE OF ANALYSIS 229301

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker
<b>Address</b>	96 Hermitage Rd, West Ryde, NSW, 2114

### Sample Details

<b>Your Reference</b>	<b>86874.01, Ultimo</b>
<b>Number of Samples</b>	13 Soil
<b>Date samples received</b>	25/10/2019
<b>Date completed instructions received</b>	25/10/2019

### Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.  
 Samples were analysed as received from the client. Results relate specifically to the samples as received.  
 Results are reported on a dry weight basis for solids and on an as received basis for other matrices.  
**Please refer to the last page of this report for any comments relating to the results.**

### Report Details

<b>Date results requested by</b>	01/11/2019
<b>Date of Issue</b>	01/11/2019
<b>Reissue Details</b>	This report replaces R00 created on 31/10/2019 due to: sample ID error
NATA Accreditation Number 2901. This document shall not be reproduced except in full.	
Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	

#### Asbestos Approved By

Analysed by Asbestos Approved Identifier: Lucy Zhu  
 Authorised by Asbestos Approved Signatory: Lucy Zhu

#### Results Approved By

Diego Bigolin, Team Leader, Inorganics  
 Jaimie Loa-Kum-Cheung, Metals Supervisor  
 Josh Williams, Chemist  
 Loren Bardwell, Senior Chemist  
 Lucy Zhu, Senior Asbestos Analyst  
 Priya Samarawickrama, Senior Chemist  
 Steven Luong, Organics Supervisor

#### Authorised By



Nancy Zhang, Laboratory Manager

VOCs in soil			
Our Reference		229301-2	229301-3
Your Reference	UNITS	BH102	BH103
Depth		0.5	1.0
Date Sampled		22/10/2019	22/10/2019
Type of sample		Soil	Soil
Date extracted	-	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019
Dichlorodifluoromethane	mg/kg	<1	<1
Chloromethane	mg/kg	<1	<1
Vinyl Chloride	mg/kg	<1	<1
Bromomethane	mg/kg	<1	<1
Chloroethane	mg/kg	<1	<1
Trichlorofluoromethane	mg/kg	<1	<1
1,1-Dichloroethene	mg/kg	<1	<1
trans-1,2-dichloroethene	mg/kg	<1	<1
1,1-dichloroethane	mg/kg	<1	<1
cis-1,2-dichloroethene	mg/kg	<1	<1
bromochloromethane	mg/kg	<1	<1
chloroform	mg/kg	<1	<1
2,2-dichloropropane	mg/kg	<1	<1
1,2-dichloroethane	mg/kg	<1	<1
1,1,1-trichloroethane	mg/kg	<1	<1
1,1-dichloropropene	mg/kg	<1	<1
Cyclohexane	mg/kg	<1	<1
carbon tetrachloride	mg/kg	<1	<1
Benzene	mg/kg	<0.2	<0.2
dibromomethane	mg/kg	<1	<1
1,2-dichloropropane	mg/kg	<1	<1
trichloroethene	mg/kg	<1	<1
bromodichloromethane	mg/kg	<1	<1
trans-1,3-dichloropropene	mg/kg	<1	<1
cis-1,3-dichloropropene	mg/kg	<1	<1
1,1,2-trichloroethane	mg/kg	<1	<1
Toluene	mg/kg	<0.5	<0.5
1,3-dichloropropane	mg/kg	<1	<1
dibromochloromethane	mg/kg	<1	<1
1,2-dibromoethane	mg/kg	<1	<1
tetrachloroethene	mg/kg	<1	<1
1,1,1,2-tetrachloroethane	mg/kg	<1	<1
chlorobenzene	mg/kg	<1	<1

VOCs in soil			
Our Reference		229301-2	229301-3
Your Reference	UNITS	BH102	BH103
Depth		0.5	1.0
Date Sampled		22/10/2019	22/10/2019
Type of sample		Soil	Soil
Ethylbenzene	mg/kg	<1	<1
bromoform	mg/kg	<1	<1
m+p-xylene	mg/kg	<2	<2
styrene	mg/kg	<1	<1
1,1,2,2-tetrachloroethane	mg/kg	<1	<1
o-Xylene	mg/kg	<1	<1
1,2,3-trichloropropane	mg/kg	<1	<1
isopropylbenzene	mg/kg	<1	<1
bromobenzene	mg/kg	<1	<1
n-propyl benzene	mg/kg	<1	<1
2-chlorotoluene	mg/kg	<1	<1
4-chlorotoluene	mg/kg	<1	<1
1,3,5-trimethyl benzene	mg/kg	<1	<1
tert-butyl benzene	mg/kg	<1	<1
1,2,4-trimethyl benzene	mg/kg	<1	<1
1,3-dichlorobenzene	mg/kg	<1	<1
sec-butyl benzene	mg/kg	<1	<1
1,4-dichlorobenzene	mg/kg	<1	<1
4-isopropyl toluene	mg/kg	<1	<1
1,2-dichlorobenzene	mg/kg	<1	<1
n-butyl benzene	mg/kg	<1	<1
1,2-dibromo-3-chloropropane	mg/kg	<1	<1
1,2,4-trichlorobenzene	mg/kg	<1	<1
hexachlorobutadiene	mg/kg	<1	<1
1,2,3-trichlorobenzene	mg/kg	<1	<1
Surrogate Dibromofluorometha	%	112	110
Surrogate aaa-Trifluorotoluene	%	98	127
Surrogate Toluene-d <sub>8</sub>	%	112	111
Surrogate 4-Bromofluorobenzene	%	119	114

vTRH(C6-C10)/BTEXN in Soil						
Our Reference		229301-1	229301-2	229301-3	229301-4	229301-5
Your Reference	UNITS	BH101	BH102	BH103	BH104	BH104
Depth		0.1	0.5	1.0	0.5	1.5
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019	29/10/2019
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	<25	<25	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	<25	<25	<25
vTPH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	<25	<25	<25
Benzene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Toluene	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Ethylbenzene	mg/kg	<1	<1	<1	<1	<1
m+p-xylene	mg/kg	<2	<2	<2	<2	<2
o-Xylene	mg/kg	<1	<1	<1	<1	<1
naphthalene	mg/kg	<1	<1	<1	<1	<1
Total +ve Xylenes	mg/kg	<3	<3	<3	<3	<3
Surrogate aaa-Trifluorotoluene	%	115	98	127	127	114

vTRH(C6-C10)/BTEXN in Soil						
Our Reference		229301-6	229301-7	229301-8	229301-9	229301-10
Your Reference	UNITS	BH104	BH104	BH108	BH109	BD2_20191022
Depth		3.0	4.0	0.1-0.2	0.15-0.2	-
Date Sampled		23/10/2019	23/10/2019	22/10/2019	22/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019	30/10/2019
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	<25	<25	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	<25	<25	<25
vTPH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	<25	<25	<25
Benzene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Toluene	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Ethylbenzene	mg/kg	<1	<1	<1	<1	<1
m+p-xylene	mg/kg	<2	<2	<2	<2	<2
o-Xylene	mg/kg	<1	<1	<1	<1	<1
naphthalene	mg/kg	<1	<1	<1	<1	<1
Total +ve Xylenes	mg/kg	<3	<3	<3	<3	<3
Surrogate aaa-Trifluorotoluene	%	115	126	132	128	86

vTRH(C6-C10)/BTEXN in Soil				
Our Reference		229301-11	229301-12	229301-13
Your Reference	UNITS	BD3_20191023	TS	TB
Depth		-	-	-
Date Sampled		23/10/2019	22/10/2019	22/10/2019
Type of sample		Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	[NA]	[NA]
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	[NA]	[NA]
vTPH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	[NA]	[NA]
Benzene	mg/kg	<0.2	113%	<0.2
Toluene	mg/kg	<0.5	108%	<0.5
Ethylbenzene	mg/kg	<1	109%	<1
m+p-xylene	mg/kg	<2	116%	<2
o-Xylene	mg/kg	<1	125%	<1
naphthalene	mg/kg	<1	[NA]	<1
Total +ve Xylenes	mg/kg	<3	[NA]	<3
Surrogate aaa-Trifluorotoluene	%	132	82	131



## svTRH (C10-C40) in Soil

Our Reference		229301-1	229301-2	229301-3	229301-4	229301-5
Your Reference	UNITS	BH101	BH102	BH103	BH104	BH104
Depth		0.1	0.5	1.0	0.5	1.5
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	28/10/2019	28/10/2019	29/10/2019	29/10/2019	29/10/2019
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	380	<100	<100	150
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100	350	<100	<100	160
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	<50	63	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	63	<50	<50	<50
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	<100	640	<100	<100	150
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	<100	200	<100	<100	160
Total +ve TRH (>C10-C40)	mg/kg	<50	900	<50	<50	310
Surrogate o-Terphenyl	%	119	129	111	110	97

## svTRH (C10-C40) in Soil

Our Reference		229301-6	229301-7	229301-8	229301-9	229301-10
Your Reference	UNITS	BH104	BH104	BH108	BH109	BD2_20191022
Depth		3.0	4.0	0.1-0.2	0.15-0.2	-
Date Sampled		23/10/2019	23/10/2019	22/10/2019	22/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019	29/10/2019
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	<100	<100	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100	<100	<100	<100	<100
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	110	<100	<100	<100	<100
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	<100	<100	<100	<100	<100
Total +ve TRH (>C10-C40)	mg/kg	110	<50	<50	<50	<50
Surrogate o-Terphenyl	%	99	95	96	92	93

svTRH (C10-C40) in Soil		
Our Reference		229301-11
Your Reference	UNITS	BD3_20191023
Depth		-
Date Sampled		23/10/2019
Type of sample		Soil
Date extracted	-	28/10/2019
Date analysed	-	29/10/2019
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	<100
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	<100
Total +ve TRH (>C10-C40)	mg/kg	<50
Surrogate o-Terphenyl	%	107

PAHs in Soil						
Our Reference		229301-1	229301-2	229301-3	229301-4	229301-5
Your Reference	UNITS	BH101	BH102	BH103	BH104	BH104
Depth		0.1	0.5	1.0	0.5	1.5
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019	29/10/2019
Naphthalene	mg/kg	<0.1	0.2	<0.1	<0.1	<0.1
Acenaphthylene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Acenaphthene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fluorene	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Phenanthrene	mg/kg	<0.1	0.9	0.1	<0.1	0.4
Anthracene	mg/kg	<0.1	0.2	<0.1	<0.1	<0.1
Fluoranthene	mg/kg	0.2	1.8	0.2	<0.1	1
Pyrene	mg/kg	0.2	2.0	0.2	<0.1	1
Benzo(a)anthracene	mg/kg	0.1	1.2	0.1	<0.1	0.5
Chrysene	mg/kg	0.1	1.1	<0.1	<0.1	0.5
Benzo(b,j+k)fluoranthene	mg/kg	0.2	2	<0.2	<0.2	1
Benzo(a)pyrene	mg/kg	0.1	1.2	0.1	<0.05	0.54
Indeno(1,2,3-c,d)pyrene	mg/kg	<0.1	0.6	<0.1	<0.1	0.3
Dibenzo(a,h)anthracene	mg/kg	<0.1	0.2	<0.1	<0.1	0.1
Benzo(g,h,i)perylene	mg/kg	<0.1	0.8	<0.1	<0.1	0.4
Total +ve PAH's	mg/kg	1.0	12	0.79	<0.05	5.7
Benzo(a)pyrene TEQ calc (zero)	mg/kg	<0.5	1.8	<0.5	<0.5	0.8
Benzo(a)pyrene TEQ calc(half)	mg/kg	<0.5	1.8	<0.5	<0.5	0.8
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	<0.5	1.8	<0.5	<0.5	0.8
Surrogate <i>p</i> -Terphenyl-d14	%	124	105	120	114	112

PAHs in Soil					
Our Reference		229301-6	229301-7	229301-8	229301-9
Your Reference	UNITS	BH104	BH104	BH108	BH109
Depth		3.0	4.0	0.1-0.2	0.15-0.2
Date Sampled		23/10/2019	23/10/2019	22/10/2019	22/10/2019
Type of sample		Soil	Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019
Naphthalene	mg/kg	0.1	<0.1	<0.1	<0.1
Acenaphthylene	mg/kg	<0.1	<0.1	<0.1	<0.1
Acenaphthene	mg/kg	0.1	<0.1	<0.1	<0.1
Fluorene	mg/kg	0.1	<0.1	<0.1	<0.1
Phenanthrene	mg/kg	1.3	0.3	<0.1	<0.1
Anthracene	mg/kg	0.4	0.1	<0.1	<0.1
Fluoranthene	mg/kg	2.2	0.8	<0.1	<0.1
Pyrene	mg/kg	2.2	0.7	<0.1	<0.1
Benzo(a)anthracene	mg/kg	1.2	0.4	<0.1	<0.1
Chrysene	mg/kg	1	0.3	<0.1	<0.1
Benzo(b,j+k)fluoranthene	mg/kg	2	0.6	<0.2	<0.2
Benzo(a)pyrene	mg/kg	1.3	0.4	<0.05	<0.05
Indeno(1,2,3-c,d)pyrene	mg/kg	0.7	0.2	<0.1	<0.1
Dibenzo(a,h)anthracene	mg/kg	0.2	<0.1	<0.1	<0.1
Benzo(g,h,i)perylene	mg/kg	0.9	0.3	<0.1	<0.1
Total +ve PAH's	mg/kg	13	4.1	<0.05	<0.05
Benzo(a)pyrene TEQ calc (zero)	mg/kg	1.9	0.5	<0.5	<0.5
Benzo(a)pyrene TEQ calc(half)	mg/kg	1.9	0.6	<0.5	<0.5
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	1.9	0.6	<0.5	<0.5
Surrogate <i>p</i> -Terphenyl-d14	%	118	113	123	125

Organochlorine Pesticides in soil						
Our Reference		229301-1	229301-2	229301-3	229301-4	229301-8
Your Reference	UNITS	BH101	BH102	BH103	BH104	BH108
Depth		0.1	0.5	1.0	0.5	0.1-0.2
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	22/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019	29/10/2019
alpha-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
HCB	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
beta-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
gamma-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Heptachlor	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
delta-BHC	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aldrin	mg/kg	<0.1	<0.1	<0.1	<0.1	7.9
Heptachlor Epoxide	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
gamma-Chlordane	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
alpha-chlordane	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endosulfan I	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
pp-DDE	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dieldrin	mg/kg	<0.1	<0.1	<0.1	<0.1	15
Endrin	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endosulfan II	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
pp-DDD	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endrin Aldehyde	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
pp-DDT	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Endosulfan Sulphate	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Methoxychlor	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve DDT+DDD+DDE	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate TCMX	%	71	104	76	77	78

Organochlorine Pesticides in soil		
Our Reference		229301-9
Your Reference	UNITS	BH109
Depth		0.15-0.2
Date Sampled		22/10/2019
Type of sample		Soil
Date extracted	-	28/10/2019
Date analysed	-	29/10/2019
alpha-BHC	mg/kg	<0.1
HCB	mg/kg	<0.1
beta-BHC	mg/kg	<0.1
gamma-BHC	mg/kg	<0.1
Heptachlor	mg/kg	<0.1
delta-BHC	mg/kg	<0.1
Aldrin	mg/kg	2.2
Heptachlor Epoxide	mg/kg	<0.1
gamma-Chlordane	mg/kg	<0.1
alpha-chlordane	mg/kg	<0.1
Endosulfan I	mg/kg	<0.1
pp-DDE	mg/kg	<0.1
Dieldrin	mg/kg	4.1
Endrin	mg/kg	<0.1
Endosulfan II	mg/kg	<0.1
pp-DDD	mg/kg	<0.1
Endrin Aldehyde	mg/kg	<0.1
pp-DDT	mg/kg	<0.1
Endosulfan Sulphate	mg/kg	<0.1
Methoxychlor	mg/kg	<0.1
Total +ve DDT+DDD+DDE	mg/kg	<0.1
Surrogate TCMX	%	77



Organophosphorus Pesticides in Soil						
Our Reference		229301-1	229301-2	229301-3	229301-4	229301-8
Your Reference	UNITS	BH101	BH102	BH103	BH104	BH108
Depth		0.1	0.5	1.0	0.5	0.1-0.2
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	22/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019	29/10/2019
Dichlorvos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Dimethoate	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Diazinon	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chlorpyrifos-methyl	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Ronnel	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Fenitrothion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Malathion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Chlorpyrifos	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Parathion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Bromophos-ethyl	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Ethion	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Azinphos-methyl (Guthion)	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate TCMX	%	71	104	76	77	78

Organophosphorus Pesticides in Soil		
Our Reference		229301-9
Your Reference	UNITS	BH109
Depth		0.15-0.2
Date Sampled		22/10/2019
Type of sample		Soil
Date extracted	-	28/10/2019
Date analysed	-	29/10/2019
Dichlorvos	mg/kg	<0.1
Dimethoate	mg/kg	<0.1
Diazinon	mg/kg	<0.1
Chlorpyrifos-methyl	mg/kg	<0.1
Ronnel	mg/kg	<0.1
Fenitrothion	mg/kg	<0.1
Malathion	mg/kg	<0.1
Chlorpyrifos	mg/kg	<0.1
Parathion	mg/kg	<0.1
Bromophos-ethyl	mg/kg	<0.1
Ethion	mg/kg	<0.1
Azinphos-methyl (Guthion)	mg/kg	<0.1
Surrogate TCMX	%	77

PCBs in Soil						
Our Reference		229301-1	229301-2	229301-3	229301-4	229301-8
Your Reference	UNITS	BH101	BH102	BH103	BH104	BH108
Depth		0.1	0.5	1.0	0.5	0.1-0.2
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	22/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019	29/10/2019
Aroclor 1016	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1221	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1232	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1242	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1248	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1254	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Aroclor 1260	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Total +ve PCBs (1016-1260)	mg/kg	<0.1	<0.1	<0.1	<0.1	<0.1
Surrogate TCMX	%	71	104	76	77	78

PCBs in Soil		
Our Reference		229301-9
Your Reference	UNITS	BH109
Depth		0.15-0.2
Date Sampled		22/10/2019
Type of sample		Soil
Date extracted	-	28/10/2019
Date analysed	-	29/10/2019
Aroclor 1016	mg/kg	<0.1
Aroclor 1221	mg/kg	<0.1
Aroclor 1232	mg/kg	<0.1
Aroclor 1242	mg/kg	<0.1
Aroclor 1248	mg/kg	<0.1
Aroclor 1254	mg/kg	<0.1
Aroclor 1260	mg/kg	<0.1
Total +ve PCBs (1016-1260)	mg/kg	<0.1
Surrogate TCMX	%	77

## Acid Extractable metals in soil

Our Reference		229301-1	229301-2	229301-3	229301-4	229301-5
Your Reference	UNITS	BH101	BH102	BH103	BH104	BH104
Depth		0.1	0.5	1.0	0.5	1.5
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Arsenic	mg/kg	<4	7	4	<4	<4
Cadmium	mg/kg	<0.4	0.8	<0.4	<0.4	<0.4
Chromium	mg/kg	8	12	36	4	43
Copper	mg/kg	21	130	21	3	66
Lead	mg/kg	100	170	42	36	120
Mercury	mg/kg	<0.1	0.3	<0.1	<0.1	0.1
Nickel	mg/kg	7	13	16	<1	40
Zinc	mg/kg	72	360	69	21	150

## Acid Extractable metals in soil

Our Reference		229301-6	229301-7	229301-8	229301-9	229301-10
Your Reference	UNITS	BH104	BH104	BH108	BH109	BD2_20191022
Depth		3.0	4.0	0.1-0.2	0.15-0.2	-
Date Sampled		23/10/2019	23/10/2019	22/10/2019	22/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Arsenic	mg/kg	12	5	<4	<4	5
Cadmium	mg/kg	<0.4	<0.4	<0.4	<0.4	<0.4
Chromium	mg/kg	32	16	34	48	36
Copper	mg/kg	280	29	10	14	17
Lead	mg/kg	1,100	160	6	5	47
Mercury	mg/kg	11	1.2	<0.1	<0.1	<0.1
Nickel	mg/kg	24	17	37	50	14
Zinc	mg/kg	460	99	20	25	62

Acid Extractable metals in soil		
Our Reference		229301-11
Your Reference	UNITS	BD3_20191023
Depth		-
Date Sampled		23/10/2019
Type of sample		Soil
Date prepared	-	28/10/2019
Date analysed	-	28/10/2019
Arsenic	mg/kg	<4
Cadmium	mg/kg	<0.4
Chromium	mg/kg	8
Copper	mg/kg	4
Lead	mg/kg	33
Mercury	mg/kg	<0.1
Nickel	mg/kg	1
Zinc	mg/kg	22

**Misc Soil - Inorg**

Our Reference		229301-1	229301-2	229301-3	229301-4	229301-8
Your Reference	UNITS	BH101	BH102	BH103	BH104	BH108
Depth		0.1	0.5	1.0	0.5	0.1-0.2
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	22/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Total Phenolics (as Phenol)	mg/kg	<5	<5	<5	<5	<5

**Misc Soil - Inorg**

Our Reference		229301-9
Your Reference	UNITS	BH109
Depth		0.15-0.2
Date Sampled		22/10/2019
Type of sample		Soil
Date prepared	-	28/10/2019
Date analysed	-	28/10/2019
Total Phenolics (as Phenol)	mg/kg	<5



Moisture						
Our Reference	UNITS	229301-1	229301-2	229301-3	229301-4	229301-5
Your Reference		BH101	BH102	BH103	BH104	BH104
Depth		0.1	0.5	1.0	0.5	1.5
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019	29/10/2019
Moisture	%	5.0	11	5.2	7.7	12

Moisture						
Our Reference	UNITS	229301-6	229301-7	229301-8	229301-9	229301-10
Your Reference		BH104	BH104	BH108	BH109	BD2_20191022
Depth		3.0	4.0	0.1-0.2	0.15-0.2	-
Date Sampled		23/10/2019	23/10/2019	22/10/2019	22/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Date analysed	-	29/10/2019	29/10/2019	29/10/2019	29/10/2019	29/10/2019
Moisture	%	22	15	3.7	4.4	6.3

Moisture		
Our Reference	UNITS	229301-11
Your Reference		BD3_20191023
Depth		-
Date Sampled		23/10/2019
Type of sample		Soil
Date prepared	-	28/10/2019
Date analysed	-	29/10/2019
Moisture	%	8.6

Asbestos ID - soils						
Our Reference	UNITS	229301-1	229301-2	229301-3	229301-4	229301-5
Your Reference		BH101	BH102	BH103	BH104	BH104
Depth		0.1	0.5	1.0	0.5	1.5
Date Sampled		22/10/2019	22/10/2019	22/10/2019	23/10/2019	23/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date analysed	-	31/10/2019	31/10/2019	31/10/2019	31/10/2019	31/10/2019
Sample mass tested	g	Approx. 50g	Approx. 30g	Approx. 40g	Approx. 35g	Approx. 40g
Sample Description	-	Brown sandy soil & rocks	Brown sandy soil & rocks	Brown sandy soil & rocks	Beige sandy soil & rocks	Brown coarse-grained soil & rocks
Asbestos ID in soil	-	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg
		Organic fibres detected	Organic fibres detected	Organic fibres detected	Organic fibres detected	Organic fibres detected
Asbestos comments	-	NO	NO	NO	NO	NO
Trace Analysis	-	No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected

Asbestos ID - soils					
Our Reference		229301-6	229301-7	229301-8	229301-9
Your Reference	UNITS	BH104	BH104	BH108	BH109
Depth		3.0	4.0	0.1-0.2	0.15-0.2
Date Sampled		23/10/2019	23/10/2019	22/10/2019	22/10/2019
Type of sample		Soil	Soil	Soil	Soil
Date analysed	-	31/10/2019	31/10/2019	31/10/2019	31/10/2019
Sample mass tested	g	Approx. 35g	Approx. 30g	Approx. 75g	Approx. 75g
Sample Description	-	Brown coarse-grained soil & rocks	Brown coarse-grained soil & rocks	Grey bituminous soil & rocks	Grey bituminous soil & rocks
Asbestos ID in soil	-	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected
Asbestos comments	-	NO	NO	NO	NO
Trace Analysis	-	No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected

Misc Inorg - Soil			
Our Reference		229301-2	229301-5
Your Reference	UNITS	BH102	BH104
Depth		0.5	1.5
Date Sampled		22/10/2019	23/10/2019
Type of sample		Soil	Soil
Date prepared	-	28/10/2019	28/10/2019
Date analysed	-	28/10/2019	28/10/2019
pH 1:5 soil:water	pH Units	8.9	9.1

CEC			
Our Reference		229301-2	229301-5
Your Reference	UNITS	BH102	BH104
Depth		0.5	1.5
Date Sampled		22/10/2019	23/10/2019
Type of sample		Soil	Soil
Date prepared	-	29/10/2019	29/10/2019
Date analysed	-	29/10/2019	29/10/2019
Exchangeable Ca	meq/100g	21	24
Exchangeable K	meq/100g	0.4	0.5
Exchangeable Mg	meq/100g	2.9	2.8
Exchangeable Na	meq/100g	0.39	0.40
Cation Exchange Capacity	meq/100g	25	28

Method ID	Methodology Summary
<b>ASB-001</b>	Asbestos ID - Qualitative identification of asbestos in bulk samples using Polarised Light Microscopy and Dispersion Staining Techniques including Synthetic Mineral Fibre and Organic Fibre as per Australian Standard 4964-2004.
<b>AT-008</b>	Determination of VOCs sampled onto coconut shell charcoal sorbent tubes, that can be desorbed using carbon disulphide, and analysed by GC-MS.
<b>Inorg-001</b>	pH - Measured using pH meter and electrode in accordance with APHA latest edition, 4500-H+. Please note that the results for water analyses are indicative only, as analysis outside of the APHA storage times.
<b>Inorg-008</b>	Moisture content determined by heating at 105+/-5 °C for a minimum of 12 hours.
<b>Inorg-031</b>	Total Phenolics by segmented flow analyser (in line distillation with colourimetric finish). Solids are extracted in a caustic media prior to analysis.
<b>Metals-009</b>	Determination of exchangeable cations and cation exchange capacity in soils using 1M Ammonium Chloride exchange and ICP-AES analytical finish.
<b>Metals-020</b>	Determination of various metals by ICP-AES.
<b>Metals-021</b>	Determination of Mercury by Cold Vapour AAS.
<b>Org-003</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.
<b>Org-003</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID.  F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.  Note, the Total +ve TRH PQL is reflective of the lowest individual PQL and is therefore "Total +ve TRH" is simply a sum of the positive individual TRH fractions (>C10-C40).
<b>Org-006</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD.
<b>Org-006</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD. Note, the Total +ve PCBs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PCBs" is simply a sum of the positive individual PCBs.
<b>Org-012/017</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS.

Method ID	Methodology Summary
<b>Org-012/017</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-MS and/or GC-MS/MS.  Note, the Total +ve reported DDD+DDE+DDT PQL is reflective of the lowest individual PQL and is therefore simply a sum of the positive individually report DDD+DDE+DDT.
<b>Org-012/017</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013. For soil results:- 1. 'EQ PQL' values are assuming all contributing PAHs reported as <PQL are actually at the PQL. This is the most conservative approach and can give false positive TEQs given that PAHs that contribute to the TEQ calculation may not be present. 2. 'EQ zero' values are assuming all contributing PAHs reported as <PQL are zero. This is the least conservative approach and is more susceptible to false negative TEQs when PAHs that contribute to the TEQ calculation are present but below PQL. 3. 'EQ half PQL' values are assuming all contributing PAHs reported as <PQL are half the stipulated PQL. Hence a mid-point between the most and least conservative approaches above. Note, the Total +ve PAHs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PAHs" is simply a sum of the positive individual PAHs.
<b>Org-014</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS.
<b>Org-016</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.
<b>Org-016</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater. Note, the Total +ve Xylene PQL is reflective of the lowest individual PQL and is therefore "Total +ve Xylenes" is simply a sum of the positive individual Xylenes.



QUALITY CONTROL: VOCs in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	229301-3
Date extracted	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	28/10/2019
Date analysed	-			29/10/2019	2	29/10/2019	29/10/2019		29/10/2019	29/10/2019
Dichlorodifluoromethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Chloromethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Vinyl Chloride	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Bromomethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Chloroethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Trichlorofluoromethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,1-Dichloroethene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
trans-1,2-dichloroethene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,1-dichloroethane	mg/kg	1	Org-014	<1	2	<1	<1	0	128	134
cis-1,2-dichloroethene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
bromochloromethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
chloroform	mg/kg	1	Org-014	<1	2	<1	<1	0	107	110
2,2-dichloropropane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,2-dichloroethane	mg/kg	1	Org-014	<1	2	<1	<1	0	108	108
1,1,1-trichloroethane	mg/kg	1	Org-014	<1	2	<1	<1	0	108	112
1,1-dichloropropene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Cyclohexane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
carbon tetrachloride	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Benzene	mg/kg	0.2	Org-014	<0.2	2	<0.2	<0.2	0	[NT]	[NT]
dibromomethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,2-dichloropropane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
trichloroethene	mg/kg	1	Org-014	<1	2	<1	<1	0	106	110
bromodichloromethane	mg/kg	1	Org-014	<1	2	<1	<1	0	103	105
trans-1,3-dichloropropene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
cis-1,3-dichloropropene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,1,2-trichloroethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Toluene	mg/kg	0.5	Org-014	<0.5	2	<0.5	<0.5	0	[NT]	[NT]
1,3-dichloropropane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
dibromochloromethane	mg/kg	1	Org-014	<1	2	<1	<1	0	108	109
1,2-dibromoethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
tetrachloroethene	mg/kg	1	Org-014	<1	2	<1	<1	0	120	126
1,1,1,2-tetrachloroethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
chlorobenzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Ethylbenzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
bromoform	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
m+p-xylene	mg/kg	2	Org-014	<2	2	<2	<2	0	[NT]	[NT]
styrene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,1,2,2-tetrachloroethane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]

QUALITY CONTROL: VOCs in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	229301-3
o-Xylene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,2,3-trichloropropane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
isopropylbenzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
bromobenzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
n-propyl benzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
2-chlorotoluene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
4-chlorotoluene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,3,5-trimethyl benzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
tert-butyl benzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,2,4-trimethyl benzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,3-dichlorobenzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
sec-butyl benzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,4-dichlorobenzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
4-isopropyl toluene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,2-dichlorobenzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
n-butyl benzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,2-dibromo-3-chloropropane	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,2,4-trichlorobenzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
hexachlorobutadiene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
1,2,3-trichlorobenzene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Surrogate Dibromofluorometha	%		Org-014	111	2	112	119	6	101	101
Surrogate aaa-Trifluorotoluene	%		Org-014	138	2	98	123	23	124	126
Surrogate Toluene-d <sub>8</sub>	%		Org-014	110	2	112	109	3	103	100
Surrogate 4-Bromofluorobenzene	%		Org-014	111	2	119	111	7	105	100

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	229301-3
Date extracted	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	28/10/2019
Date analysed	-			29/10/2019	2	29/10/2019	29/10/2019		29/10/2019	29/10/2019
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	25	Org-016	<25	2	<25	<25	0	123	125
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	25	Org-016	<25	2	<25	<25	0	123	125
Benzene	mg/kg	0.2	Org-016	<0.2	2	<0.2	<0.2	0	115	120
Toluene	mg/kg	0.5	Org-016	<0.5	2	<0.5	<0.5	0	120	125
Ethylbenzene	mg/kg	1	Org-016	<1	2	<1	<1	0	126	126
m+p-xylene	mg/kg	2	Org-016	<2	2	<2	<2	0	126	126
o-Xylene	mg/kg	1	Org-016	<1	2	<1	<1	0	123	122
naphthalene	mg/kg	1	Org-014	<1	2	<1	<1	0	[NT]	[NT]
Surrogate aaa-Trifluorotoluene	%		Org-016	113	2	98	123	23	124	126

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Soil						Duplicate		Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	8	28/10/2019	28/10/2019		[NT]	[NT]
Date analysed	-			[NT]	8	29/10/2019	29/10/2019		[NT]	[NT]
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	25	Org-016	[NT]	8	<25	<25	0	[NT]	[NT]
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	25	Org-016	[NT]	8	<25	<25	0	[NT]	[NT]
Benzene	mg/kg	0.2	Org-016	[NT]	8	<0.2	<0.2	0	[NT]	[NT]
Toluene	mg/kg	0.5	Org-016	[NT]	8	<0.5	<0.5	0	[NT]	[NT]
Ethylbenzene	mg/kg	1	Org-016	[NT]	8	<1	<1	0	[NT]	[NT]
m+p-xylene	mg/kg	2	Org-016	[NT]	8	<2	<2	0	[NT]	[NT]
o-Xylene	mg/kg	1	Org-016	[NT]	8	<1	<1	0	[NT]	[NT]
naphthalene	mg/kg	1	Org-014	[NT]	8	<1	<1	0	[NT]	[NT]
Surrogate aaa-Trifluorotoluene	%		Org-016	[NT]	8	132	78	51	[NT]	[NT]

QUALITY CONTROL: svTRH (C10-C40) in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	229301-3
Date extracted	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	28/10/2019
Date analysed	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	29/10/2019
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	50	Org-003	<50	2	<50	<50	0	129	117
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	100	Org-003	<100	2	380	400	5	107	116
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	100	Org-003	<100	2	350	370	6	108	86
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	50	Org-003	<50	2	63	59	7	129	117
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	100	Org-003	<100	2	640	680	6	107	116
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	100	Org-003	<100	2	200	220	10	108	86
Surrogate o-Terphenyl	%		Org-003	118	2	129	112	14	115	111

QUALITY CONTROL: svTRH (C10-C40) in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	8	28/10/2019	28/10/2019		[NT]	[NT]
Date analysed	-			[NT]	8	29/10/2019	29/10/2019		[NT]	[NT]
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	50	Org-003	[NT]	8	<50	<50	0	[NT]	[NT]
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	100	Org-003	[NT]	8	<100	<100	0	[NT]	[NT]
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	100	Org-003	[NT]	8	<100	<100	0	[NT]	[NT]
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	50	Org-003	[NT]	8	<50	<50	0	[NT]	[NT]
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	100	Org-003	[NT]	8	<100	<100	0	[NT]	[NT]
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	100	Org-003	[NT]	8	<100	<100	0	[NT]	[NT]
Surrogate o-Terphenyl	%		Org-003	[NT]	8	96	94	2	[NT]	[NT]

QUALITY CONTROL: PAHs in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	229301-3
Date extracted	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	28/10/2019
Date analysed	-			29/10/2019	2	29/10/2019	29/10/2019		29/10/2019	29/10/2019
Naphthalene	mg/kg	0.1	Org-012/017	<0.1	2	0.2	0.2	0	116	121
Acenaphthylene	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Acenaphthene	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Fluorene	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	118	124
Phenanthrene	mg/kg	0.1	Org-012/017	<0.1	2	0.9	0.9	0	106	122
Anthracene	mg/kg	0.1	Org-012/017	<0.1	2	0.2	0.3	40	[NT]	[NT]
Fluoranthene	mg/kg	0.1	Org-012/017	<0.1	2	1.8	2.2	20	104	120
Pyrene	mg/kg	0.1	Org-012/017	<0.1	2	2.0	2.5	22	104	120
Benzo(a)anthracene	mg/kg	0.1	Org-012/017	<0.1	2	1.2	1.6	29	[NT]	[NT]
Chrysene	mg/kg	0.1	Org-012/017	<0.1	2	1.1	1.4	24	106	120
Benzo(b,j+k)fluoranthene	mg/kg	0.2	Org-012/017	<0.2	2	2	2.5	22	[NT]	[NT]
Benzo(a)pyrene	mg/kg	0.05	Org-012/017	<0.05	2	1.2	1.7	34	114	126
Indeno(1,2,3-c,d)pyrene	mg/kg	0.1	Org-012/017	<0.1	2	0.6	0.8	29	[NT]	[NT]
Dibenzo(a,h)anthracene	mg/kg	0.1	Org-012/017	<0.1	2	0.2	0.3	40	[NT]	[NT]
Benzo(g,h,i)perylene	mg/kg	0.1	Org-012/017	<0.1	2	0.8	1	22	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-012/017	123	2	105	104	1	109	107

QUALITY CONTROL: PAHs in Soil						Duplicate		Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	8	28/10/2019	28/10/2019		[NT]	[NT]
Date analysed	-			[NT]	8	29/10/2019	29/10/2019		[NT]	[NT]
Naphthalene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Acenaphthylene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Acenaphthene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Fluorene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Phenanthrene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Anthracene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Fluoranthene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Pyrene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Benzo(a)anthracene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Chrysene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Benzo(b,j+k)fluoranthene	mg/kg	0.2	Org-012/017	[NT]	8	<0.2	<0.2	0	[NT]	[NT]
Benzo(a)pyrene	mg/kg	0.05	Org-012/017	[NT]	8	<0.05	<0.05	0	[NT]	[NT]
Indeno(1,2,3-c,d)pyrene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Dibenzo(a,h)anthracene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Benzo(g,h,i)perylene	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-012/017	[NT]	8	123	123	0	[NT]	[NT]

QUALITY CONTROL: Organochlorine Pesticides in soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	229301-3
Date extracted	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	28/10/2019
Date analysed	-			29/10/2019	2	29/10/2019	29/10/2019		29/10/2019	29/10/2019
alpha-BHC	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	116	118
HCB	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
beta-BHC	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	92	89
gamma-BHC	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Heptachlor	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	92	93
delta-BHC	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Aldrin	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	106	108
Heptachlor Epoxide	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	104	102
gamma-Chlordane	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
alpha-chlordane	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Endosulfan I	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
pp-DDE	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	104	104
Dieldrin	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	128	114
Endrin	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	116	116
Endosulfan II	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
pp-DDD	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	104	102
Endrin Aldehyde	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
pp-DDT	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Endosulfan Sulphate	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	96	85
Methoxychlor	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	72	2	104	99	5	132	112

QUALITY CONTROL: Organochlorine Pesticides in soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	8	28/10/2019	28/10/2019		[NT]	[NT]
Date analysed	-			[NT]	8	29/10/2019	29/10/2019		[NT]	[NT]
alpha-BHC	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
HCB	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
beta-BHC	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
gamma-BHC	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Heptachlor	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
delta-BHC	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Aldrin	mg/kg	0.1	Org-012/017	[NT]	8	7.9	13	49	[NT]	[NT]
Heptachlor Epoxide	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
gamma-Chlordane	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
alpha-chlordane	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Endosulfan I	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
pp-DDE	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Dieldrin	mg/kg	0.1	Org-012/017	[NT]	8	15	24	46	[NT]	[NT]
Endrin	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Endosulfan II	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
pp-DDD	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Endrin Aldehyde	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
pp-DDT	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Endosulfan Sulphate	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Methoxychlor	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	[NT]	8	78	81	4	[NT]	[NT]



QUALITY CONTROL: Organophosphorus Pesticides in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	229301-3
Date extracted	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	28/10/2019
Date analysed	-			29/10/2019	2	29/10/2019	29/10/2019		29/10/2019	29/10/2019
Dichlorvos	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	106	106
Dimethoate	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Diazinon	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Chlorpyrifos-methyl	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Ronnel	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	114	116
Fenitrothion	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	116	116
Malathion	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	117	113
Chlorpyrifos	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	128	130
Parathion	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	122	122
Bromophos-ethyl	mg/kg	0.1	AT-008	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Ethion	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	124	128
Azinphos-methyl (Guthion)	mg/kg	0.1	Org-012/017	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	72	2	104	99	5	132	112

QUALITY CONTROL: Organophosphorus Pesticides in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	8	28/10/2019	28/10/2019		[NT]	[NT]
Date analysed	-			[NT]	8	29/10/2019	29/10/2019		[NT]	[NT]
Dichlorvos	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Dimethoate	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Diazinon	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Chlorpyrifos-methyl	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Ronnel	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Fenitrothion	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Malathion	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Chlorpyrifos	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Parathion	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Bromophos-ethyl	mg/kg	0.1	AT-008	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Ethion	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Azinphos-methyl (Guthion)	mg/kg	0.1	Org-012/017	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	[NT]	8	78	81	4	[NT]	[NT]

QUALITY CONTROL: PCBs in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	229301-3
Date extracted	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	28/10/2019
Date analysed	-			29/10/2019	2	29/10/2019	29/10/2019		29/10/2019	29/10/2019
Aroclor 1016	mg/kg	0.1	Org-006	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1221	mg/kg	0.1	Org-006	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1232	mg/kg	0.1	Org-006	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1242	mg/kg	0.1	Org-006	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1248	mg/kg	0.1	Org-006	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1254	mg/kg	0.1	Org-006	<0.1	2	<0.1	<0.1	0	80	76
Aroclor 1260	mg/kg	0.1	Org-006	<0.1	2	<0.1	<0.1	0	[NT]	[NT]
Surrogate TCMX	%		Org-006	72	2	104	99	5	132	112

QUALITY CONTROL: PCBs in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date extracted	-			[NT]	8	28/10/2019	28/10/2019		[NT]	[NT]
Date analysed	-			[NT]	8	29/10/2019	29/10/2019		[NT]	[NT]
Aroclor 1016	mg/kg	0.1	Org-006	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1221	mg/kg	0.1	Org-006	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1232	mg/kg	0.1	Org-006	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1242	mg/kg	0.1	Org-006	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1248	mg/kg	0.1	Org-006	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1254	mg/kg	0.1	Org-006	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Aroclor 1260	mg/kg	0.1	Org-006	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Surrogate TCMX	%		Org-006	[NT]	8	78	81	4	[NT]	[NT]

QUALITY CONTROL: Acid Extractable metals in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	229301-3
Date prepared	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	28/10/2019
Date analysed	-			28/10/2019	2	28/10/2019	28/10/2019		28/10/2019	28/10/2019
Arsenic	mg/kg	4	Metals-020	<4	2	7	9	25	108	107
Cadmium	mg/kg	0.4	Metals-020	<0.4	2	0.8	0.6	29	107	93
Chromium	mg/kg	1	Metals-020	<1	2	12	13	8	112	95
Copper	mg/kg	1	Metals-020	<1	2	130	120	8	107	110
Lead	mg/kg	1	Metals-020	<1	2	170	180	6	110	91
Mercury	mg/kg	0.1	Metals-021	<0.1	2	0.3	0.3	0	81	72
Nickel	mg/kg	1	Metals-020	<1	2	13	17	27	105	89
Zinc	mg/kg	1	Metals-020	<1	2	360	380	5	106	97

QUALITY CONTROL: Acid Extractable metals in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	[NT]	[NT]
Date prepared	-			[NT]	8	28/10/2019	28/10/2019		[NT]	[NT]
Date analysed	-			[NT]	8	28/10/2019	28/10/2019		[NT]	[NT]
Arsenic	mg/kg	4	Metals-020	[NT]	8	<4	<4	0	[NT]	[NT]
Cadmium	mg/kg	0.4	Metals-020	[NT]	8	<0.4	<0.4	0	[NT]	[NT]
Chromium	mg/kg	1	Metals-020	[NT]	8	34	44	26	[NT]	[NT]
Copper	mg/kg	1	Metals-020	[NT]	8	10	12	18	[NT]	[NT]
Lead	mg/kg	1	Metals-020	[NT]	8	6	6	0	[NT]	[NT]
Mercury	mg/kg	0.1	Metals-021	[NT]	8	<0.1	<0.1	0	[NT]	[NT]
Nickel	mg/kg	1	Metals-020	[NT]	8	37	41	10	[NT]	[NT]
Zinc	mg/kg	1	Metals-020	[NT]	8	20	20	0	[NT]	[NT]

QUALITY CONTROL: Misc Soil - Inorg					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	[NT]
Date prepared	-			28/10/2019	1	28/10/2019	28/10/2019		28/10/2019	[NT]
Date analysed	-			28/10/2019	1	28/10/2019	28/10/2019		28/10/2019	[NT]
Total Phenolics (as Phenol)	mg/kg	5	Inorg-031	<5	1	<5	<5	0	100	[NT]

QUALITY CONTROL: Misc Inorg - Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	[NT]
Date prepared	-			28/10/2019	[NT]	[NT]	[NT]	[NT]	28/10/2019	[NT]
Date analysed	-			28/10/2019	[NT]	[NT]	[NT]	[NT]	28/10/2019	[NT]
pH 1:5 soil:water	pH Units		Inorg-001	[NT]	[NT]	[NT]	[NT]	[NT]	102	[NT]

QUALITY CONTROL: CEC					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-7	[NT]
Date prepared	-			29/10/2019	[NT]	[NT]	[NT]	[NT]	29/10/2019	[NT]
Date analysed	-			29/10/2019	[NT]	[NT]	[NT]	[NT]	29/10/2019	[NT]
Exchangeable Ca	meq/100g	0.1	Metals-009	<0.1	[NT]	[NT]	[NT]	[NT]	83	[NT]
Exchangeable K	meq/100g	0.1	Metals-009	<0.1	[NT]	[NT]	[NT]	[NT]	93	[NT]
Exchangeable Mg	meq/100g	0.1	Metals-009	<0.1	[NT]	[NT]	[NT]	[NT]	80	[NT]
Exchangeable Na	meq/100g	0.1	Metals-009	<0.1	[NT]	[NT]	[NT]	[NT]	108	[NT]

## Result Definitions

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<b>&lt;</b>	Less than
<b>&gt;</b>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	



## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the same sample will be re-analysed. When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

## Report Comments

Asbestos: A portion of the supplied sample was sub-sampled for asbestos analysis according to Envirolab procedures.

We cannot guarantee that this sub-sample is indicative of the entire sample. Envirolab recommends supplying 40-50g of sample in its own container.

Note: Samples 229301-8 & 9 were sub-sampled from jars provided by the client.

<b>Project No:</b> 86874-01				<b>Suburb:</b> U1 time				<b>To:</b> Envirolab Services			
<b>Project Name:</b> U1 time				<b>Order Number</b>				12 Ashley Street, Chatswood NSW 2067			
<b>Project Manager:</b> David Walker				<b>Sampler:</b> T. Graham, R. Keavney				<b>Attn:</b>			
<b>Emails:</b> david.walker@douglaspartners.com.au								<b>Phone:</b> 02 9910 6200			
<b>Date Required:</b> Same day <input type="checkbox"/> 24 hours <input type="checkbox"/> 48 hours <input type="checkbox"/> 72 hours <input type="checkbox"/> Standard <input checked="" type="checkbox"/>								<b>Email:</b> sydney@envirolab.com.au			
<b>Prior Storage:</b> <input type="checkbox"/> Esky <input checked="" type="checkbox"/> Fridge <input type="checkbox"/> Shelved				<b>Do samples contain 'potential' HBM?</b> Yes <input type="checkbox"/> No <input checked="" type="checkbox"/> (If YES, then handle, transport and store in accordance with FPM HAZID)							

Sample ID	Lab ID	Date Sampled	Sample Type S - soil W - water	Container Type G - glass P - plastic	Analytes										Notes/preservation	
					Combo 8a	VOC's	Combo 3a	Combo 1a	TRN/PTX	Metals (8)	BTX only	PH + CEC				
BH101	0.1	1	22/10/19	S	G/P	✓										subsample OK for asbestos analysis
BH102	0.5	2	22/10/19	S	G/P	✓	✓						✓			
BH103	1.0	3	22/10/19	S	G/P	✓	✓						✓			
BH104	0.5	4	23/10/19	S	G/P	✓										
BH104	1.5	5	23/10/19	S	G/P			✓					✓			
BH104	3.0	6	23/10/19	S	G/P			✓								
BH104	4.0	7	23/10/19	S	G/P			✓								
BH108	0.1-0.2	8	22/10/19	S	G	✓										
BH109	0.15-0.2	9	22/10/19	S	G	✓										
BD2-20191022		10	22/10/19	S	G				✓							
BD3-20191023		11	23/10/19	S	G				✓							
BDA/20191022		12	22/10/19	S	G					✓	✓					Send sample to Enviro for analysis
TS		12		S	G							✓				
TB		13		S	G							✓				
<b>PQL (S) mg/kg</b>																<b>ANZECC PQLs req'd for all water analytes</b> <input type="checkbox"/>
<b>PQL = practical quantitation limit. If none given, default to Laboratory Method Detection Limit</b>																<b>Lab Report/Reference No.:</b>
<b>Metals to Analyse: 8HM unless specified here:</b>																
<b>Total number of samples in container:</b>				<b>Relinquished by:</b> DW				<b>Transported to laboratory by:</b>				<b>Courier</b>				
<b>Send Results to:</b> Douglas Partners Pty Ltd				<b>Address:</b> 96 Hermitage Road, West Ryde				<b>Phone:</b> 98090666				<b>Fax:</b> 98094095				
<b>Signed:</b>				<b>Received by:</b> <i>[Signature]</i>				<b>Date &amp; Time:</b> 25/10/19								

## SAMPLE RECEIPT ADVICE

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker

### Sample Login Details

<b>Your reference</b>	86874.01, Ultimo
<b>Envirolab Reference</b>	229301
<b>Date Sample Received</b>	25/10/2019
<b>Date Instructions Received</b>	25/10/2019
<b>Date Results Expected to be Reported</b>	01/11/2019

### Sample Condition

<b>Samples received in appropriate condition for analysis</b>	Yes
<b>No. of Samples Provided</b>	13 Soil
<b>Turnaround Time Requested</b>	Standard
<b>Temperature on Receipt (°C)</b>	12.7
<b>Cooling Method</b>	Ice Pack
<b>Sampling Date Provided</b>	YES

### Comments

Nil

Please direct any queries to:

#### Aileen Hie

**Phone:** 02 9910 6200  
**Fax:** 02 9910 6201  
**Email:** ahie@envirolab.com.au

#### Jacinta Hurst

**Phone:** 02 9910 6200  
**Fax:** 02 9910 6201  
**Email:** jhurst@envirolab.com.au

*Analysis Underway, details on the following page:*

Sample ID	VOCs in soil	VTRH(C6-C10)/BTEXN in Soil	svTRH (C10-C40) in Soil	PAHs in Soil	Organochlorine Pesticides in soil	Organophosphorus Pesticides in Soil	PCBs in Soil	Acid Extractable metals in soil	Misc Soil - Inorg	Asbestos ID - soils	Misc Inorg - Soil	CEC
BH101-0.01		✓	✓	✓	✓	✓	✓	✓	✓	✓		
BH102-0.5	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
BH103-1.0	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓		
BH104-0.5		✓	✓	✓	✓	✓	✓	✓	✓	✓		
BH104-1.5		✓	✓	✓				✓		✓	✓	✓
BH104-3.0		✓	✓	✓				✓		✓		
BH104-4.0		✓	✓	✓				✓		✓		
BH108-0.1-0.2		✓	✓	✓	✓	✓	✓	✓	✓	✓		
BH109-0.15-0.2		✓	✓	✓	✓	✓	✓	✓	✓	✓		
BD2_20191022		✓	✓					✓				
BD3_20191023		✓	✓					✓				
TS		✓										
TB		✓										

The '✓' indicates the testing you have requested. **THIS IS NOT A REPORT OF THE RESULTS.**

### Additional Info

Sample storage - Waters are routinely disposed of approximately 1 month and soils approximately 2 months from receipt.

Requests for longer term sample storage must be received in writing.

Please contact the laboratory immediately if observed settled sediment present in water samples is to be included in the extraction and/or analysis (exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS analysis where solids are included by default.

TAT for Micro is dependent on incubation. This varies from 3 to 6 days.

## **CERTIFICATE OF ANALYSIS 229301-A**

### **Client Details**

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker
<b>Address</b>	96 Hermitage Rd, West Ryde, NSW, 2114

### **Sample Details**

<b>Your Reference</b>	<b><u>86874.01, Ultimo</u></b>
<b>Number of Samples</b>	13 Soil
<b>Date samples received</b>	25/10/2019
<b>Date completed instructions received</b>	01/11/2019

### **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.  
Samples were analysed as received from the client. Results relate specifically to the samples as received.  
Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

### **Report Details**

<b>Date results requested by</b>	08/11/2019
<b>Date of Issue</b>	05/11/2019
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Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	

#### **Results Approved By**

Josh Williams, Senior Chemist

#### **Authorised By**



Nancy Zhang, Laboratory Manager

sTPH in Soil (C10-C40)-Silica		
Our Reference		229301-A-2
Your Reference	UNITS	BH102
Depth		0.5
Date Sampled		22/10/2019
Type of sample		Soil
Date extracted	-	04/11/2019
Date analysed	-	05/11/2019
TPH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50
TPH C <sub>15</sub> - C <sub>28</sub>	mg/kg	260
TPH C <sub>29</sub> - C <sub>36</sub>	mg/kg	240
TPH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	<50
TPH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	440
TPH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	180
Surrogate o-Terphenyl	%	90



Method ID	Methodology Summary
Org-003	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.

QUALITY CONTROL: sTPH in Soil (C10-C40)-Silica					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	[NT]
Date extracted	-			04/11/2019	[NT]	[NT]	[NT]	[NT]	04/11/2019	[NT]
Date analysed	-			05/11/2019	[NT]	[NT]	[NT]	[NT]	05/11/2019	[NT]
TPH C <sub>10</sub> - C <sub>14</sub>	mg/kg	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	111	[NT]
TPH C <sub>15</sub> - C <sub>28</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	130	[NT]
TPH C <sub>29</sub> - C <sub>36</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	121	[NT]
TPH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	111	[NT]
TPH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	130	[NT]
TPH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	121	[NT]
Surrogate o-Terphenyl	%		Org-003	70	[NT]	[NT]	[NT]	[NT]	122	[NT]

## Result Definitions

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<b>&lt;</b>	Less than
<b>&gt;</b>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	

## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the same sample will be re-analysed. When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

## Andrew Fitzsimons

---

**From:** David Walker <David.Walker@douglaspartners.com.au>  
**Sent:** Friday, 1 November 2019 2:03 PM  
**To:** Samplereceipt  
**Subject:** Analysis request

Ref: 229301-A

TAT: Std

Due: 8/11/19

*FA*

**Follow Up Flag:** Follow up  
**Flag Status:** Flagged

Hi

2

Please conduct TRH with silica gel clean up analysis on 229301-2 BH102 0.5m. Standard t/a.

thanks

---

**David Walker** | Associate / Environmental Engineer  
**Douglas Partners Pty Ltd** | ABN 75 053 980 117 | [www.douglaspartners.com.au](http://www.douglaspartners.com.au)  
96 Hermitage Road West Ryde NSW 2114 | PO Box 472 West Ryde NSW 1685  
P: 02 9809 0666 | M: 0407 540 537 | E: [David.Walker@douglaspartners.com.au](mailto:David.Walker@douglaspartners.com.au)

FINANCIAL REVIEW

**CLIENT CHOICE AWARDS 2019**

**WINNER**

**beaton**



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## SAMPLE RECEIPT ADVICE

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker

### Sample Login Details

<b>Your reference</b>	86874.01, Ultimo
<b>Envirolab Reference</b>	229301-A
<b>Date Sample Received</b>	25/10/2019
<b>Date Instructions Received</b>	01/11/2019
<b>Date Results Expected to be Reported</b>	08/11/2019

### Sample Condition

<b>Samples received in appropriate condition for analysis</b>	Yes
<b>No. of Samples Provided</b>	13 Soil
<b>Turnaround Time Requested</b>	Standard
<b>Temperature on Receipt (°C)</b>	12.7
<b>Cooling Method</b>	Ice Pack
<b>Sampling Date Provided</b>	YES

### Comments

Nil

Please direct any queries to:

#### Aileen Hie

**Phone:** 02 9910 6200  
**Fax:** 02 9910 6201  
**Email:** ahie@envirolab.com.au

#### Jacinta Hurst

**Phone:** 02 9910 6200  
**Fax:** 02 9910 6201  
**Email:** jhurst@envirolab.com.au

*Analysis Underway, details on the following page:*

**Envirolab Services Pty Ltd**

ABN 37 112 535 645

12 Ashley St Chatswood NSW 2067

ph 02 9910 6200 fax 02 9910 6201

customerservice@envirolab.com.au

www.envirolab.com.au

Sample ID	sTPH in Soil (C10-C40)-Silica	On Hold
BH101-0.1		✓
BH102-0.5	✓	
BH103-1.0		✓
BH104-0.5		✓
BH104-1.5		✓
BH104-3.0		✓
BH104-4.0		✓
BH108-0.1-0.2		✓
BH109-0.15-0.2		✓
BD2_20191022		✓
BD3_20191023		✓
TS		✓
TB		✓

The '✓' indicates the testing you have requested. **THIS IS NOT A REPORT OF THE RESULTS.**

**Additional Info**

Sample storage - Waters are routinely disposed of approximately 1 month and soils approximately 2 months from receipt.

Requests for longer term sample storage must be received in writing.

Please contact the laboratory immediately if observed settled sediment present in water samples is to be included in the extraction and/or analysis (exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS analysis where solids are included by default.

TAT for Micro is dependent on incubation. This varies from 3 to 6 days.



## CERTIFICATE OF ANALYSIS 229472

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker
<b>Address</b>	96 Hermitage Rd, West Ryde, NSW, 2114

### Sample Details

<b>Your Reference</b>	<b>86874.01</b>
<b>Number of Samples</b>	6 Soil
<b>Date samples received</b>	29/10/2019
<b>Date completed instructions received</b>	29/10/2019

### Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.  
 Samples were analysed as received from the client. Results relate specifically to the samples as received.  
 Results are reported on a dry weight basis for solids and on an as received basis for other matrices.  
**Please refer to the last page of this report for any comments relating to the results.**

### Report Details

<b>Date results requested by</b>	05/11/2019
<b>Date of Issue</b>	04/11/2019
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Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	


#### Asbestos Approved By

Analysed by Asbestos Approved Identifier: Aida Marner  
 Authorised by Asbestos Approved Signatory: Lucy Zhu

#### Results Approved By

Diego Bigolin, Team Leader, Inorganics  
 Dragana Tomas, Senior Chemist  
 Jaimie Loa-Kum-Cheung, Metals Supervisor  
 Josh Williams, Chemist  
 Loren Bardwell, Senior Chemist  
 Lucy Zhu, Senior Asbestos Analyst  
 Priya Samarawickrama, Senior Chemist  
 Steven Luong, Organics Supervisor

#### Authorised By



Nancy Zhang, Laboratory Manager

VOCs in soil		
Our Reference		229472-3
Your Reference	UNITS	BH105
Depth		3.0
Date Sampled		28/10/2019
Type of sample		Soil
Date extracted	-	31/10/2019
Date analysed	-	01/11/2019
Dichlorodifluoromethane	mg/kg	<1
Chloromethane	mg/kg	<1
Vinyl Chloride	mg/kg	<1
Bromomethane	mg/kg	<1
Chloroethane	mg/kg	<1
Trichlorofluoromethane	mg/kg	<1
1,1-Dichloroethene	mg/kg	<1
trans-1,2-dichloroethene	mg/kg	<1
1,1-dichloroethane	mg/kg	<1
cis-1,2-dichloroethene	mg/kg	<1
bromochloromethane	mg/kg	<1
chloroform	mg/kg	<1
2,2-dichloropropane	mg/kg	<1
1,2-dichloroethane	mg/kg	<1
1,1,1-trichloroethane	mg/kg	<1
1,1-dichloropropene	mg/kg	<1
Cyclohexane	mg/kg	<1
carbon tetrachloride	mg/kg	<1
Benzene	mg/kg	<0.2
dibromomethane	mg/kg	<1
1,2-dichloropropane	mg/kg	<1
trichloroethene	mg/kg	<1
bromodichloromethane	mg/kg	<1
trans-1,3-dichloropropene	mg/kg	<1
cis-1,3-dichloropropene	mg/kg	<1
1,1,2-trichloroethane	mg/kg	<1
Toluene	mg/kg	<0.5
1,3-dichloropropane	mg/kg	<1
dibromochloromethane	mg/kg	<1
1,2-dibromoethane	mg/kg	<1
tetrachloroethene	mg/kg	<1
1,1,1,2-tetrachloroethane	mg/kg	<1
chlorobenzene	mg/kg	<1

VOCs in soil		
Our Reference		229472-3
Your Reference	UNITS	BH105
Depth		3.0
Date Sampled		28/10/2019
Type of sample		Soil
Ethylbenzene	mg/kg	<1
bromoform	mg/kg	<1
m+p-xylene	mg/kg	<2
styrene	mg/kg	<1
1,1,2,2-tetrachloroethane	mg/kg	<1
o-Xylene	mg/kg	<1
1,2,3-trichloropropane	mg/kg	<1
isopropylbenzene	mg/kg	<1
bromobenzene	mg/kg	<1
n-propyl benzene	mg/kg	<1
2-chlorotoluene	mg/kg	<1
4-chlorotoluene	mg/kg	<1
1,3,5-trimethyl benzene	mg/kg	<1
tert-butyl benzene	mg/kg	<1
1,2,4-trimethyl benzene	mg/kg	<1
1,3-dichlorobenzene	mg/kg	<1
sec-butyl benzene	mg/kg	<1
1,4-dichlorobenzene	mg/kg	<1
4-isopropyl toluene	mg/kg	<1
1,2-dichlorobenzene	mg/kg	<1
n-butyl benzene	mg/kg	<1
1,2-dibromo-3-chloropropane	mg/kg	<1
1,2,4-trichlorobenzene	mg/kg	<1
hexachlorobutadiene	mg/kg	<1
1,2,3-trichlorobenzene	mg/kg	<1
Surrogate Dibromofluorometha	%	107
Surrogate aaa-Trifluorotoluene	%	92
Surrogate Toluene-d <sub>8</sub>	%	109
Surrogate 4-Bromofluorobenzene	%	118

vTRH(C6-C10)/BTEXN in Soil						
Our Reference		229472-1	229472-2	229472-3	229472-4	229472-5
Your Reference	UNITS	BH105	BH105	BH105	BH106	BH106
Depth		1.0	2.0	3.0	0.1	1.0
Date Sampled		28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	31/10/2019	31/10/2019	31/10/2019	31/10/2019	31/10/2019
Date analysed	-	01/11/2019	01/11/2019	01/11/2019	01/11/2019	01/11/2019
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	<25	<25	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	<25	<25	<25
vTPH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	<25	<25	<25
Benzene	mg/kg	<0.2	<0.2	<0.2	<0.2	<0.2
Toluene	mg/kg	<0.5	<0.5	<0.5	<0.5	<0.5
Ethylbenzene	mg/kg	<1	<1	<1	<1	<1
m+p-xylene	mg/kg	<2	<2	<2	<2	<2
o-Xylene	mg/kg	<1	<1	<1	<1	<1
naphthalene	mg/kg	<1	2	4	<1	<1
Total +ve Xylenes	mg/kg	<3	<3	<3	<3	<3
Surrogate aaa-Trifluorotoluene	%	85	121	92	85	96

vTRH(C6-C10)/BTEXN in Soil		
Our Reference		229472-6
Your Reference	UNITS	BH106
Depth		2.0
Date Sampled		28/10/2019
Type of sample		Soil
Date extracted	-	31/10/2019
Date analysed	-	01/11/2019
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25
vTPH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25
Benzene	mg/kg	<0.2
Toluene	mg/kg	<0.5
Ethylbenzene	mg/kg	<1
m+p-xylene	mg/kg	<2
o-Xylene	mg/kg	<1
naphthalene	mg/kg	<1
Total +ve Xylenes	mg/kg	<3
Surrogate aaa-Trifluorotoluene	%	123

## svTRH (C10-C40) in Soil

Our Reference		229472-1	229472-2	229472-3	229472-4	229472-5
Your Reference	UNITS	BH105	BH105	BH105	BH106	BH106
Depth		1.0	2.0	3.0	0.1	1.0
Date Sampled		28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	30/10/2019	30/10/2019	30/10/2019	30/10/2019	30/10/2019
Date analysed	-	31/10/2019	31/10/2019	31/10/2019	31/10/2019	31/10/2019
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	240	210	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	110	210	170	<100	<100
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	<50	<50	<50	<50
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	160	390	340	<100	100
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	140	190	150	100	<100
Total +ve TRH (>C10-C40)	mg/kg	300	570	480	100	100
Surrogate o-Terphenyl	%	75	77	76	78	76

## svTRH (C10-C40) in Soil

Our Reference		229472-6
Your Reference	UNITS	BH106
Depth		2.0
Date Sampled		28/10/2019
Type of sample		Soil
Date extracted	-	30/10/2019
Date analysed	-	31/10/2019
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	<100
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	<100
Total +ve TRH (>C10-C40)	mg/kg	<50
Surrogate o-Terphenyl	%	81

PAHs in Soil						
Our Reference		229472-1	229472-2	229472-3	229472-4	229472-5
Your Reference	UNITS	BH105	BH105	BH105	BH106	BH106
Depth		1.0	2.0	3.0	0.1	1.0
Date Sampled		28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date extracted	-	30/10/2019	30/10/2019	30/10/2019	30/10/2019	30/10/2019
Date analysed	-	31/10/2019	31/10/2019	31/10/2019	31/10/2019	31/10/2019
Naphthalene	mg/kg	0.2	3.9	1.9	<0.1	<0.1
Acenaphthylene	mg/kg	0.2	0.8	0.6	<0.1	0.1
Acenaphthene	mg/kg	0.1	0.6	0.3	<0.1	<0.1
Fluorene	mg/kg	0.1	1.2	0.6	<0.1	<0.1
Phenanthrene	mg/kg	2.3	17	10	0.1	0.6
Anthracene	mg/kg	0.4	3.2	2.0	<0.1	0.2
Fluoranthene	mg/kg	4.2	20	13	0.3	1.3
Pyrene	mg/kg	4.0	18	12	0.3	1.3
Benzo(a)anthracene	mg/kg	1.6	6.2	4.6	0.2	0.7
Chrysene	mg/kg	1.4	5.5	4.3	0.2	0.7
Benzo(b,j+k)fluoranthene	mg/kg	1	5.5	4.1	<0.2	0.7
Benzo(a)pyrene	mg/kg	2.0	7.9	5.8	0.2	0.91
Indeno(1,2,3-c,d)pyrene	mg/kg	1.2	4.7	3.3	<0.1	0.5
Dibenzo(a,h)anthracene	mg/kg	0.3	1.0	0.9	<0.1	0.2
Benzo(g,h,i)perylene	mg/kg	1.7	6.7	4.6	0.1	0.6
Total +ve PAH's	mg/kg	21	100	67	1.4	7.7
Benzo(a)pyrene TEQ calc (zero)	mg/kg	2.8	11	7.9	<0.5	1.3
Benzo(a)pyrene TEQ calc(half)	mg/kg	2.8	11	7.9	<0.5	1.3
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	2.8	11	7.9	<0.5	1.3
Surrogate <i>p</i> -Terphenyl-d14	%	94	96	96	94	95

PAHs in Soil		
Our Reference		229472-6
Your Reference	UNITS	BH106
Depth		2.0
Date Sampled		28/10/2019
Type of sample		Soil
Date extracted	-	30/10/2019
Date analysed	-	31/10/2019
Naphthalene	mg/kg	0.1
Acenaphthylene	mg/kg	<0.1
Acenaphthene	mg/kg	<0.1
Fluorene	mg/kg	<0.1
Phenanthrene	mg/kg	0.4
Anthracene	mg/kg	<0.1
Fluoranthene	mg/kg	0.4
Pyrene	mg/kg	0.3
Benzo(a)anthracene	mg/kg	0.1
Chrysene	mg/kg	<0.1
Benzo(b,j+k)fluoranthene	mg/kg	<0.2
Benzo(a)pyrene	mg/kg	0.1
Indeno(1,2,3-c,d)pyrene	mg/kg	<0.1
Dibenzo(a,h)anthracene	mg/kg	<0.1
Benzo(g,h,i)perylene	mg/kg	<0.1
Total +ve PAH's	mg/kg	1.4
Benzo(a)pyrene TEQ calc (zero)	mg/kg	<0.5
Benzo(a)pyrene TEQ calc(half)	mg/kg	<0.5
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	<0.5
Surrogate <i>p</i> -Terphenyl-d14	%	98

Organochlorine Pesticides in soil			
Our Reference		229472-1	229472-4
Your Reference	UNITS	BH105	BH106
Depth		1.0	0.1
Date Sampled		28/10/2019	28/10/2019
Type of sample		Soil	Soil
Date extracted	-	30/10/2019	30/10/2019
Date analysed	-	31/10/2019	31/10/2019
alpha-BHC	mg/kg	<0.1	<0.1
HCB	mg/kg	<0.1	<0.1
beta-BHC	mg/kg	<0.1	<0.1
gamma-BHC	mg/kg	<0.1	<0.1
Heptachlor	mg/kg	<0.1	<0.1
delta-BHC	mg/kg	<0.1	<0.1
Aldrin	mg/kg	<0.1	<0.1
Heptachlor Epoxide	mg/kg	<0.1	<0.1
gamma-Chlordane	mg/kg	<0.1	<0.1
alpha-chlordane	mg/kg	<0.1	<0.1
Endosulfan I	mg/kg	<0.1	<0.1
pp-DDE	mg/kg	<0.1	<0.1
Dieldrin	mg/kg	<0.1	<0.1
Endrin	mg/kg	<0.1	<0.1
Endosulfan II	mg/kg	<0.1	<0.1
pp-DDD	mg/kg	<0.1	<0.1
Endrin Aldehyde	mg/kg	<0.1	<0.1
pp-DDT	mg/kg	<0.1	<0.1
Endosulfan Sulphate	mg/kg	<0.1	<0.1
Methoxychlor	mg/kg	<0.1	<0.1
Total +ve DDT+DDD+DDE	mg/kg	<0.1	<0.1
Surrogate TCMX	%	93	93



Organophosphorus Pesticides in Soil			
Our Reference		229472-1	229472-4
Your Reference	UNITS	BH105	BH106
Depth		1.0	0.1
Date Sampled		28/10/2019	28/10/2019
Type of sample		Soil	Soil
Date extracted	-	30/10/2019	30/10/2019
Date analysed	-	31/10/2019	31/10/2019
Dichlorvos	mg/kg	<0.1	<0.1
Dimethoate	mg/kg	<0.1	<0.1
Diazinon	mg/kg	<0.1	<0.1
Chlorpyrifos-methyl	mg/kg	<0.1	<0.1
Ronnel	mg/kg	<0.1	<0.1
Fenitrothion	mg/kg	<0.1	<0.1
Malathion	mg/kg	<0.1	<0.1
Chlorpyrifos	mg/kg	<0.1	<0.1
Parathion	mg/kg	<0.1	<0.1
Bromophos-ethyl	mg/kg	<0.1	<0.1
Ethion	mg/kg	<0.1	<0.1
Azinphos-methyl (Guthion)	mg/kg	<0.1	<0.1
Surrogate TCMX	%	93	93

PCBs in Soil			
Our Reference		229472-1	229472-4
Your Reference	UNITS	BH105	BH106
Depth		1.0	0.1
Date Sampled		28/10/2019	28/10/2019
Type of sample		Soil	Soil
Date extracted	-	30/10/2019	30/10/2019
Date analysed	-	31/10/2019	31/10/2019
Aroclor 1016	mg/kg	<0.1	<0.1
Aroclor 1221	mg/kg	<0.1	<0.1
Aroclor 1232	mg/kg	<0.1	<0.1
Aroclor 1242	mg/kg	<0.1	<0.1
Aroclor 1248	mg/kg	<0.1	<0.1
Aroclor 1254	mg/kg	<0.1	<0.1
Aroclor 1260	mg/kg	<0.1	<0.1
Total +ve PCBs (1016-1260)	mg/kg	<0.1	<0.1
Surrogate TCMX	%	93	93

## Acid Extractable metals in soil

Our Reference		229472-1	229472-2	229472-3	229472-4	229472-5
Your Reference	UNITS	BH105	BH105	BH105	BH106	BH106
Depth		1.0	2.0	3.0	0.1	1.0
Date Sampled		28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	30/10/2019	30/10/2019	30/10/2019	30/10/2019	30/10/2019
Date analysed	-	30/10/2019	30/10/2019	30/10/2019	30/10/2019	30/10/2019
Arsenic	mg/kg	8	18	10	<4	7
Cadmium	mg/kg	<0.4	0.6	<0.4	<0.4	<0.4
Chromium	mg/kg	37	16	12	11	15
Copper	mg/kg	42	51	45	44	45
Lead	mg/kg	74	130	95	15	99
Mercury	mg/kg	0.2	0.4	0.4	0.1	0.4
Nickel	mg/kg	17	28	41	86	29
Zinc	mg/kg	87	160	150	41	120

## Acid Extractable metals in soil

Our Reference		229472-6
Your Reference	UNITS	BH106
Depth		2.0
Date Sampled		28/10/2019
Type of sample		Soil
Date prepared	-	30/10/2019
Date analysed	-	30/10/2019
Arsenic	mg/kg	<4
Cadmium	mg/kg	<0.4
Chromium	mg/kg	14
Copper	mg/kg	3
Lead	mg/kg	7
Mercury	mg/kg	<0.1
Nickel	mg/kg	3
Zinc	mg/kg	8

Misc Soil - Inorg			
Our Reference		229472-1	229472-4
Your Reference	UNITS	BH105	BH106
Depth		1.0	0.1
Date Sampled		28/10/2019	28/10/2019
Type of sample		Soil	Soil
Date prepared	-	30/10/2019	30/10/2019
Date analysed	-	30/10/2019	30/10/2019
Total Phenolics (as Phenol)	mg/kg	<5	<5

Misc Inorg - Soil		
Our Reference		229472-5
Your Reference	UNITS	BH106
Depth		1.0
Date Sampled		28/10/2019
Type of sample		Soil
Date prepared	-	30/10/2019
Date analysed	-	30/10/2019
pH 1:5 soil:water	pH Units	9.0

CEC		
Our Reference		229472-5
Your Reference	UNITS	BH106
Depth		1.0
Date Sampled		28/10/2019
Type of sample		Soil
Date prepared	-	31/10/2019
Date analysed	-	31/10/2019
Exchangeable Ca	meq/100g	11
Exchangeable K	meq/100g	0.1
Exchangeable Mg	meq/100g	0.98
Exchangeable Na	meq/100g	<0.1
Cation Exchange Capacity	meq/100g	12

Moisture						
Our Reference		229472-1	229472-2	229472-3	229472-4	229472-5
Your Reference	UNITS	BH105	BH105	BH105	BH106	BH106
Depth		1.0	2.0	3.0	0.1	1.0
Date Sampled		28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date prepared	-	30/10/2019	30/10/2019	30/10/2019	30/10/2019	30/10/2019
Date analysed	-	31/10/2019	31/10/2019	31/10/2019	31/10/2019	31/10/2019
Moisture	%	9.0	9.4	9.7	7.1	6.7

Moisture		
Our Reference		229472-6
Your Reference	UNITS	BH106
Depth		2.0
Date Sampled		28/10/2019
Type of sample		Soil
Date prepared	-	30/10/2019
Date analysed	-	31/10/2019
Moisture	%	15

Asbestos ID - soils						
Our Reference	UNITS	229472-1	229472-2	229472-3	229472-4	229472-5
Your Reference		BH105	BH105	BH105	BH106	BH106
Depth		1.0	2.0	3.0	0.1	1.0
Date Sampled		28/10/2019	28/10/2019	28/10/2019	28/10/2019	28/10/2019
Type of sample		Soil	Soil	Soil	Soil	Soil
Date analysed	-	31/10/2019	31/10/2019	31/10/2019	31/10/2019	31/10/2019
Sample mass tested	g	Approx. 30g	Approx. 35g	Approx. 55g	Approx. 45g	Approx. 55g
Sample Description	-	Brown coarse-grained soil & rocks	Brown coarse-grained soil & rocks	Brown coarse-grained soil & rocks	Brown coarse-grained soil & rocks	Brown coarse-grained soil & rocks
Asbestos ID in soil	-	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected
Asbestos comments	-	NO	NO	NO	NO	NO
Trace Analysis	-	No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected	No asbestos detected



Method ID	Methodology Summary
<b>ASB-001</b>	Asbestos ID - Qualitative identification of asbestos in bulk samples using Polarised Light Microscopy and Dispersion Staining Techniques including Synthetic Mineral Fibre and Organic Fibre as per Australian Standard 4964-2004.
<b>AT-008</b>	Determination of VOCs sampled onto coconut shell charcoal sorbent tubes, that can be desorbed using carbon disulphide, and analysed by GC-MS.
<b>Inorg-001</b>	pH - Measured using pH meter and electrode in accordance with APHA latest edition, 4500-H+. Please note that the results for water analyses are indicative only, as analysis outside of the APHA storage times.
<b>Inorg-008</b>	Moisture content determined by heating at 105+/-5 °C for a minimum of 12 hours.
<b>Inorg-031</b>	Total Phenolics by segmented flow analyser (in line distillation with colourimetric finish). Solids are extracted in a caustic media prior to analysis.
<b>Metals-009</b>	Determination of exchangeable cations and cation exchange capacity in soils using 1M Ammonium Chloride exchange and ICP-AES analytical finish.
<b>Metals-020</b>	Determination of various metals by ICP-AES.
<b>Metals-021</b>	Determination of Mercury by Cold Vapour AAS.
<b>Org-003</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.
<b>Org-003</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID.  F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.  Note, the Total +ve TRH PQL is reflective of the lowest individual PQL and is therefore "Total +ve TRH" is simply a sum of the positive individual TRH fractions (>C10-C40).
<b>Org-006</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD.
<b>Org-006</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD. Note, the Total +ve PCBs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PCBs" is simply a sum of the positive individual PCBs.
<b>Org-012/017</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS.

Method ID	Methodology Summary
<b>Org-012/017</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-MS and/or GC-MS/MS.  Note, the Total +ve reported DDD+DDE+DDT PQL is reflective of the lowest individual PQL and is therefore simply a sum of the positive individually report DDD+DDE+DDT.
<b>Org-012/017</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013. For soil results:- 1. 'EQ PQL' values are assuming all contributing PAHs reported as <PQL are actually at the PQL. This is the most conservative approach and can give false positive TEQs given that PAHs that contribute to the TEQ calculation may not be present. 2. 'EQ zero' values are assuming all contributing PAHs reported as <PQL are zero. This is the least conservative approach and is more susceptible to false negative TEQs when PAHs that contribute to the TEQ calculation are present but below PQL. 3. 'EQ half PQL' values are assuming all contributing PAHs reported as <PQL are half the stipulated PQL. Hence a mid-point between the most and least conservative approaches above. Note, the Total +ve PAHs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PAHs" is simply a sum of the positive individual PAHs.
<b>Org-014</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS.
<b>Org-016</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.
<b>Org-016</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater. Note, the Total +ve Xylene PQL is reflective of the lowest individual PQL and is therefore "Total +ve Xylenes" is simply a sum of the positive individual Xylenes.

QUALITY CONTROL: VOCs in soil					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date extracted	-			31/10/2019	[NT]	[NT]	[NT]	[NT]	31/10/2019	[NT]
Date analysed	-			01/11/2019	[NT]	[NT]	[NT]	[NT]	01/11/2019	[NT]
Dichlorodifluoromethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chloromethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Vinyl Chloride	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Bromomethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chloroethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Trichlorofluoromethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,1-Dichloroethene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
trans-1,2-dichloroethene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,1-dichloroethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	99	[NT]
cis-1,2-dichloroethene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
bromochloromethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
chloroform	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	90	[NT]
2,2-dichloropropane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2-dichloroethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	91	[NT]
1,1,1-trichloroethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	97	[NT]
1,1-dichloropropene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Cyclohexane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
carbon tetrachloride	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzene	mg/kg	0.2	Org-014	<0.2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
dibromomethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2-dichloropropane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
trichloroethene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	104	[NT]
bromodichloromethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	92	[NT]
trans-1,3-dichloropropene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
cis-1,3-dichloropropene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,1,2-trichloroethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Toluene	mg/kg	0.5	Org-014	<0.5	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,3-dichloropropane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
dibromochloromethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	108	[NT]
1,2-dibromoethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
tetrachloroethene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	117	[NT]
1,1,1,2-tetrachloroethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
chlorobenzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ethylbenzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
bromoform	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
m+p-xylene	mg/kg	2	Org-014	<2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
styrene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,1,2,2-tetrachloroethane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]

QUALITY CONTROL: VOCs in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
o-Xylene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2,3-trichloropropane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
isopropylbenzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
bromobenzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
n-propyl benzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
2-chlorotoluene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
4-chlorotoluene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,3,5-trimethyl benzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
tert-butyl benzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2,4-trimethyl benzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,3-dichlorobenzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
sec-butyl benzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,4-dichlorobenzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
4-isopropyl toluene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2-dichlorobenzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
n-butyl benzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2-dibromo-3-chloropropane	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2,4-trichlorobenzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
hexachlorobutadiene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2,3-trichlorobenzene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate Dibromofluorometha	%		Org-014	108	[NT]	[NT]	[NT]	[NT]	97	[NT]
Surrogate aaa-Trifluorotoluene	%		Org-014	101	[NT]	[NT]	[NT]	[NT]	120	[NT]
Surrogate Toluene-d <sub>8</sub>	%		Org-014	109	[NT]	[NT]	[NT]	[NT]	101	[NT]
Surrogate 4-Bromofluorobenzene	%		Org-014	122	[NT]	[NT]	[NT]	[NT]	123	[NT]

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date extracted	-			31/10/2019	[NT]	[NT]	[NT]	[NT]	31/10/2019	[NT]
Date analysed	-			01/11/2019	[NT]	[NT]	[NT]	[NT]	01/11/2019	[NT]
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	25	Org-016	<25	[NT]	[NT]	[NT]	[NT]	123	[NT]
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	25	Org-016	<25	[NT]	[NT]	[NT]	[NT]	123	[NT]
Benzene	mg/kg	0.2	Org-016	<0.2	[NT]	[NT]	[NT]	[NT]	102	[NT]
Toluene	mg/kg	0.5	Org-016	<0.5	[NT]	[NT]	[NT]	[NT]	110	[NT]
Ethylbenzene	mg/kg	1	Org-016	<1	[NT]	[NT]	[NT]	[NT]	134	[NT]
m+p-xylene	mg/kg	2	Org-016	<2	[NT]	[NT]	[NT]	[NT]	134	[NT]
o-Xylene	mg/kg	1	Org-016	<1	[NT]	[NT]	[NT]	[NT]	132	[NT]
naphthalene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate aaa-Trifluorotoluene	%		Org-016	101	[NT]	[NT]	[NT]	[NT]	120	[NT]

QUALITY CONTROL: svTRH (C10-C40) in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date extracted	-			31/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Date analysed	-			01/11/2019	[NT]	[NT]	[NT]	[NT]	31/10/2019	[NT]
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	119	[NT]
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	103	[NT]
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	108	[NT]
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	119	[NT]
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	103	[NT]
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	108	[NT]
Surrogate o-Terphenyl	%		Org-003	79	[NT]	[NT]	[NT]	[NT]	96	[NT]

QUALITY CONTROL: PAHs in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date extracted	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Date analysed	-			31/10/2019	[NT]	[NT]	[NT]	[NT]	31/10/2019	[NT]
Naphthalene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	108	[NT]
Acenaphthylene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Acenaphthene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Fluorene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	98	[NT]
Phenanthrene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	100	[NT]
Anthracene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Fluoranthene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	98	[NT]
Pyrene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	100	[NT]
Benzo(a)anthracene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chrysene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	102	[NT]
Benzo(b,j+k)fluoranthene	mg/kg	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzo(a)pyrene	mg/kg	0.05	Org-012/017	<0.05	[NT]	[NT]	[NT]	[NT]	100	[NT]
Indeno(1,2,3-c,d)pyrene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Dibenzo(a,h)anthracene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzo(g,h,i)perylene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-012/017	95	[NT]	[NT]	[NT]	[NT]	100	[NT]

QUALITY CONTROL: Organochlorine Pesticides in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date extracted	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Date analysed	-			31/10/2019	[NT]	[NT]	[NT]	[NT]	31/10/2019	[NT]
alpha-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	110	[NT]
HCB	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
beta-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	92	[NT]
gamma-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Heptachlor	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	92	[NT]
delta-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aldrin	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	104	[NT]
Heptachlor Epoxide	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	96	[NT]
gamma-Chlordane	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
alpha-chlordane	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Endosulfan I	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDE	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	100	[NT]
Dieldrin	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	106	[NT]
Endrin	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	90	[NT]
Endosulfan II	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDD	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	96	[NT]
Endrin Aldehyde	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDT	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Endosulfan Sulphate	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	94	[NT]
Methoxychlor	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	97	[NT]	[NT]	[NT]	[NT]	95	[NT]



QUALITY CONTROL: Organophosphorus Pesticides in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date extracted	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Date analysed	-			31/10/2019	[NT]	[NT]	[NT]	[NT]	31/10/2019	[NT]
Dichlorvos	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	91	[NT]
Dimethoate	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Diazinon	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chlorpyrifos-methyl	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ronnel	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	112	[NT]
Fenitrothion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	96	[NT]
Malathion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	67	[NT]
Chlorpyrifos	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	124	[NT]
Parathion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	122	[NT]
Bromophos-ethyl	mg/kg	0.1	AT-008	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ethion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	112	[NT]
Azinphos-methyl (Guthion)	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	97	[NT]	[NT]	[NT]	[NT]	95	[NT]

QUALITY CONTROL: PCBs in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date extracted	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Date analysed	-			31/10/2019	[NT]	[NT]	[NT]	[NT]	31/10/2019	[NT]
Aroclor 1016	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1221	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1232	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1242	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1248	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1254	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	86	[NT]
Aroclor 1260	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-006	97	[NT]	[NT]	[NT]	[NT]	95	[NT]

QUALITY CONTROL: Acid Extractable metals in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-9	[NT]
Date prepared	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Date analysed	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Arsenic	mg/kg	4	Metals-020	<4	[NT]	[NT]	[NT]	[NT]	108	[NT]
Cadmium	mg/kg	0.4	Metals-020	<0.4	[NT]	[NT]	[NT]	[NT]	105	[NT]
Chromium	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	110	[NT]
Copper	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	110	[NT]
Lead	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	114	[NT]
Mercury	mg/kg	0.1	Metals-021	<0.1	[NT]	[NT]	[NT]	[NT]	87	[NT]
Nickel	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	107	[NT]
Zinc	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	105	[NT]

QUALITY CONTROL: Misc Soil - Inorg						Duplicate		Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date prepared	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Date analysed	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Total Phenolics (as Phenol)	mg/kg	5	Inorg-031	<5	[NT]	[NT]	[NT]	[NT]	103	[NT]

QUALITY CONTROL: Misc Inorg - Soil						Duplicate		Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date prepared	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
Date analysed	-			30/10/2019	[NT]	[NT]	[NT]	[NT]	30/10/2019	[NT]
pH 1:5 soil:water	pH Units		Inorg-001	[NT]	[NT]	[NT]	[NT]	[NT]	102	[NT]

QUALITY CONTROL: CEC					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date prepared	-			31/10/2019	[NT]	[NT]	[NT]	[NT]	31/10/2019	[NT]
Date analysed	-			31/10/2019	[NT]	[NT]	[NT]	[NT]	31/10/2019	[NT]
Exchangeable Ca	meq/100g	0.1	Metals-009	<0.1	[NT]	[NT]	[NT]	[NT]	108	[NT]
Exchangeable K	meq/100g	0.1	Metals-009	<0.1	[NT]	[NT]	[NT]	[NT]	110	[NT]
Exchangeable Mg	meq/100g	0.1	Metals-009	<0.1	[NT]	[NT]	[NT]	[NT]	107	[NT]
Exchangeable Na	meq/100g	0.1	Metals-009	<0.1	[NT]	[NT]	[NT]	[NT]	110	[NT]

## Result Definitions

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<b>&lt;</b>	Less than
<b>&gt;</b>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	

## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the same sample will be re-analysed. When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.



## Report Comments

Asbestos: Excessive sample volume was provided for asbestos analysis. A portion of the supplied sample was sub-sampled according to Envirolab procedures. We cannot guarantee that this sub-sample is indicative of the entire sample. Envirolab recommends supplying 40-50g (50mL) of sample in its own container as per AS4964-2004.

Note: Samples 229472-1 to 5 were sub-sampled from bags provided by the client.

M - ENVID/Form COC 02

## SAMPLE RECEIPT ADVICE

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker

### Sample Login Details

<b>Your reference</b>	86874.01
<b>Envirolab Reference</b>	229472
<b>Date Sample Received</b>	29/10/2019
<b>Date Instructions Received</b>	29/10/2019
<b>Date Results Expected to be Reported</b>	05/11/2019

### Sample Condition

<b>Samples received in appropriate condition for analysis</b>	Yes
<b>No. of Samples Provided</b>	6 Soil
<b>Turnaround Time Requested</b>	Standard
<b>Temperature on Receipt (°C)</b>	11.6
<b>Cooling Method</b>	Ice Pack
<b>Sampling Date Provided</b>	YES

### Comments

Nil

Please direct any queries to:

#### Aileen Hie

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**Fax:** 02 9910 6201  
**Email:** ahie@envirolab.com.au

#### Jacinta Hurst

**Phone:** 02 9910 6200  
**Fax:** 02 9910 6201  
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*Analysis Underway, details on the following page:*



**Envirolab Services Pty Ltd**

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Sample ID	VOCs in soil	VTRH(C6-C10)/BTEXN in Soil	svTRH (C10-C40) in Soil	PAHs in Soil	Organochlorine Pesticides in soil	Organophosphorus Pesticides in Soil	PCBs in Soil	Acid Extractable metals in soil	Misc Soil - Inorg	Misc Inorg - Soil	CEC	Asbestos ID - soils
BH105-1.0		✓	✓	✓	✓	✓	✓	✓	✓			✓
BH105-2.0		✓	✓	✓				✓				✓
BH105-3.0	✓	✓	✓	✓				✓				✓
BH106-0.1		✓	✓	✓	✓	✓	✓	✓	✓			✓
BH106-1.0		✓	✓	✓				✓		✓	✓	✓
BH106-2.0		✓	✓	✓				✓				

The '✓' indicates the testing you have requested. **THIS IS NOT A REPORT OF THE RESULTS.**

### Additional Info

Sample storage - Waters are routinely disposed of approximately 1 month and soils approximately 2 months from receipt.

Requests for longer term sample storage must be received in writing.

Please contact the laboratory immediately if observed settled sediment present in water samples is to be included in the extraction and/or analysis (exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS analysis where solids are included by default.

TAT for Micro is dependent on incubation. This varies from 3 to 6 days.

## CERTIFICATE OF ANALYSIS 230944

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker
<b>Address</b>	96 Hermitage Rd, West Ryde, NSW, 2114

### Sample Details

<b>Your Reference</b>	<b>86874.01, Ultimo</b>
<b>Number of Samples</b>	5 Water
<b>Date samples received</b>	15/11/2019
<b>Date completed instructions received</b>	15/11/2019

### Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

### Report Details

<b>Date results requested by</b>	22/11/2019
<b>Date of Issue</b>	22/11/2019
NATA Accreditation Number 2901. This document shall not be reproduced except in full.	
Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	

#### Results Approved By

Diego Bigolin, Team Leader, Inorganics  
 Jaimie Loa-Kum-Cheung, Metals Supervisor  
 Josh Williams, Senior Chemist  
 Loren Bardwell, Senior Chemist  
 Nick Sarlamis, Inorganics Supervisor  
 Steven Luong, Organics Supervisor

#### Authorised By



Nancy Zhang, Laboratory Manager

VOCs in water			
Our Reference		230944-1	230944-2
Your Reference	UNITS	MW105	MW104
Date Sampled		15/11/2019	15/11/2019
Type of sample		Water	Water
Date extracted	-	19/11/2019	19/11/2019
Date analysed	-	20/11/2019	20/11/2019
Dichlorodifluoromethane	µg/L	<10	<10
Chloromethane	µg/L	<10	<10
Vinyl Chloride	µg/L	<10	<10
Bromomethane	µg/L	<10	<10
Chloroethane	µg/L	<10	<10
Trichlorofluoromethane	µg/L	<10	<10
1,1-Dichloroethene	µg/L	<1	<1
Trans-1,2-dichloroethene	µg/L	<1	<1
1,1-dichloroethane	µg/L	<1	<1
Cis-1,2-dichloroethene	µg/L	<1	<1
Bromochloromethane	µg/L	<1	<1
Chloroform	µg/L	<1	<1
2,2-dichloropropane	µg/L	<1	<1
1,2-dichloroethane	µg/L	<1	<1
1,1,1-trichloroethane	µg/L	<1	<1
1,1-dichloropropene	µg/L	<1	<1
Cyclohexane	µg/L	<1	<1
Carbon tetrachloride	µg/L	<1	<1
Benzene	µg/L	<1	<1
Dibromomethane	µg/L	<1	<1
1,2-dichloropropane	µg/L	<1	<1
Trichloroethene	µg/L	<1	<1
Bromodichloromethane	µg/L	<1	<1
trans-1,3-dichloropropene	µg/L	<1	<1
cis-1,3-dichloropropene	µg/L	<1	<1
1,1,2-trichloroethane	µg/L	<1	<1
Toluene	µg/L	<1	<1
1,3-dichloropropane	µg/L	<1	<1
Dibromochloromethane	µg/L	<1	<1
1,2-dibromoethane	µg/L	<1	<1
Tetrachloroethene	µg/L	<1	<1
1,1,1,2-tetrachloroethane	µg/L	<1	<1
Chlorobenzene	µg/L	<1	<1
Ethylbenzene	µg/L	<1	<1

VOCs in water			
Our Reference		230944-1	230944-2
Your Reference	UNITS	MW105	MW104
Date Sampled		15/11/2019	15/11/2019
Type of sample		Water	Water
Bromoform	µg/L	<1	<1
m+p-xylene	µg/L	<2	<2
Styrene	µg/L	<1	<1
1,1,2,2-tetrachloroethane	µg/L	<1	<1
o-xylene	µg/L	<1	<1
1,2,3-trichloropropane	µg/L	<1	<1
Isopropylbenzene	µg/L	<1	<1
Bromobenzene	µg/L	<1	<1
n-propyl benzene	µg/L	<1	<1
2-chlorotoluene	µg/L	<1	<1
4-chlorotoluene	µg/L	<1	<1
1,3,5-trimethyl benzene	µg/L	<1	<1
Tert-butyl benzene	µg/L	<1	<1
1,2,4-trimethyl benzene	µg/L	<1	<1
1,3-dichlorobenzene	µg/L	<1	<1
Sec-butyl benzene	µg/L	<1	<1
1,4-dichlorobenzene	µg/L	<1	<1
4-isopropyl toluene	µg/L	<1	<1
1,2-dichlorobenzene	µg/L	<1	<1
n-butyl benzene	µg/L	<1	<1
1,2-dibromo-3-chloropropane	µg/L	<1	<1
1,2,4-trichlorobenzene	µg/L	<1	<1
Hexachlorobutadiene	µg/L	<1	<1
1,2,3-trichlorobenzene	µg/L	<1	<1
Surrogate Dibromofluoromethane	%	83	91
Surrogate toluene-d8	%	62	66
Surrogate 4-BFB	%	100	106

vTRH(C6-C10)/BTEXN in Water						
Our Reference		230944-1	230944-2	230944-3	230944-4	230944-5
Your Reference	UNITS	MW105	MW104	BD1/20191115	TS	TB
Date Sampled		15/11/2019	15/11/2019	15/11/2019	15/11/2019	15/11/2019
Type of sample		Water	Water	Water	Water	Water
Date extracted	-	19/11/2019	19/11/2019	19/11/2019	19/11/2019	19/11/2019
Date analysed	-	20/11/2019	20/11/2019	20/11/2019	20/11/2019	20/11/2019
TRH C <sub>6</sub> - C <sub>9</sub>	µg/L	<10	<10	<10	[NA]	[NA]
TRH C <sub>6</sub> - C <sub>10</sub>	µg/L	<10	<10	<10	[NA]	[NA]
TRH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	µg/L	<10	<10	<10	[NA]	[NA]
Benzene	µg/L	<1	<1	<1	103%	<1
Toluene	µg/L	<1	<1	<1	108%	<1
Ethylbenzene	µg/L	<1	<1	<1	100%	<1
m+p-xylene	µg/L	<2	<2	<2	95%	<2
o-xylene	µg/L	<1	<1	<1	101%	<1
Naphthalene	µg/L	<1	<1	<1	[NA]	<1
Surrogate Dibromofluoromethane	%	83	91	84	106	105
Surrogate toluene-d8	%	62	66	85	106	103
Surrogate 4-BFB	%	100	106	102	104	105



svTRH (C10-C40) in Water				
Our Reference		230944-1	230944-2	230944-3
Your Reference	UNITS	MW105	MW104	BD1/20191115
Date Sampled		15/11/2019	15/11/2019	15/11/2019
Type of sample		Water	Water	Water
Date extracted	-	18/11/2019	18/11/2019	18/11/2019
Date analysed	-	19/11/2019	19/11/2019	19/11/2019
TRH C <sub>10</sub> - C <sub>14</sub>	µg/L	<50	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	µg/L	<100	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	µg/L	<100	<100	<100
TRH >C <sub>10</sub> - C <sub>16</sub>	µg/L	<50	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	µg/L	<50	<50	<50
TRH >C <sub>16</sub> - C <sub>34</sub>	µg/L	<100	<100	<100
TRH >C <sub>34</sub> - C <sub>40</sub>	µg/L	<100	<100	<100
Surrogate o-Terphenyl	%	92	86	78

PAHs in Water - Low Level			
Our Reference		230944-1	230944-2
Your Reference	UNITS	MW105	MW104
Date Sampled		15/11/2019	15/11/2019
Type of sample		Water	Water
Date extracted	-	18/11/2019	18/11/2019
Date analysed	-	19/11/2019	19/11/2019
Naphthalene	µg/L	<0.2	<0.2
Acenaphthylene	µg/L	<0.1	<0.1
Acenaphthene	µg/L	<0.1	<0.1
Fluorene	µg/L	<0.1	<0.1
Phenanthrene	µg/L	<0.1	<0.1
Anthracene	µg/L	<0.1	<0.1
Fluoranthene	µg/L	<0.1	<0.1
Pyrene	µg/L	<0.1	<0.1
Benzo(a)anthracene	µg/L	<0.1	<0.1
Chrysene	µg/L	<0.1	<0.1
Benzo(b,j+k)fluoranthene	µg/L	<0.2	<0.2
Benzo(a)pyrene	µg/L	<0.1	<0.1
Indeno(1,2,3-c,d)pyrene	µg/L	<0.1	<0.1
Dibenzo(a,h)anthracene	µg/L	<0.1	<0.1
Benzo(g,h,i)perylene	µg/L	<0.1	<0.1
Benzo(a)pyrene TEQ	µg/L	<0.5	<0.5
Total +ve PAH's	µg/L	<0.1	<0.1
Surrogate <i>p</i> -Terphenyl-d14	%	76	77

OCPs in Water - Low Level			
Our Reference		230944-1	230944-2
Your Reference	UNITS	MW105	MW104
Date Sampled		15/11/2019	15/11/2019
Type of sample		Water	Water
Date extracted	-	18/11/2019	18/11/2019
Date analysed	-	19/11/2019	19/11/2019
alpha-BHC	µg/L	<0.01	<0.01
HCB	µg/L	<0.01	<0.01
beta-BHC	µg/L	<0.01	<0.01
gamma-BHC	µg/L	<0.01	<0.01
Heptachlor	µg/L	<0.01	<0.01
delta-BHC	µg/L	<0.01	<0.01
Aldrin	µg/L	<0.01	<0.01
Heptachlor Epoxide	µg/L	<0.01	<0.01
gamma-Chlordane	µg/L	<0.01	<0.01
alpha-Chlordane	µg/L	<0.01	<0.01
Endosulfan I	µg/L	<0.01	<0.01
pp-DDE	µg/L	<0.01	<0.01
Dieldrin	µg/L	<0.01	<0.01
Endrin	µg/L	<0.01	<0.01
Endosulfan II	µg/L	<0.01	<0.01
pp-DDD	µg/L	<0.01	<0.01
Endrin Aldehyde	µg/L	<0.01	<0.01
pp-DDT	µg/L	<0.006	<0.006
Endosulfan Sulphate	µg/L	<0.01	<0.01
Methoxychlor	µg/L	<0.01	<0.01
Surrogate TCMX	%	69	64

OP in water LL ANZECCF/ADWG			
Our Reference		230944-1	230944-2
Your Reference	UNITS	MW105	MW104
Date Sampled		15/11/2019	15/11/2019
Type of sample		Water	Water
Date extracted	-	18/11/2019	18/11/2019
Date analysed	-	19/11/2019	19/11/2019
Dichlorovos	µg/L	<0.2	<0.2
Dimethoate	µg/L	<0.15	<0.15
Diazinon	µg/L	<0.01	<0.01
Chlorpyrifos-methyl	µg/L	<0.2	<0.2
Methyl Parathion	µg/L	<0.2	<0.2
Ronnel	µg/L	<0.2	<0.2
Fenitrothion	µg/L	<0.2	<0.2
Malathion	µg/L	<0.05	<0.05
Chlorpyrifos	µg/L	<0.01	<0.01
Parathion	µg/L	<0.01	<0.01
Bromophos ethyl	µg/L	<0.2	<0.2
Ethion	µg/L	<0.2	<0.2
Azinphos-methyl (Guthion)	µg/L	<0.02	<0.02
Surrogate TCMX	%	69	64

PCBs in Water - Low Level			
Our Reference		230944-1	230944-2
Your Reference	UNITS	MW105	MW104
Date Sampled		15/11/2019	15/11/2019
Type of sample		Water	Water
Date extracted	-	18/11/2019	18/11/2019
Date analysed	-	19/11/2019	19/11/2019
Aroclor 1016	µg/L	<0.1	<0.1
Aroclor 1221	µg/L	<0.1	<0.1
Aroclor 1232	µg/L	<0.1	<0.1
Aroclor 1242	µg/L	<0.1	<0.1
Aroclor 1248	µg/L	<0.1	<0.1
Aroclor 1254	µg/L	<0.1	<0.1
Aroclor 1260	µg/L	<0.1	<0.1
Surrogate TCMX	%	69	64

Total Phenolics in Water			
Our Reference		230944-1	230944-2
Your Reference	UNITS	MW105	MW104
Date Sampled		15/11/2019	15/11/2019
Type of sample		Water	Water
Date extracted	-	19/11/2019	19/11/2019
Date analysed	-	19/11/2019	19/11/2019
Total Phenolics (as Phenol)	mg/L	<0.05	<0.05

HM in water - dissolved				
Our Reference		230944-1	230944-2	230944-3
Your Reference	UNITS	MW105	MW104	BD1/20191115
Date Sampled		15/11/2019	15/11/2019	15/11/2019
Type of sample		Water	Water	Water
Date prepared	-	19/11/2019	19/11/2019	19/11/2019
Date analysed	-	19/11/2019	19/11/2019	19/11/2019
Arsenic-Dissolved	µg/L	1	<1	<1
Cadmium-Dissolved	µg/L	<0.1	<0.1	<0.1
Chromium-Dissolved	µg/L	2	<1	<1
Copper-Dissolved	µg/L	4	1	2
Lead-Dissolved	µg/L	5	<1	<1
Mercury-Dissolved	µg/L	0.06	<0.05	<0.05
Nickel-Dissolved	µg/L	12	<1	11
Zinc-Dissolved	µg/L	18	11	11

Miscellaneous Inorganics			
Our Reference		230944-1	230944-2
Your Reference	UNITS	MW105	MW104
Date Sampled		15/11/2019	15/11/2019
Type of sample		Water	Water
Date prepared	-	15/11/2019	15/11/2019
Date analysed	-	15/11/2019	15/11/2019
Ammonia as N in water	mg/L	0.22	8.3



Cations in water Dissolved			
Our Reference		230944-1	230944-2
Your Reference	UNITS	MW105	MW104
Date Sampled		15/11/2019	15/11/2019
Type of sample		Water	Water
Date digested	-	19/11/2019	19/11/2019
Date analysed	-	21/11/2019	21/11/2019
Calcium - Dissolved	mg/L	120	190
Magnesium - Dissolved	mg/L	23	48
Hardness	mgCaCO <sub>3</sub> /L	400	680

Method ID	Methodology Summary
<b>AT-008</b>	Determination of VOCs sampled onto coconut shell charcoal sorbent tubes, that can be desorbed using carbon disulphide, and analysed by GC-MS.
<b>Inorg-031</b>	Total Phenolics by segmented flow analyser (in line distillation with colourimetric finish). Solids are extracted in a caustic media prior to analysis.
<b>Inorg-057</b>	Ammonia - determined colourimetrically, based on APHA latest edition 4500-NH3 F. Waters samples are filtered on receipt prior to analysis. Soils are analysed following a KCl extraction.
<b>Metals-020</b>	Determination of various metals by ICP-AES.
<b>Metals-021</b>	Determination of Mercury by Cold Vapour AAS.
<b>Metals-022</b>	Determination of various metals by ICP-MS.
<b>Org-003</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.
<b>Org-006</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD.
<b>Org-012/017</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS.
<b>Org-012/017</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013.
<b>Org-013</b>	Water samples are analysed directly by purge and trap GC-MS.
<b>Org-016</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTEX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.

QUALITY CONTROL: VOCs in water					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W2	[NT]
Date extracted	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			20/11/2019	[NT]	[NT]	[NT]	[NT]	20/11/2019	[NT]
Dichlorodifluoromethane	µg/L	10	Org-013	<10	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chloromethane	µg/L	10	Org-013	<10	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Vinyl Chloride	µg/L	10	Org-013	<10	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Bromomethane	µg/L	10	Org-013	<10	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chloroethane	µg/L	10	Org-013	<10	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Trichlorofluoromethane	µg/L	10	Org-013	<10	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,1-Dichloroethene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Trans-1,2-dichloroethene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,1-dichloroethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	85	[NT]
Cis-1,2-dichloroethene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Bromochloromethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chloroform	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	84	[NT]
2,2-dichloropropane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2-dichloroethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	80	[NT]
1,1,1-trichloroethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	78	[NT]
1,1-dichloropropene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Cyclohexane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Carbon tetrachloride	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Dibromomethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2-dichloropropane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Trichloroethene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	100	[NT]
Bromodichloromethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	93	[NT]
trans-1,3-dichloropropene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
cis-1,3-dichloropropene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,1,2-trichloroethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Toluene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,3-dichloropropane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Dibromochloromethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	75	[NT]
1,2-dibromoethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Tetrachloroethene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	75	[NT]
1,1,1,2-tetrachloroethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chlorobenzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ethylbenzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Bromoform	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
m+p-xylene	µg/L	2	Org-013	<2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Styrene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,1,2,2-tetrachloroethane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]

QUALITY CONTROL: VOCs in water					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W2	[NT]
o-xylene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2,3-trichloropropane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Isopropylbenzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Bromobenzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
n-propyl benzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
2-chlorotoluene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
4-chlorotoluene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,3,5-trimethyl benzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Tert-butyl benzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2,4-trimethyl benzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,3-dichlorobenzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Sec-butyl benzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,4-dichlorobenzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
4-isopropyl toluene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2-dichlorobenzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
n-butyl benzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2-dibromo-3-chloropropane	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2,4-trichlorobenzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Hexachlorobutadiene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
1,2,3-trichlorobenzene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate Dibromofluoromethane	%		Org-013	83	[NT]	[NT]	[NT]	[NT]	74	[NT]
Surrogate toluene-d8	%		Org-013	83	[NT]	[NT]	[NT]	[NT]	81	[NT]
Surrogate 4-BFB	%		Org-013	108	[NT]	[NT]	[NT]	[NT]	97	[NT]

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Water					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W2	[NT]
Date extracted	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			20/11/2019	[NT]	[NT]	[NT]	[NT]	20/11/2019	[NT]
TRH C <sub>6</sub> - C <sub>9</sub>	µg/L	10	Org-016	<10	[NT]	[NT]	[NT]	[NT]	96	[NT]
TRH C <sub>6</sub> - C <sub>10</sub>	µg/L	10	Org-016	<10	[NT]	[NT]	[NT]	[NT]	96	[NT]
Benzene	µg/L	1	Org-016	<1	[NT]	[NT]	[NT]	[NT]	101	[NT]
Toluene	µg/L	1	Org-016	<1	[NT]	[NT]	[NT]	[NT]	82	[NT]
Ethylbenzene	µg/L	1	Org-016	<1	[NT]	[NT]	[NT]	[NT]	97	[NT]
m+p-xylene	µg/L	2	Org-016	<2	[NT]	[NT]	[NT]	[NT]	96	[NT]
o-xylene	µg/L	1	Org-016	<1	[NT]	[NT]	[NT]	[NT]	104	[NT]
Naphthalene	µg/L	1	Org-013	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate Dibromofluoromethane	%		Org-016	83	[NT]	[NT]	[NT]	[NT]	74	[NT]
Surrogate toluene-d8	%		Org-016	83	[NT]	[NT]	[NT]	[NT]	81	[NT]
Surrogate 4-BFB	%		Org-016	108	[NT]	[NT]	[NT]	[NT]	97	[NT]

QUALITY CONTROL: svTRH (C10-C40) in Water					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W2	[NT]
Date extracted	-			18/11/2019	[NT]	[NT]	[NT]	[NT]	18/11/2019	[NT]
Date analysed	-			18/11/2019	[NT]	[NT]	[NT]	[NT]	18/11/2019	[NT]
TRH C <sub>10</sub> - C <sub>14</sub>	µg/L	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	104	[NT]
TRH C <sub>15</sub> - C <sub>28</sub>	µg/L	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	97	[NT]
TRH C <sub>29</sub> - C <sub>36</sub>	µg/L	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	108	[NT]
TRH >C <sub>10</sub> - C <sub>16</sub>	µg/L	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	104	[NT]
TRH >C <sub>16</sub> - C <sub>34</sub>	µg/L	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	97	[NT]
TRH >C <sub>34</sub> - C <sub>40</sub>	µg/L	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	108	[NT]
Surrogate o-Terphenyl	%		Org-003	78	[NT]	[NT]	[NT]	[NT]	111	[NT]

QUALITY CONTROL: PAHs in Water - Low Level					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date extracted	-			18/11/2019	[NT]	[NT]	[NT]	[NT]	18/11/2019	[NT]
Date analysed	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Naphthalene	µg/L	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	116	[NT]
Acenaphthylene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Acenaphthene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Fluorene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	108	[NT]
Phenanthrene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	92	[NT]
Anthracene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Fluoranthene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	82	[NT]
Pyrene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	94	[NT]
Benzo(a)anthracene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chrysene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	80	[NT]
Benzo(b,j+k)fluoranthene	µg/L	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzo(a)pyrene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	86	[NT]
Indeno(1,2,3-c,d)pyrene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Dibenzo(a,h)anthracene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzo(g,h,i)perylene	µg/L	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-012/017	72	[NT]	[NT]	[NT]	[NT]	75	[NT]

QUALITY CONTROL: OCPs in Water - Low Level					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date extracted	-			18/11/2019	[NT]	[NT]	[NT]	[NT]	18/11/2019	[NT]
Date analysed	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
alpha-BHC	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	91	[NT]
HCB	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
beta-BHC	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	88	[NT]
gamma-BHC	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Heptachlor	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	84	[NT]
delta-BHC	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aldrin	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	94	[NT]
Heptachlor Epoxide	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	89	[NT]
gamma-Chlordane	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
alpha-Chlordane	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Endosulfan I	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDE	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	92	[NT]
Dieldrin	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	106	[NT]
Endrin	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	101	[NT]
Endosulfan II	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDD	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	91	[NT]
Endrin Aldehyde	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDT	µg/L	0.006	AT-008	<0.006	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Endosulfan Sulphate	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	98	[NT]
Methoxychlor	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	70	[NT]	[NT]	[NT]	[NT]	75	[NT]



QUALITY CONTROL: OP in water LL ANZECCF/ADWG					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date extracted	-			18/11/2019	[NT]	[NT]	[NT]	[NT]	18/11/2019	[NT]
Date analysed	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Dichlorovos	µg/L	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	120	[NT]
Dimethoate	µg/L	0.15	Org-012/017	<0.15	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Diazinon	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chlorpyrifos-methyl	µg/L	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Methyl Parathion	µg/L	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ronnel	µg/L	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	78	[NT]
Fenitrothion	µg/L	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	94	[NT]
Malathion	µg/L	0.05	Org-012/017	<0.05	[NT]	[NT]	[NT]	[NT]	71	[NT]
Chlorpyrifos	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	108	[NT]
Parathion	µg/L	0.01	Org-012/017	<0.01	[NT]	[NT]	[NT]	[NT]	96	[NT]
Bromophos ethyl	µg/L	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ethion	µg/L	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	124	[NT]
Azinphos-methyl (Guthion)	µg/L	0.02	Org-012/017	<0.02	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	78	[NT]	[NT]	[NT]	[NT]	62	[NT]

QUALITY CONTROL: PCBs in Water - Low Level					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date extracted	-			18/11/2019	[NT]	[NT]	[NT]	[NT]	18/11/2019	[NT]
Date analysed	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Aroclor 1016	µg/L	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1221	µg/L	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1232	µg/L	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1242	µg/L	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1248	µg/L	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1254	µg/L	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	73	[NT]
Aroclor 1260	µg/L	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-006	70	[NT]	[NT]	[NT]	[NT]	75	[NT]

QUALITY CONTROL: Total Phenolics in Water						Duplicate		Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date extracted	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Total Phenolics (as Phenol)	mg/L	0.05	Inorg-031	<0.05	[NT]	[NT]	[NT]	[NT]	98	[NT]

QUALITY CONTROL: HM in water - dissolved						Duplicate		Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W2	230944-2
Date prepared	-			19/11/2019	1	19/11/2019	19/11/2019		19/11/2019	19/11/2019
Date analysed	-			19/11/2019	1	19/11/2019	19/11/2019		19/11/2019	19/11/2019
Arsenic-Dissolved	µg/L	1	Metals-022	<1	1	1	1	0	96	97
Cadmium-Dissolved	µg/L	0.1	Metals-022	<0.1	1	<0.1	<0.1	0	99	104
Chromium-Dissolved	µg/L	1	Metals-022	<1	1	2	2	0	104	101
Copper-Dissolved	µg/L	1	Metals-022	<1	1	4	4	0	105	98
Lead-Dissolved	µg/L	1	Metals-022	<1	1	5	5	0	109	104
Mercury-Dissolved	µg/L	0.05	Metals-021	<0.05	1	0.06	0.06	0	102	95
Nickel-Dissolved	µg/L	1	Metals-022	<1	1	12	12	0	104	100
Zinc-Dissolved	µg/L	1	Metals-022	<1	1	18	18	0	100	96

QUALITY CONTROL: Miscellaneous Inorganics						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date prepared	-			15/11/2019	[NT]	[NT]	[NT]	[NT]	15/11/2019	[NT]
Date analysed	-			15/11/2019	[NT]	[NT]	[NT]	[NT]	15/11/2019	[NT]
Ammonia as N in water	mg/L	0.005	Inorg-057	<0.005	[NT]	[NT]	[NT]	[NT]	95	[NT]

QUALITY CONTROL: Cations in water Dissolved					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-W1	[NT]
Date digested	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			21/11/2019	[NT]	[NT]	[NT]	[NT]	21/11/2019	[NT]
Calcium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	91	[NT]
Magnesium - Dissolved	mg/L	0.5	Metals-020	<0.5	[NT]	[NT]	[NT]	[NT]	93	[NT]

## Result Definitions

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<b>&lt;</b>	Less than
<b>&gt;</b>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	

## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the same sample will be re-analysed. When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.



Rev4/October2016

## SAMPLE RECEIPT ADVICE

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker

### Sample Login Details

<b>Your reference</b>	86874.01, Ultimo
<b>Envirolab Reference</b>	230944
<b>Date Sample Received</b>	15/11/2019
<b>Date Instructions Received</b>	15/11/2019
<b>Date Results Expected to be Reported</b>	22/11/2019

### Sample Condition

<b>Samples received in appropriate condition for analysis</b>	Yes
<b>No. of Samples Provided</b>	5 Water
<b>Turnaround Time Requested</b>	Standard
<b>Temperature on Receipt (°C)</b>	21.5
<b>Cooling Method</b>	Ice Pack
<b>Sampling Date Provided</b>	YES

### Comments

Nil

Please direct any queries to:

#### Aileen Hie

**Phone:** 02 9910 6200  
**Fax:** 02 9910 6201  
**Email:** ahie@envirolab.com.au

#### Jacinta Hurst

**Phone:** 02 9910 6200  
**Fax:** 02 9910 6201  
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*Analysis Underway, details on the following page:*



**Envirolab Services Pty Ltd**

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Sample ID	VOCs in water	VTRH(C6-C10)/BTEXN in Water	svTRH (C10-C40) in Water	PAHs in Water - Low Level	Organochlorine Pesticides in Water	OP Pesticides in Water	PCBs in Water	Total Phenolics in Water	HM in water - dissolved	Ammonia as N in water	Cations in water Dissolved
MW105	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
MW104	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
BD1/20191115		✓	✓						✓		
TS		✓									
TB		✓									

The '✓' indicates the testing you have requested. **THIS IS NOT A REPORT OF THE RESULTS.**

### Additional Info

Sample storage - Waters are routinely disposed of approximately 1 month and soils approximately 2 months from receipt.

Requests for longer term sample storage must be received in writing.

Please contact the laboratory immediately if observed settled sediment present in water samples is to be included in the extraction and/or analysis (exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS analysis where solids are included by default.

TAT for Micro is dependent on incubation. This varies from 3 to 6 days.

## **CERTIFICATE OF ANALYSIS 231002**

### **Client Details**

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker
<b>Address</b>	96 Hermitage Rd, West Ryde, NSW, 2114

### **Sample Details**

<b>Your Reference</b>	<b><u>86874.01, Ultimo</u></b>
<b>Number of Samples</b>	2 soil
<b>Date samples received</b>	18/11/2019
<b>Date completed instructions received</b>	18/11/2019

### **Analysis Details**

Please refer to the following pages for results, methodology summary and quality control data.  
 Samples were analysed as received from the client. Results relate specifically to the samples as received.  
 Results are reported on a dry weight basis for solids and on an as received basis for other matrices.  
**Please refer to the last page of this report for any comments relating to the results.**

### **Report Details**

<b>Date results requested by</b>	25/11/2019
<b>Date of Issue</b>	22/11/2019
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Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	

#### **Asbestos Approved By**

Analysed by Asbestos Approved Identifier: Lucy Zhu  
 Authorised by Asbestos Approved Signatory: Lucy Zhu

#### **Results Approved By**

Diego Bigolin, Team Leader, Inorganics  
 Jaimie Loa-Kum-Cheung, Metals Supervisor  
 Josh Williams, Senior Chemist  
 Lucy Zhu, Senior Asbestos Analyst  
 Steven Luong, Organics Supervisor

#### **Authorised By**



Nancy Zhang, Laboratory Manager

vTRH(C6-C10)/BTEXN in Soil			
Our Reference		231002-1	231002-2
Your Reference	UNITS	201	201
Depth		0.9-1.0	1.4-1.5
Date Sampled		15/11/2019	15/11/2019
Type of sample		soil	soil
Date extracted	-	19/11/2019	19/11/2019
Date analysed	-	20/11/2019	20/11/2019
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25
vTPH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25
Benzene	mg/kg	<0.2	<0.2
Toluene	mg/kg	<0.5	<0.5
Ethylbenzene	mg/kg	<1	<1
m+p-xylene	mg/kg	<2	<2
o-Xylene	mg/kg	<1	<1
naphthalene	mg/kg	<1	<1
Total +ve Xylenes	mg/kg	<3	<3
Surrogate aaa-Trifluorotoluene	%	91	91

svTRH (C10-C40) in Soil			
Our Reference		231002-1	231002-2
Your Reference	UNITS	201	201
Depth		0.9-1.0	1.4-1.5
Date Sampled		15/11/2019	15/11/2019
Type of sample		soil	soil
Date extracted	-	19/11/2019	19/11/2019
Date analysed	-	20/11/2019	20/11/2019
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	<50	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	<100	<100
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	<100	<100
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	<50	<50
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	<50	<50
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	<100	<100
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	<100	<100
Total +ve TRH (C10-C36)	mg/kg	<50	<50
Total +ve TRH (>C10-C40)	mg/kg	<50	<50
Surrogate o-Terphenyl	%	79	79

PAHs in Soil			
Our Reference		231002-1	231002-2
Your Reference	UNITS	201	201
Depth		0.9-1.0	1.4-1.5
Date Sampled		15/11/2019	15/11/2019
Type of sample		soil	soil
Date extracted	-	19/11/2019	19/11/2019
Date analysed	-	20/11/2019	20/11/2019
Naphthalene	mg/kg	<0.1	<0.1
Acenaphthylene	mg/kg	<0.1	<0.1
Acenaphthene	mg/kg	<0.1	<0.1
Fluorene	mg/kg	<0.1	<0.1
Phenanthrene	mg/kg	0.2	<0.1
Anthracene	mg/kg	<0.1	<0.1
Fluoranthene	mg/kg	0.3	<0.1
Pyrene	mg/kg	0.3	<0.1
Benzo(a)anthracene	mg/kg	0.2	<0.1
Chrysene	mg/kg	0.2	<0.1
Benzo(b,j+k)fluoranthene	mg/kg	0.2	<0.2
Benzo(a)pyrene	mg/kg	0.2	<0.05
Indeno(1,2,3-c,d)pyrene	mg/kg	<0.1	<0.1
Dibenzo(a,h)anthracene	mg/kg	<0.1	<0.1
Benzo(g,h,i)perylene	mg/kg	<0.1	<0.1
Total +ve PAH's	mg/kg	1.5	<0.05
Benzo(a)pyrene TEQ calc (zero)	mg/kg	<0.5	<0.5
Benzo(a)pyrene TEQ calc(half)	mg/kg	<0.5	<0.5
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	<0.5	<0.5
Surrogate <i>p</i> -Terphenyl-d14	%	103	99

Organochlorine Pesticides in soil		
Our Reference		231002-1
Your Reference	UNITS	201
Depth		0.9-1.0
Date Sampled		15/11/2019
Type of sample		soil
Date extracted	-	19/11/2019
Date analysed	-	20/11/2019
alpha-BHC	mg/kg	<0.1
HCB	mg/kg	<0.1
beta-BHC	mg/kg	<0.1
gamma-BHC	mg/kg	<0.1
Heptachlor	mg/kg	<0.1
delta-BHC	mg/kg	<0.1
Aldrin	mg/kg	<0.1
Heptachlor Epoxide	mg/kg	<0.1
gamma-Chlordane	mg/kg	<0.1
alpha-chlordane	mg/kg	<0.1
Endosulfan I	mg/kg	<0.1
pp-DDE	mg/kg	<0.1
Dieldrin	mg/kg	<0.1
Endrin	mg/kg	<0.1
Endosulfan II	mg/kg	<0.1
pp-DDD	mg/kg	<0.1
Endrin Aldehyde	mg/kg	<0.1
pp-DDT	mg/kg	<0.1
Endosulfan Sulphate	mg/kg	<0.1
Methoxychlor	mg/kg	<0.1
Total +ve DDT+DDD+DDE	mg/kg	<0.1
Surrogate TCMX	%	95



Organophosphorus Pesticides in Soil		
Our Reference		231002-1
Your Reference	UNITS	201
Depth		0.9-1.0
Date Sampled		15/11/2019
Type of sample		soil
Date extracted	-	19/11/2019
Date analysed	-	20/11/2019
Dichlorvos	mg/kg	<0.1
Dimethoate	mg/kg	<0.1
Diazinon	mg/kg	<0.1
Chlorpyrifos-methyl	mg/kg	<0.1
Ronnel	mg/kg	<0.1
Fenitrothion	mg/kg	<0.1
Malathion	mg/kg	<0.1
Chlorpyrifos	mg/kg	<0.1
Parathion	mg/kg	<0.1
Bromophos-ethyl	mg/kg	<0.1
Ethion	mg/kg	<0.1
Azinphos-methyl (Guthion)	mg/kg	<0.1
Surrogate TCMX	%	95

PCBs in Soil		
Our Reference		231002-1
Your Reference	UNITS	201
Depth		0.9-1.0
Date Sampled		15/11/2019
Type of sample		soil
Date extracted	-	19/11/2019
Date analysed	-	20/11/2019
Aroclor 1016	mg/kg	<0.1
Aroclor 1221	mg/kg	<0.1
Aroclor 1232	mg/kg	<0.1
Aroclor 1242	mg/kg	<0.1
Aroclor 1248	mg/kg	<0.1
Aroclor 1254	mg/kg	<0.1
Aroclor 1260	mg/kg	<0.1
Total +ve PCBs (1016-1260)	mg/kg	<0.1
Surrogate TCMX	%	95

Acid Extractable metals in soil			
Our Reference		231002-1	231002-2
Your Reference	UNITS	201	201
Depth		0.9-1.0	1.4-1.5
Date Sampled		15/11/2019	15/11/2019
Type of sample		soil	soil
Date prepared	-	19/11/2019	19/11/2019
Date analysed	-	19/11/2019	19/11/2019
Arsenic	mg/kg	<4	9
Cadmium	mg/kg	<0.4	<0.4
Chromium	mg/kg	8	19
Copper	mg/kg	9	54
Lead	mg/kg	25	38
Mercury	mg/kg	0.2	0.1
Nickel	mg/kg	2	5
Zinc	mg/kg	23	100

Misc Soil - Inorg		
Our Reference		231002-1
Your Reference	UNITS	201
Depth		0.9-1.0
Date Sampled		15/11/2019
Type of sample		soil
Date prepared	-	19/11/2019
Date analysed	-	19/11/2019
Total Phenolics (as Phenol)	mg/kg	<5

Moisture			
Our Reference		231002-1	231002-2
Your Reference	UNITS	201	201
Depth		0.9-1.0	1.4-1.5
Date Sampled		15/11/2019	15/11/2019
Type of sample		soil	soil
Date prepared	-	19/11/2019	19/11/2019
Date analysed	-	20/11/2019	20/11/2019
Moisture	%	7.5	3.7

Asbestos ID - soils		
Our Reference		231002-1
Your Reference	UNITS	201
Depth		0.9-1.0
Date Sampled		15/11/2019
Type of sample		soil
Date analysed	-	21/11/2019
Sample mass tested	g	Approx. 50g
Sample Description	-	Brown sandy soil & rocks
Asbestos ID in soil	-	No asbestos detected at reporting limit of 0.1g/kg  Organic fibres detected
Trace Analysis	-	No asbestos detected

Method ID	Methodology Summary
<b>ASB-001</b>	Asbestos ID - Qualitative identification of asbestos in bulk samples using Polarised Light Microscopy and Dispersion Staining Techniques including Synthetic Mineral Fibre and Organic Fibre as per Australian Standard 4964-2004.
<b>AT-008</b>	Determination of VOCs sampled onto coconut shell charcoal sorbent tubes, that can be desorbed using carbon disulphide, and analysed by GC-MS.
<b>Inorg-008</b>	Moisture content determined by heating at 105+/-5 °C for a minimum of 12 hours.
<b>Inorg-031</b>	Total Phenolics by segmented flow analyser (in line distillation with colourimetric finish). Solids are extracted in a caustic media prior to analysis.
<b>Metals-020</b>	Determination of various metals by ICP-AES.
<b>Metals-021</b>	Determination of Mercury by Cold Vapour AAS.
<b>Org-003</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.
<b>Org-003</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID.  F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.  Note, the Total +ve TRH PQL is reflective of the lowest individual PQL and is therefore "Total +ve TRH" is simply a sum of the positive individual TRH fractions (>C10-C40).
<b>Org-006</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD.
<b>Org-006</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD. Note, the Total +ve PCBs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PCBs" is simply a sum of the positive individual PCBs.
<b>Org-012/017</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS.
<b>Org-012/017</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-MS and/or GC-MS/MS.  Note, the Total +ve reported DDD+DDE+DDT PQL is reflective of the lowest individual PQL and is therefore simply a sum of the positive individually report DDD+DDE+DDT.

Method ID	Methodology Summary
<b>Org-012/017</b>	<p>Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013.</p> <p>For soil results:-</p> <ol style="list-style-type: none"> <li>1. 'EQ PQL' values are assuming all contributing PAHs reported as &lt;PQL are actually at the PQL. This is the most conservative approach and can give false positive TEQs given that PAHs that contribute to the TEQ calculation may not be present.</li> <li>2. 'EQ zero' values are assuming all contributing PAHs reported as &lt;PQL are zero. This is the least conservative approach and is more susceptible to false negative TEQs when PAHs that contribute to the TEQ calculation are present but below PQL.</li> <li>3. 'EQ half PQL' values are assuming all contributing PAHs reported as &lt;PQL are half the stipulated PQL. Hence a mid-point between the most and least conservative approaches above.</li> </ol> <p>Note, the Total +ve PAHs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PAHs" is simply a sum of the positive individual PAHs.</p>
<b>Org-014</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS.
<b>Org-016</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.
<b>Org-016</b>	<p>Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.</p> <p>Note, the Total +ve Xylene PQL is reflective of the lowest individual PQL and is therefore "Total +ve Xylenes" is simply a sum of the positive individual Xylenes.</p>



QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Soil					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date extracted	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			20/11/2019	[NT]	[NT]	[NT]	[NT]	20/11/2019	[NT]
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	25	Org-016	<25	[NT]	[NT]	[NT]	[NT]	103	[NT]
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	25	Org-016	<25	[NT]	[NT]	[NT]	[NT]	103	[NT]
Benzene	mg/kg	0.2	Org-016	<0.2	[NT]	[NT]	[NT]	[NT]	97	[NT]
Toluene	mg/kg	0.5	Org-016	<0.5	[NT]	[NT]	[NT]	[NT]	105	[NT]
Ethylbenzene	mg/kg	1	Org-016	<1	[NT]	[NT]	[NT]	[NT]	102	[NT]
m+p-xylene	mg/kg	2	Org-016	<2	[NT]	[NT]	[NT]	[NT]	105	[NT]
o-Xylene	mg/kg	1	Org-016	<1	[NT]	[NT]	[NT]	[NT]	103	[NT]
naphthalene	mg/kg	1	Org-014	<1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate aaa-Trifluorotoluene	%		Org-016	81	[NT]	[NT]	[NT]	[NT]	95	[NT]

QUALITY CONTROL: svTRH (C10-C40) in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-10	[NT]
Date extracted	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	97	[NT]
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	68	[NT]
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	92	[NT]
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	97	[NT]
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	68	[NT]
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	92	[NT]
Surrogate o-Terphenyl	%		Org-003	81	[NT]	[NT]	[NT]	[NT]	130	[NT]

QUALITY CONTROL: PAHs in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-10	[NT]
Date extracted	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			20/11/2019	[NT]	[NT]	[NT]	[NT]	20/11/2019	[NT]
Naphthalene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	116	[NT]
Acenaphthylene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Acenaphthene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Fluorene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	106	[NT]
Phenanthrene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	104	[NT]
Anthracene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Fluoranthene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	102	[NT]
Pyrene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	106	[NT]
Benzo(a)anthracene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chrysene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	106	[NT]
Benzo(b,j+k)fluoranthene	mg/kg	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzo(a)pyrene	mg/kg	0.05	Org-012/017	<0.05	[NT]	[NT]	[NT]	[NT]	102	[NT]
Indeno(1,2,3-c,d)pyrene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Dibenzo(a,h)anthracene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzo(g,h,i)perylene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-012/017	101	[NT]	[NT]	[NT]	[NT]	101	[NT]

QUALITY CONTROL: Organochlorine Pesticides in soil					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-10	[NT]
Date extracted	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			20/11/2019	[NT]	[NT]	[NT]	[NT]	20/11/2019	[NT]
alpha-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	112	[NT]
HCB	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
beta-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	118	[NT]
gamma-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Heptachlor	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	82	[NT]
delta-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aldrin	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	108	[NT]
Heptachlor Epoxide	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	106	[NT]
gamma-Chlordane	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
alpha-chlordane	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Endosulfan I	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDE	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	108	[NT]
Dieldrin	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	106	[NT]
Endrin	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	96	[NT]
Endosulfan II	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDD	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	104	[NT]
Endrin Aldehyde	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDT	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Endosulfan Sulphate	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	102	[NT]
Methoxychlor	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	94	[NT]	[NT]	[NT]	[NT]	91	[NT]

QUALITY CONTROL: Organophosphorus Pesticides in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-10	[NT]
Date extracted	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			20/11/2019	[NT]	[NT]	[NT]	[NT]	20/11/2019	[NT]
Dichlorvos	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	87	[NT]
Dimethoate	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Diazinon	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chlorpyrifos-methyl	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ronnel	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	83	[NT]
Fenitrothion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	85	[NT]
Malathion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	82	[NT]
Chlorpyrifos	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	89	[NT]
Parathion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	103	[NT]
Bromophos-ethyl	mg/kg	0.1	AT-008	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ethion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	85	[NT]
Azinphos-methyl (Guthion)	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	94	[NT]	[NT]	[NT]	[NT]	91	[NT]

QUALITY CONTROL: PCBs in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-10	[NT]
Date extracted	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			20/11/2019	[NT]	[NT]	[NT]	[NT]	20/11/2019	[NT]
Aroclor 1016	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1221	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1232	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1242	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1248	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1254	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	83	[NT]
Aroclor 1260	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-006	94	[NT]	[NT]	[NT]	[NT]	91	[NT]

QUALITY CONTROL: Acid Extractable metals in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-8	[NT]
Date prepared	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Arsenic	mg/kg	4	Metals-020	<4	[NT]	[NT]	[NT]	[NT]	92	[NT]
Cadmium	mg/kg	0.4	Metals-020	<0.4	[NT]	[NT]	[NT]	[NT]	89	[NT]
Chromium	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	98	[NT]
Copper	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	91	[NT]
Lead	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	101	[NT]
Mercury	mg/kg	0.1	Metals-021	<0.1	[NT]	[NT]	[NT]	[NT]	84	[NT]
Nickel	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	88	[NT]
Zinc	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	97	[NT]

QUALITY CONTROL: Misc Soil - Inorg						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-10	[NT]
Date prepared	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Date analysed	-			19/11/2019	[NT]	[NT]	[NT]	[NT]	19/11/2019	[NT]
Total Phenolics (as Phenol)	mg/kg	5	Inorg-031	<5	[NT]	[NT]	[NT]	[NT]	98	[NT]



## Result Definitions

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<b>&lt;</b>	Less than
<b>&gt;</b>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	

## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the same sample will be re-analysed. When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

## Report Comments

Asbestos: A portion of the supplied sample was sub-sampled for asbestos analysis according to Envirolab procedures.

We cannot guarantee that this sub-sample is indicative of the entire sample. Envirolab recommends supplying 40-50g of sample in its own container.

Note: Sample 231002-1 was sub-sampled from a jar provided by the client.

FPM - ENVID/Form COC 02

## SAMPLE RECEIPT ADVICE

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker

### Sample Login Details

<b>Your reference</b>	86874.01, Ultimo
<b>Envirolab Reference</b>	231002
<b>Date Sample Received</b>	18/11/2019
<b>Date Instructions Received</b>	18/11/2019
<b>Date Results Expected to be Reported</b>	25/11/2019

### Sample Condition

<b>Samples received in appropriate condition for analysis</b>	Yes
<b>No. of Samples Provided</b>	2 soil
<b>Turnaround Time Requested</b>	Standard
<b>Temperature on Receipt (°C)</b>	14.1
<b>Cooling Method</b>	Ice Pack
<b>Sampling Date Provided</b>	YES

### Comments

Nil

Please direct any queries to:

#### Aileen Hie

**Phone:** 02 9910 6200  
**Fax:** 02 9910 6201  
**Email:** ahie@envirolab.com.au

#### Jacinta Hurst

**Phone:** 02 9910 6200  
**Fax:** 02 9910 6201  
**Email:** jhurst@envirolab.com.au

*Analysis Underway, details on the following page:*



**Envirolab Services Pty Ltd**

ABN 37 112 535 645

12 Ashley St Chatswood NSW 2067

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customerservice@envirolab.com.au

www.envirolab.com.au

Sample ID	VTRH(C6-C10)/BTEXN in Soil	svTRH (C10-C40) in Soil	PAHs in Soil	Organochlorine Pesticides in soil	Organophosphorus Pesticides in Soil	PCBs in Soil	Acid Extractable metals in soil	Misc Soil - Inorg	Asbestos ID - soils
201-0.9-1.0	✓	✓	✓	✓	✓	✓	✓	✓	✓
201-1.4-1.5	✓	✓	✓				✓		

The '✓' indicates the testing you have requested. **THIS IS NOT A REPORT OF THE RESULTS.**

### Additional Info

Sample storage - Waters are routinely disposed of approximately 1 month and soils approximately 2 months from receipt.

Requests for longer term sample storage must be received in writing.

Please contact the laboratory immediately if observed settled sediment present in water samples is to be included in the extraction and/or analysis (exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS analysis where solids are included by default.

TAT for Micro is dependent on incubation. This varies from 3 to 6 days.

## CERTIFICATE OF ANALYSIS 231640

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker
<b>Address</b>	96 Hermitage Rd, West Ryde, NSW, 2114

### Sample Details

<b>Your Reference</b>	<b>86874.01, Ultimo</b>
<b>Number of Samples</b>	4 soil
<b>Date samples received</b>	26/11/2019
<b>Date completed instructions received</b>	26/11/2019

### Analysis Details

Please refer to the following pages for results, methodology summary and quality control data.

Samples were analysed as received from the client. Results relate specifically to the samples as received.

Results are reported on a dry weight basis for solids and on an as received basis for other matrices.

**Please refer to the last page of this report for any comments relating to the results.**

### Report Details

<b>Date results requested by</b>	29/11/2019
<b>Date of Issue</b>	29/11/2019
<b>Reissue Details</b>	This report replaces R00 created on 29/11/2019 due to: result entry error for Acenaphthylene for sample 1.
NATA Accreditation Number 2901. This document shall not be reproduced except in full.	
Accredited for compliance with ISO/IEC 17025 - Testing. <b>Tests not covered by NATA are denoted with *</b>	

#### Asbestos Approved By

Analysed by Asbestos Approved Identifier: Panika Wongchanda  
 Authorised by Asbestos Approved Signatory: Lucy Zhu

#### Results Approved By

Diego Bigolin, Team Leader, Inorganics  
 Jaimie Loa-Kum-Cheung, Metals Supervisor  
 Lucy Zhu, Senior Asbestos Analyst  
 Steven Luong, Organics Supervisor

#### Authorised By



Nancy Zhang, Laboratory Manager

VOCs in soil		
Our Reference		231640-1
Your Reference	UNITS	202
Depth		1.2-1.3
Date Sampled		26/11/2019
Type of sample		soil
Date extracted	-	27/11/2019
Date analysed	-	28/11/2019
Dichlorodifluoromethane	mg/kg	<1
Chloromethane	mg/kg	<1
Vinyl Chloride	mg/kg	<1
Bromomethane	mg/kg	<1
Chloroethane	mg/kg	<1
Trichlorofluoromethane	mg/kg	<1
1,1-Dichloroethene	mg/kg	<1
trans-1,2-dichloroethene	mg/kg	<1
1,1-dichloroethane	mg/kg	<1
cis-1,2-dichloroethene	mg/kg	<1
bromochloromethane	mg/kg	<1
chloroform	mg/kg	<1
2,2-dichloropropane	mg/kg	<1
1,2-dichloroethane	mg/kg	<1
1,1,1-trichloroethane	mg/kg	<1
1,1-dichloropropene	mg/kg	<1
Cyclohexane	mg/kg	<1
carbon tetrachloride	mg/kg	<1
Benzene	mg/kg	<0.2
dibromomethane	mg/kg	<1
1,2-dichloropropane	mg/kg	<1
trichloroethene	mg/kg	<1
bromodichloromethane	mg/kg	<1
trans-1,3-dichloropropene	mg/kg	<1
cis-1,3-dichloropropene	mg/kg	<1
1,1,2-trichloroethane	mg/kg	<1
Toluene	mg/kg	<0.5
1,3-dichloropropane	mg/kg	<1
dibromochloromethane	mg/kg	<1
1,2-dibromoethane	mg/kg	<1
tetrachloroethene	mg/kg	<1
1,1,1,2-tetrachloroethane	mg/kg	<1
chlorobenzene	mg/kg	<1



VOCs in soil		
Our Reference		231640-1
Your Reference	UNITS	202
Depth		1.2-1.3
Date Sampled		26/11/2019
Type of sample		soil
Ethylbenzene	mg/kg	<1
bromoform	mg/kg	<1
m+p-xylene	mg/kg	<2
styrene	mg/kg	<1
1,1,2,2-tetrachloroethane	mg/kg	<1
o-Xylene	mg/kg	<1
1,2,3-trichloropropane	mg/kg	<1
isopropylbenzene	mg/kg	<1
bromobenzene	mg/kg	<1
n-propyl benzene	mg/kg	<1
2-chlorotoluene	mg/kg	<1
4-chlorotoluene	mg/kg	<1
1,3,5-trimethyl benzene	mg/kg	<1
tert-butyl benzene	mg/kg	<1
1,2,4-trimethyl benzene	mg/kg	<1
1,3-dichlorobenzene	mg/kg	<1
sec-butyl benzene	mg/kg	<1
1,4-dichlorobenzene	mg/kg	<1
4-isopropyl toluene	mg/kg	<1
1,2-dichlorobenzene	mg/kg	<1
n-butyl benzene	mg/kg	<1
1,2-dibromo-3-chloropropane	mg/kg	<1
1,2,4-trichlorobenzene	mg/kg	<1
hexachlorobutadiene	mg/kg	<1
1,2,3-trichlorobenzene	mg/kg	<1
Surrogate Dibromofluorometha	%	96
Surrogate aaa-Trifluorotoluene	%	76
Surrogate Toluene-d <sub>8</sub>	%	96
Surrogate 4-Bromofluorobenzene	%	75

vTRH(C6-C10)/BTEXN in Soil					
Our Reference		231640-1	231640-2	231640-3	231640-4
Your Reference	UNITS	202	202	TS	TB
Depth		1.2-1.3	2.0-2.1	-	-
Date Sampled		26/11/2019	26/11/2019	26/11/2019	26/11/2019
Type of sample		soil	soil	soil	soil
Date extracted	-	27/11/2019	27/11/2019	27/11/2019	27/11/2019
Date analysed	-	28/11/2019	28/11/2019	27/11/2019	27/11/2019
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	<25	<25	[NA]	<25
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	<25	<25	[NA]	<25
vTPH C <sub>6</sub> - C <sub>10</sub> less BTEX (F1)	mg/kg	<25	<25	[NA]	<25
Benzene	mg/kg	<0.2	<0.2	103%	<0.2
Toluene	mg/kg	<0.5	<0.5	103%	<0.5
Ethylbenzene	mg/kg	<1	<1	107%	<1
m+p-xylene	mg/kg	<2	<2	106%	<2
o-Xylene	mg/kg	<1	<1	106%	<1
naphthalene	mg/kg	73	6	[NA]	<1
Total +ve Xylenes	mg/kg	<3	<3	[NA]	<3
Surrogate aaa-Trifluorotoluene	%	76	68	102	84

svTRH (C10-C40) in Soil			
Our Reference		231640-1	231640-2
Your Reference	UNITS	202	202
Depth		1.2-1.3	2.0-2.1
Date Sampled		26/11/2019	26/11/2019
Type of sample		soil	soil
Date extracted	-	27/11/2019	27/11/2019
Date analysed	-	28/11/2019	27/11/2019
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	380	<50
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	6,700	620
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	3,000	270
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	1,100	82
TRH >C <sub>10</sub> - C <sub>16</sub> less Naphthalene (F2)	mg/kg	1,000	76
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	8,500	780
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	1,300	<100
Total +ve TRH (>C10-C40)	mg/kg	11,000	860
Surrogate o-Terphenyl	%	#	83

PAHs in Soil			
Our Reference		231640-1	231640-2
Your Reference	UNITS	202	202
Depth		1.2-1.3	2.0-2.1
Date Sampled		26/11/2019	26/11/2019
Type of sample		soil	soil
Date extracted	-	27/11/2019	27/11/2019
Date analysed	-	28/11/2019	28/11/2019
Naphthalene	mg/kg	100	8.0
Acenaphthylene	mg/kg	2.0	2.5
Acenaphthene	mg/kg	220	12
Fluorene	mg/kg	240	13
Phenanthrene	mg/kg	1,200	83
Anthracene	mg/kg	390	25
Fluoranthene	mg/kg	1,000	71
Pyrene	mg/kg	830	60
Benzo(a)anthracene	mg/kg	430	31
Chrysene	mg/kg	340	22
Benzo(b,j+k)fluoranthene	mg/kg	480	33
Benzo(a)pyrene	mg/kg	290	21
Indeno(1,2,3-c,d)pyrene	mg/kg	73	1.2
Dibenzo(a,h)anthracene	mg/kg	24	3.2
Benzo(g,h,i)perylene	mg/kg	82	12
Total +ve PAH's	mg/kg	5,700	400
Benzo(a)pyrene TEQ calc (zero)	mg/kg	420	31
Benzo(a)pyrene TEQ calc(half)	mg/kg	420	31
Benzo(a)pyrene TEQ calc(PQL)	mg/kg	420	31
Surrogate <i>p</i> -Terphenyl-d14	%	130	110

Organochlorine Pesticides in soil		
Our Reference		231640-1
Your Reference	UNITS	202
Depth		1.2-1.3
Date Sampled		26/11/2019
Type of sample		soil
Date extracted	-	27/11/2019
Date analysed	-	28/11/2019
alpha-BHC	mg/kg	<1
HCB	mg/kg	<1
beta-BHC	mg/kg	<1
gamma-BHC	mg/kg	<1
Heptachlor	mg/kg	<1
delta-BHC	mg/kg	<1
Aldrin	mg/kg	<1
Heptachlor Epoxide	mg/kg	<1
gamma-Chlordane	mg/kg	<1
alpha-chlordane	mg/kg	<1
Endosulfan I	mg/kg	<1
pp-DDE	mg/kg	<1
Dieldrin	mg/kg	<1
Endrin	mg/kg	<1
Endosulfan II	mg/kg	<1
pp-DDD	mg/kg	<1
Endrin Aldehyde	mg/kg	<1
pp-DDT	mg/kg	<1
Endosulfan Sulphate	mg/kg	<1
Methoxychlor	mg/kg	<1
Total +ve DDT+DDD+DDE	mg/kg	<1
Surrogate TCMX	%	110

Organophosphorus Pesticides in Soil		
Our Reference		231640-1
Your Reference	UNITS	202
Depth		1.2-1.3
Date Sampled		26/11/2019
Type of sample		soil
Date extracted	-	27/11/2019
Date analysed	-	28/11/2019
Dichlorvos	mg/kg	<1
Dimethoate	mg/kg	<1
Diazinon	mg/kg	<1
Chlorpyrifos-methyl	mg/kg	<1
Ronnel	mg/kg	<1
Fenitrothion	mg/kg	<1
Malathion	mg/kg	<1
Chlorpyrifos	mg/kg	<1
Parathion	mg/kg	<1
Bromophos-ethyl	mg/kg	<1
Ethion	mg/kg	<1
Azinphos-methyl (Guthion)	mg/kg	<1
Surrogate TCMX	%	110

PCBs in Soil		
Our Reference		231640-1
Your Reference	UNITS	202
Depth		1.2-1.3
Date Sampled		26/11/2019
Type of sample		soil
Date extracted	-	27/11/2019
Date analysed	-	28/11/2019
Aroclor 1016	mg/kg	<1
Aroclor 1221	mg/kg	<1
Aroclor 1232	mg/kg	<1
Aroclor 1242	mg/kg	<1
Aroclor 1248	mg/kg	<1
Aroclor 1254	mg/kg	<1
Aroclor 1260	mg/kg	<1
Total +ve PCBs (1016-1260)	mg/kg	<1
Surrogate TCMX	%	110

Acid Extractable metals in soil			
Our Reference		231640-1	231640-2
Your Reference	UNITS	202	202
Depth		1.2-1.3	2.0-2.1
Date Sampled		26/11/2019	26/11/2019
Type of sample		soil	soil
Date prepared	-	27/11/2019	27/11/2019
Date analysed	-	27/11/2019	27/11/2019
Arsenic	mg/kg	<4	4
Cadmium	mg/kg	<0.4	<0.4
Chromium	mg/kg	45	30
Copper	mg/kg	130	70
Lead	mg/kg	30	320
Mercury	mg/kg	<0.1	1.4
Nickel	mg/kg	35	36
Zinc	mg/kg	96	120



Misc Soil - Inorg		
Our Reference		231640-1
Your Reference	UNITS	202
Depth		1.2-1.3
Date Sampled		26/11/2019
Type of sample		soil
Date prepared	-	27/11/2019
Date analysed	-	27/11/2019
Total Phenolics (as Phenol)	mg/kg	<5

Moisture			
Our Reference		231640-1	231640-2
Your Reference	UNITS	202	202
Depth		1.2-1.3	2.0-2.1
Date Sampled		26/11/2019	26/11/2019
Type of sample		soil	soil
Date prepared	-	27/11/2019	27/11/2019
Date analysed	-	28/11/2019	28/11/2019
Moisture	%	5.3	9.8

Asbestos ID - soils			
Our Reference		231640-1	231640-2
Your Reference	UNITS	202	202
Depth		1.2-1.3	2.0-2.1
Date Sampled		26/11/2019	26/11/2019
Type of sample		soil	soil
Date analysed	-	27/11/2019	27/11/2019
Sample mass tested	g	Approx. 45g	Approx. 25g
Sample Description	-	Brown fine-grained soil & rocks	Brown fine-grained soil & rocks
Asbestos ID in soil	-	No asbestos detected at reporting limit of 0.1g/kg	No asbestos detected at reporting limit of 0.1g/kg
		Organic fibres detected	Organic fibres detected
Trace Analysis	-	No asbestos detected	No asbestos detected

Method ID	Methodology Summary
<b>ASB-001</b>	Asbestos ID - Qualitative identification of asbestos in bulk samples using Polarised Light Microscopy and Dispersion Staining Techniques including Synthetic Mineral Fibre and Organic Fibre as per Australian Standard 4964-2004.
<b>AT-008</b>	Determination of VOCs sampled onto coconut shell charcoal sorbent tubes, that can be desorbed using carbon disulphide, and analysed by GC-MS.
<b>Inorg-008</b>	Moisture content determined by heating at 105+/-5 °C for a minimum of 12 hours.
<b>Inorg-031</b>	Total Phenolics by segmented flow analyser (in line distillation with colourimetric finish). Solids are extracted in a caustic media prior to analysis.
<b>Metals-020</b>	Determination of various metals by ICP-AES.
<b>Metals-021</b>	Determination of Mercury by Cold Vapour AAS.
<b>Org-003</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID. F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.
<b>Org-003</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-FID.  F2 = (>C10-C16)-Naphthalene as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater (HSLs Tables 1A (3, 4)). Note Naphthalene is determined from the VOC analysis.  Note, the Total +ve TRH PQL is reflective of the lowest individual PQL and is therefore "Total +ve TRH" is simply a sum of the positive individual TRH fractions (>C10-C40).
<b>Org-006</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD.
<b>Org-006</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-ECD. Note, the Total +ve PCBs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PCBs" is simply a sum of the positive individual PCBs.
<b>Org-012/017</b>	Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS.
<b>Org-012/017</b>	Soil samples are extracted with dichloromethane/acetone and waters with dichloromethane and analysed by GC-MS and/or GC-MS/MS.  Note, the Total +ve reported DDD+DDE+DDT PQL is reflective of the lowest individual PQL and is therefore simply a sum of the positive individually report DDD+DDE+DDT.

Method ID	Methodology Summary
<b>Org-012/017</b>	<p>Soil samples are extracted with Dichloromethane/Acetone and waters with Dichloromethane and analysed by GC-MS and/or GC-MS/MS. Benzo(a)pyrene TEQ as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater - 2013.</p> <p>For soil results:-</p> <ol style="list-style-type: none"> <li>1. 'EQ PQL' values are assuming all contributing PAHs reported as &lt;PQL are actually at the PQL. This is the most conservative approach and can give false positive TEQs given that PAHs that contribute to the TEQ calculation may not be present.</li> <li>2. 'EQ zero' values are assuming all contributing PAHs reported as &lt;PQL are zero. This is the least conservative approach and is more susceptible to false negative TEQs when PAHs that contribute to the TEQ calculation are present but below PQL.</li> <li>3. 'EQ half PQL' values are assuming all contributing PAHs reported as &lt;PQL are half the stipulated PQL. Hence a mid-point between the most and least conservative approaches above.</li> </ol> <p>Note, the Total +ve PAHs PQL is reflective of the lowest individual PQL and is therefore "Total +ve PAHs" is simply a sum of the positive individual PAHs.</p>
<b>Org-014</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS.
<b>Org-016</b>	Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.
<b>Org-016</b>	<p>Soil samples are extracted with methanol and spiked into water prior to analysing by purge and trap GC-MS. Water samples are analysed directly by purge and trap GC-MS. F1 = (C6-C10)-BTX as per NEPM B1 Guideline on Investigation Levels for Soil and Groundwater.</p> <p>Note, the Total +ve Xylene PQL is reflective of the lowest individual PQL and is therefore "Total +ve Xylenes" is simply a sum of the positive individual Xylenes.</p>

QUALITY CONTROL: VOCs in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	231640-1
Date extracted	-			27/11/2019	1	27/11/2019	27/11/2019		27/11/2019	27/11/2019
Date analysed	-			28/11/2019	1	28/11/2019	28/11/2019		28/11/2019	28/11/2019
Dichlorodifluoromethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Chloromethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Vinyl Chloride	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Bromomethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Chloroethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Trichlorofluoromethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,1-Dichloroethene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
trans-1,2-dichloroethene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,1-dichloroethane	mg/kg	1	Org-014	<1	1	<1	<1	0	86	71
cis-1,2-dichloroethene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
bromochloromethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
chloroform	mg/kg	1	Org-014	<1	1	<1	<1	0	70	60
2,2-dichloropropane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,2-dichloroethane	mg/kg	1	Org-014	<1	1	<1	<1	0	92	77
1,1,1-trichloroethane	mg/kg	1	Org-014	<1	1	<1	<1	0	90	74
1,1-dichloropropene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Cyclohexane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
carbon tetrachloride	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Benzene	mg/kg	0.2	Org-014	<0.2	1	<0.2	<0.2	0	[NT]	[NT]
dibromomethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,2-dichloropropane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
trichloroethene	mg/kg	1	Org-014	<1	1	<1	<1	0	87	74
bromodichloromethane	mg/kg	1	Org-014	<1	1	<1	<1	0	92	80
trans-1,3-dichloropropene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
cis-1,3-dichloropropene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,1,2-trichloroethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Toluene	mg/kg	0.5	Org-014	<0.5	1	<0.5	<0.5	0	[NT]	[NT]
1,3-dichloropropane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
dibromochloromethane	mg/kg	1	Org-014	<1	1	<1	<1	0	88	74
1,2-dibromoethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
tetrachloroethene	mg/kg	1	Org-014	<1	1	<1	<1	0	95	77
1,1,1,2-tetrachloroethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
chlorobenzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Ethylbenzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
bromoform	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
m+p-xylene	mg/kg	2	Org-014	<2	1	<2	<2	0	[NT]	[NT]
styrene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,1,2,2-tetrachloroethane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]

QUALITY CONTROL: VOCs in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	231640-1
o-Xylene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,2,3-trichloropropane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
isopropylbenzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
bromobenzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
n-propyl benzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
2-chlorotoluene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
4-chlorotoluene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,3,5-trimethyl benzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
tert-butyl benzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,2,4-trimethyl benzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,3-dichlorobenzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
sec-butyl benzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,4-dichlorobenzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
4-isopropyl toluene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,2-dichlorobenzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
n-butyl benzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,2-dibromo-3-chloropropane	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,2,4-trichlorobenzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
hexachlorobutadiene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
1,2,3-trichlorobenzene	mg/kg	1	Org-014	<1	1	<1	<1	0	[NT]	[NT]
Surrogate Dibromofluorometha	%		Org-014	94	1	96	89	8	92	78
Surrogate aaa-Trifluorotoluene	%		Org-014	79	1	76	88	15	80	69
Surrogate Toluene-d <sub>8</sub>	%		Org-014	97	1	96	108	12	93	99
Surrogate 4-Bromofluorobenzene	%		Org-014	72	1	75	79	5	88	89

QUALITY CONTROL: vTRH(C6-C10)/BTEXN in Soil						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	231640-1
Date extracted	-			27/11/2019	1	27/11/2019	27/11/2019		27/11/2019	27/11/2019
Date analysed	-			28/11/2019	1	28/11/2019	28/11/2019		28/11/2019	28/11/2019
TRH C <sub>6</sub> - C <sub>9</sub>	mg/kg	25	Org-016	<25	1	<25	<25	0	90	86
TRH C <sub>6</sub> - C <sub>10</sub>	mg/kg	25	Org-016	<25	1	<25	<25	0	90	86
Benzene	mg/kg	0.2	Org-016	<0.2	1	<0.2	<0.2	0	91	81
Toluene	mg/kg	0.5	Org-016	<0.5	1	<0.5	<0.5	0	92	90
Ethylbenzene	mg/kg	1	Org-016	<1	1	<1	<1	0	79	80
m+p-xylene	mg/kg	2	Org-016	<2	1	<2	<2	0	93	89
o-Xylene	mg/kg	1	Org-016	<1	1	<1	<1	0	91	87
naphthalene	mg/kg	1	Org-014	<1	1	73	56	26	[NT]	[NT]
Surrogate aaa-Trifluorotoluene	%		Org-016	79	1	76	88	15	80	69



QUALITY CONTROL: svTRH (C10-C40) in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	231640-1
Date extracted	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	27/11/2019
Date analysed	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	27/11/2019
TRH C <sub>10</sub> - C <sub>14</sub>	mg/kg	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	94	#
TRH C <sub>15</sub> - C <sub>28</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	77	#
TRH C <sub>29</sub> - C <sub>36</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	77	#
TRH >C <sub>10</sub> -C <sub>16</sub>	mg/kg	50	Org-003	<50	[NT]	[NT]	[NT]	[NT]	94	#
TRH >C <sub>16</sub> -C <sub>34</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	77	#
TRH >C <sub>34</sub> -C <sub>40</sub>	mg/kg	100	Org-003	<100	[NT]	[NT]	[NT]	[NT]	77	#
Surrogate o-Terphenyl	%		Org-003	88	[NT]	[NT]	[NT]	[NT]	106	#

QUALITY CONTROL: PAHs in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	231640-1
Date extracted	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	27/11/2019
Date analysed	-			28/11/2019	[NT]	[NT]	[NT]	[NT]	28/11/2019	28/11/2019
Naphthalene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	116	#
Acenaphthylene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Acenaphthene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Fluorene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	114	#
Phenanthrene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	112	#
Anthracene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Fluoranthene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	114	#
Pyrene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	118	#
Benzo(a)anthracene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chrysene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	116	#
Benzo(b,j+k)fluoranthene	mg/kg	0.2	Org-012/017	<0.2	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzo(a)pyrene	mg/kg	0.05	Org-012/017	<0.05	[NT]	[NT]	[NT]	[NT]	114	#
Indeno(1,2,3-c,d)pyrene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Dibenzo(a,h)anthracene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Benzo(g,h,i)perylene	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate p-Terphenyl-d14	%		Org-012/017	97	[NT]	[NT]	[NT]	[NT]	109	130

QUALITY CONTROL: Organochlorine Pesticides in soil					Duplicate				Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	231640-1
Date extracted	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	27/11/2019
Date analysed	-			28/11/2019	[NT]	[NT]	[NT]	[NT]	28/11/2019	28/11/2019
alpha-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	132	#
HCB	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
beta-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	108	#
gamma-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Heptachlor	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	108	120
delta-BHC	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aldrin	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	118	#
Heptachlor Epoxide	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	116	125
gamma-Chlordane	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
alpha-chlordane	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Endosulfan I	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDE	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	120	114
Dieldrin	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	114	114
Endrin	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	98	#
Endosulfan II	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDD	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	118	#
Endrin Aldehyde	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
pp-DDT	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Endosulfan Sulphate	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	120	128
Methoxychlor	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	94	[NT]	[NT]	[NT]	[NT]	105	125

QUALITY CONTROL: Organophosphorus Pesticides in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	231640-1
Date extracted	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	27/11/2019
Date analysed	-			28/11/2019	[NT]	[NT]	[NT]	[NT]	28/11/2019	28/11/2019
Dichlorvos	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	104	106
Dimethoate	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Diazinon	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Chlorpyrifos-methyl	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ronnel	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	90	100
Fenitrothion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	82	107
Malathion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	80	114
Chlorpyrifos	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	106	#
Parathion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	98	100
Bromophos-ethyl	mg/kg	0.1	AT-008	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Ethion	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	92	#
Azinphos-methyl (Guthion)	mg/kg	0.1	Org-012/017	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-012/017	94	[NT]	[NT]	[NT]	[NT]	105	125

QUALITY CONTROL: PCBs in Soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	231640-1
Date extracted	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	27/11/2019
Date analysed	-			28/11/2019	[NT]	[NT]	[NT]	[NT]	28/11/2019	28/11/2019
Aroclor 1016	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1221	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1232	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1242	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1248	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Aroclor 1254	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	114	106
Aroclor 1260	mg/kg	0.1	Org-006	<0.1	[NT]	[NT]	[NT]	[NT]	[NT]	[NT]
Surrogate TCMX	%		Org-006	94	[NT]	[NT]	[NT]	[NT]	105	125

QUALITY CONTROL: Acid Extractable metals in soil					Duplicate			Spike Recovery %		
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	231640-1
Date prepared	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	27/11/2019
Date analysed	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	27/11/2019
Arsenic	mg/kg	4	Metals-020	<4	[NT]	[NT]	[NT]	[NT]	106	84
Cadmium	mg/kg	0.4	Metals-020	<0.4	[NT]	[NT]	[NT]	[NT]	100	79
Chromium	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	110	72
Copper	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	108	92
Lead	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	113	80
Mercury	mg/kg	0.1	Metals-021	<0.1	[NT]	[NT]	[NT]	[NT]	100	83
Nickel	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	101	73
Zinc	mg/kg	1	Metals-020	<1	[NT]	[NT]	[NT]	[NT]	109	#

QUALITY CONTROL: Misc Soil - Inorg						Duplicate			Spike Recovery %	
Test Description	Units	PQL	Method	Blank	#	Base	Dup.	RPD	LCS-1	[NT]
Date prepared	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	[NT]
Date analysed	-			27/11/2019	[NT]	[NT]	[NT]	[NT]	27/11/2019	[NT]
Total Phenolics (as Phenol)	mg/kg	5	Inorg-031	<5	[NT]	[NT]	[NT]	[NT]	103	[NT]

## Result Definitions

<b>NT</b>	Not tested
<b>NA</b>	Test not required
<b>INS</b>	Insufficient sample for this test
<b>PQL</b>	Practical Quantitation Limit
<b>&lt;</b>	Less than
<b>&gt;</b>	Greater than
<b>RPD</b>	Relative Percent Difference
<b>LCS</b>	Laboratory Control Sample
<b>NS</b>	Not specified
<b>NEPM</b>	National Environmental Protection Measure
<b>NR</b>	Not Reported

## Quality Control Definitions

<b>Blank</b>	This is the component of the analytical signal which is not derived from the sample but from reagents, glassware etc, can be determined by processing solvents and reagents in exactly the same manner as for samples.
<b>Duplicate</b>	This is the complete duplicate analysis of a sample from the process batch. If possible, the sample selected should be one where the analyte concentration is easily measurable.
<b>Matrix Spike</b>	A portion of the sample is spiked with a known concentration of target analyte. The purpose of the matrix spike is to monitor the performance of the analytical method used and to determine whether matrix interferences exist.
<b>LCS (Laboratory Control Sample)</b>	This comprises either a standard reference material or a control matrix (such as a blank sand or water) fortified with analytes representative of the analyte class. It is simply a check sample.
<b>Surrogate Spike</b>	Surrogates are known additions to each sample, blank, matrix spike and LCS in a batch, of compounds which are similar to the analyte of interest, however are not expected to be found in real samples.
Australian Drinking Water Guidelines recommend that Thermotolerant Coliform, Faecal Enterococci, & E.Coli levels are less than 1cfu/100mL. The recommended maximums are taken from "Australian Drinking Water Guidelines", published by NHMRC & ARMC 2011.	



## Laboratory Acceptance Criteria

Duplicate sample and matrix spike recoveries may not be reported on smaller jobs, however, were analysed at a frequency to meet or exceed NEPM requirements. All samples are tested in batches of 20. The duplicate sample RPD and matrix spike recoveries for the batch were within the laboratory acceptance criteria.

Filters, swabs, wipes, tubes and badges will not have duplicate data as the whole sample is generally extracted during sample extraction.

Spikes for Physical and Aggregate Tests are not applicable.

For VOCs in water samples, three vials are required for duplicate or spike analysis.

Duplicates: >10xPQL - RPD acceptance criteria will vary depending on the analytes and the analytical techniques but is typically in the range 20%-50% – see ELN-P05 QA/QC tables for details; <10xPQL - RPD are higher as the results approach PQL and the estimated measurement uncertainty will statistically increase.

Matrix Spikes, LCS and Surrogate recoveries: Generally 70-130% for inorganics/metals (not SPOCAS); 60-140% for organics/SPOCAS (+/-50% surrogates) and 10-140% for labile SVOCs (including labile surrogates), ultra trace organics and speciated phenols is acceptable.

In circumstances where no duplicate and/or sample spike has been reported at 1 in 10 and/or 1 in 20 samples respectively, the sample volume submitted was insufficient in order to satisfy laboratory QA/QC protocols.

When samples are received where certain analytes are outside of recommended technical holding times (THTs), the analysis has proceeded. Where analytes are on the verge of breaching THTs, every effort will be made to analyse within the THT or as soon as practicable.

Where sampling dates are not provided, Envirolab are not in a position to comment on the validity of the analysis where recommended technical holding times may have been breached.

Measurement Uncertainty estimates are available for most tests upon request.

Analysis of aqueous samples typically involves the extraction/digestion and/or analysis of the liquid phase only (i.e. NOT any settled sediment phase but inclusive of suspended particles if present), unless stipulated on the Envirolab COC and/or by correspondence. Notable exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, total recoverable metals and PFAS where solids are included by default.

Samples for Microbiological analysis (not Amoeba forms) received outside of the 2-8°C temperature range do not meet the ideal cooling conditions as stated in AS2031-2012.

## Report Comments

Asbestos: A portion of the supplied samples were sub-sampled for asbestos analysis according to Envirolab procedures.

We cannot guarantee that these sub-samples are indicative of the entire sample.

Envirolab recommends supplying 40-50g of sample in its own container.

Note: Samples requested for asbestos testing were sub-sampled from jars provided by the client.

8 metals in soil - # Percent recovery is not possible to report due to the inhomogeneous nature of the element/s in the sample/s. However an acceptable recovery was obtained for the LCS.

TRH Soil C10-C40 NEPM - # Percent recovery for the surrogate and matrix spike is not possible to report as the high concentration of analytes in sample 231640-1 has caused interference.

PAHs in Soil - # Percent recovery for the matrix spike is not possible to report as the high concentration of analytes in samples 231640-1 has caused interference.

OC's in Soil - The PQL has been raised due to interferences from analytes (other than those being tested) in sample 231640-1.  
# Percent recovery for the matrix spike is not possible to report due to interference from analytes (other than those being tested) in sample 231640-1.

OP's in Soil - The PQL has been raised due to interferences from analytes (other than those being tested) in sample 231640-1.  
# Percent recovery for the matrix spike is not possible to report due to interference from analytes (other than those being tested) in sample 231640-1.

PCBs in Soil - The PQL has been raised due to interferences from analytes (other than those being tested) in sample 231640-1.  
# Percent recovery for the matrix spike is not possible to report due to interference from analytes (other than those being tested) in sample 231640-1.

Rev4/October2016

## SAMPLE RECEIPT ADVICE

### Client Details

<b>Client</b>	Douglas Partners Pty Ltd
<b>Attention</b>	David Walker

### Sample Login Details

<b>Your reference</b>	86874.01, Ultimo
<b>Envirolab Reference</b>	231640
<b>Date Sample Received</b>	26/11/2019
<b>Date Instructions Received</b>	26/11/2019
<b>Date Results Expected to be Reported</b>	29/11/2019

### Sample Condition

<b>Samples received in appropriate condition for analysis</b>	Yes
<b>No. of Samples Provided</b>	4 soil
<b>Turnaround Time Requested</b>	3 days
<b>Temperature on Receipt (°C)</b>	20..8
<b>Cooling Method</b>	Ice
<b>Sampling Date Provided</b>	YES

### Comments

Nil

Please direct any queries to:

<b>Aileen Hie</b>	<b>Jacinta Hurst</b>
<b>Phone:</b> 02 9910 6200	<b>Phone:</b> 02 9910 6200
<b>Fax:</b> 02 9910 6201	<b>Fax:</b> 02 9910 6201
<b>Email:</b> ahie@envirolab.com.au	<b>Email:</b> jhurst@envirolab.com.au

Analysis Underway, details on the following page:



**EnviroLab Services Pty Ltd**

ABN 37 112 535 645

12 Ashley St Chatswood NSW 2067

ph 02 9910 6200 fax 02 9910 6201

customerservice@envirolab.com.au

www.envirolab.com.au

Sample ID	VOCs in soil	vTRH(C6-C10)/BTEXN in Soil	svTRH (C10-C40) in Soil	PAHs in Soil	Organochlorine Pesticides in soil	Organophosphorus Pesticides in Soil	PCBs in Soil	Acid Extractable metals in soil	Misc Soil - Inorg	Asbestos ID - soils
202-1.2-1.3	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
202-2.0-2.1		✓	✓	✓				✓		✓
TS		✓								
TB		✓								

The '✓' indicates the testing you have requested. **THIS IS NOT A REPORT OF THE RESULTS.**

### Additional Info

Sample storage - Waters are routinely disposed of approximately 1 month and soils approximately 2 months from receipt.

Requests for longer term sample storage must be received in writing.

Please contact the laboratory immediately if observed settled sediment present in water samples is to be included in the extraction and/or analysis (exceptions include certain Physical Tests (pH/EC/BOD/COD/Apparent Colour etc.), Solids testing, Total Recoverable metals and PFAS analysis where solids are included by default.

TAT for Micro is dependent on incubation. This varies from 3 to 6 days.



# 684979

**CHAIN OF CUSTODY DESPATCH SHEET**

<b>Project No:</b> 86874.01				<b>Suburb:</b> Olinda				<b>To:</b> Envirolab Services			
<b>Project Name:</b> Olinda				<b>Order Number</b>				12 Ashley Street, Chatswood NSW 2067			
<b>Project Manager:</b> David Walker				<b>Sampler:</b> T. Graham, R. Keavney				<b>Attn:</b>			
<b>Emails:</b> david.walker@douglaspartners.com.au				<b>Phone:</b> 02 9910 6200				<b>Email:</b> sydney@envirolab.com.au			
<b>Date Required:</b> Same day <input type="checkbox"/> 24 hours <input type="checkbox"/> 48 hours <input type="checkbox"/> 72 hours <input type="checkbox"/> Standard <input checked="" type="checkbox"/>				<b>Do samples contain 'potential' HBM?</b> Yes <input type="checkbox"/> No <input checked="" type="checkbox"/> (If YES, then handle, transport and store in accordance with FPM HAZID)							
<b>Prior Storage:</b> <input type="checkbox"/> Esky <input checked="" type="checkbox"/> Fridge <input type="checkbox"/> Shelved											

Sample ID	Lab ID	Date Sampled	Sample Type S - soil W - water	Container Type G - glass P - plastic	Analytes										Notes/preservation	
					Combo 8a	VOC	Combo 3a	Combo 1a	TRN/ BTEX	Metals (8)	BTEX only	PH+ CEC				
BH101	0.1	1	22/10/19	S	G/P	✓										subsample OK for asbestos analysis
BH102	0.5	2	22/10/19	S	G/P	✓	✓						✓			
BH103	1.0	3	22/10/19	S	G/P	✓	✓									
BH104	0.5	4	23/10/19	S	G/P	✓										
BH104	1.5	5	23/10/19	S	G/P			✓					✓			
BH104	3.0	6	23/10/19	S	G/P			✓								
BH104	4.0	7	23/10/19	S	G/P			✓								
BH108	0.1-0.2	8	22/10/19	S	G	✓										
BH109	0.15-0.2	9	22/10/19	S	G	✓										
BD2-20191022	10	22/10/19	S	G					✓							
BD3-20191023	11	23/10/19	S	G					✓							
BD4-20191022	12	22/10/19	S	G						✓	✓					Send sample to Enviro for analysis
TS	12		S	G								✓				
TB	13		S	G								✓				
<b>PQL (S) mg/kg</b>															<b>ANZECC PQLs req'd for all water analytes</b> <input type="checkbox"/>	

**PQL = practical quantitation limit.** If none given, default to Laboratory Method Detection Limit

**Metals to Analyse:** 8HM unless specified here:

**Total number of samples in container:** Relinquished by: DW Transported to laboratory by: Courier

**Send Results to:** Douglas Partners Pty Ltd **Address:** 96 Hermitage Road, West Ryde **Phone:** 98090666 **Fax:** 98094095

**Signed:** Received by: [Signature] **Date & Time:** 25/10/19



**Melbourne**

6 Monterey Road  
Dandenong South Vic 3175  
Phone : +61 3 8564 5000  
NATA # 1261  
Site # 1254 & 14271

**Sydney**

Unit F3, Building F  
16 Mars Road  
Lane Cove West NSW 2066  
Phone : +61 2 9900 8400  
NATA # 1261 Site # 18217

**Brisbane**

1/21 Smallwood Place  
Murarrie QLD 4172  
Phone : +61 7 3902 4600  
NATA # 1261 Site # 20794

**Perth**

2/91 Leach Highway  
Kewdale WA 6105  
Phone : +61 8 9251 9600  
NATA # 1261 Site # 23736

ABN – 50 005 085 521

e.mail : EnviroSales@eurofins.com

web : www.eurofins.com.au

## Sample Receipt Advice

Company name: **Douglas Partners (Syd)**  
Contact name: David Walker  
Project name: ULTIMO  
Project ID: 86874.01  
COC number: Not provided  
Turn around time: 5 Day  
Date/Time received: Oct 28, 2019 4:11 PM  
Eurofins reference: **684979**

### Sample information

- ☒ A detailed list of analytes logged into our LIMS, is included in the attached summary table.
- ☒ All samples have been received as described on the above COC.
- ☒ COC has been completed correctly.
- ☒ Attempt to chill was evident.
- ☒ Appropriately preserved sample containers have been used.
- ☒ All samples were received in good condition.
- ☒ Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.
- ☒ Appropriate sample containers have been used.
- ☐ Split sample sent to requested external lab.
- ☐ Some samples have been subcontracted.
- N/A Custody Seals intact (if used).

### Contact notes

If you have any questions with respect to these samples please contact:

Ursula Long on Phone : or by e.mail: UrsulaLong@eurofins.com

Results will be delivered electronically via e.mail to David Walker - david.walker@douglaspartners.com.au.

**Company Name:** Douglas Partners (Syd)  
**Address:** 96 Hermitage Road  
West Ryde  
NSW 2114  
**Project Name:** ULTIMO  
**Project ID:** 86874.01

**Order No.:**  
**Report #:** 684979  
**Phone:** 02 9809 0666  
**Fax:**

**Received:** Oct 28, 2019 4:11 PM  
**Due:** Nov 4, 2019  
**Priority:** 5 Day  
**Contact Name:** David Walker

**Eurofins Analytical Services Manager : Ursula Long**

Sample Detail						Moisture Set	Eurofins   mgt Suite B6
Melbourne Laboratory - NATA Site # 1254 & 14271							
Sydney Laboratory - NATA Site # 18217						X	X
Brisbane Laboratory - NATA Site # 20794							
Perth Laboratory - NATA Site # 23736							
External Laboratory							
No	Sample ID	Sample Date	Sampling Time	Matrix	LAB ID		
1	BDA/20191022	Oct 22, 2019		Rock	S19-Oc43532	X	X
Test Counts						1	1



**Douglas Partners (Syd)**  
**96 Hermitage Road**  
**West Ryde**  
**NSW 2114**



**NATA Accredited**  
**Accreditation Number 1261**  
**Site Number 18217**

Accredited for compliance with ISO/IEC 17025 – Testing  
 The results of the tests, calibrations and/or  
 measurements included in this document are traceable  
 to Australian/national standards.

**Attention:** **David Walker**

**Report** **684979-S**  
 Project name **ULTIMO**  
 Project ID **86874.01**  
 Received Date **Oct 28, 2019**

<b>Client Sample ID</b>			<b>BDA/20191022</b>
<b>Sample Matrix</b>			<b>Rock</b>
<b>Eurofins Sample No.</b>			<b>S19-Oc43532</b>
<b>Date Sampled</b>			<b>Oct 22, 2019</b>
Test/Reference	LOR	Unit	
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>			
TRH C6-C9	20	mg/kg	< 20
TRH C10-C14	20	mg/kg	< 20
TRH C15-C28	50	mg/kg	< 50
TRH C29-C36	50	mg/kg	< 50
TRH C10-C36 (Total)	50	mg/kg	< 50
<b>BTEX</b>			
Benzene	0.1	mg/kg	< 0.1
Toluene	0.1	mg/kg	< 0.1
Ethylbenzene	0.1	mg/kg	< 0.1
m&p-Xylenes	0.2	mg/kg	< 0.2
o-Xylene	0.1	mg/kg	< 0.1
Xylenes - Total	0.3	mg/kg	< 0.3
4-Bromofluorobenzene (surr.)	1	%	84
<b>Total Recoverable Hydrocarbons - 2013 NEPM Fractions</b>			
Naphthalene <sup>N02</sup>	0.5	mg/kg	< 0.5
TRH C6-C10	20	mg/kg	< 20
TRH C6-C10 less BTEX (F1) <sup>N04</sup>	20	mg/kg	< 20
TRH >C10-C16	50	mg/kg	< 50
TRH >C10-C16 less Naphthalene (F2) <sup>N01</sup>	50	mg/kg	< 50
TRH >C16-C34	100	mg/kg	< 100
TRH >C34-C40	100	mg/kg	< 100
TRH >C10-C40 (total)*	100	mg/kg	< 100
<b>Heavy Metals</b>			
Arsenic	2	mg/kg	< 2
Cadmium	0.4	mg/kg	< 0.4
Chromium	5	mg/kg	67
Copper	5	mg/kg	22
Lead	5	mg/kg	14
Mercury	0.1	mg/kg	< 0.1
Nickel	5	mg/kg	67
Zinc	5	mg/kg	31
% Moisture	1	%	3.7

## Sample History

Where samples are submitted/analysed over several days, the last date of extraction and analysis is reported.

A recent review of our LIMS has resulted in the correction or clarification of some method identifications. Due to this, some of the method reference information on reports has changed. However, no substantive change has been made to our laboratory methods, and as such there is no change in the validity of current or previous results.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Eurofins   mgt Suite B6			
Total Recoverable Hydrocarbons - 1999 NEPM Fractions	Sydney	Oct 30, 2019	14 Days
- Method: LTM-ORG-2010 TRH C6-C40			
BTEX	Sydney	Oct 30, 2019	14 Days
- Method: LTM-ORG-2010 TRH C6-C40			
Total Recoverable Hydrocarbons - 2013 NEPM Fractions	Sydney	Oct 30, 2019	14 Days
- Method: LTM-ORG-2010 TRH C6-C40			
Total Recoverable Hydrocarbons - 2013 NEPM Fractions	Sydney	Oct 30, 2019	
- Method: LTM-ORG-2010 TRH C6-C40			
Metals M8	Sydney	Oct 30, 2019	180 Days
- Method: LTM-MET-3040 Metals in Waters, Soils & Sediments by ICP-MS			
% Moisture	Sydney	Oct 28, 2019	14 Days
- Method: LTM-GEN-7080 Moisture			

**Company Name:** Douglas Partners (Syd)  
**Address:** 96 Hermitage Road  
West Ryde  
NSW 2114  
**Project Name:** ULTIMO  
**Project ID:** 86874.01

**Order No.:**  
**Report #:** 684979  
**Phone:** 02 9809 0666  
**Fax:**

**Received:** Oct 28, 2019 4:11 PM  
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**Contact Name:** David Walker

**Eurofins Analytical Services Manager : Ursula Long**

Sample Detail						Moisture Set	Eurofins   mgt Suite B6
Melbourne Laboratory - NATA Site # 1254 & 14271							
Sydney Laboratory - NATA Site # 18217						X	X
Brisbane Laboratory - NATA Site # 20794							
Perth Laboratory - NATA Site # 23736							
External Laboratory							
No	Sample ID	Sample Date	Sampling Time	Matrix	LAB ID		
1	BDA/20191022	Oct 22, 2019		Rock	S19-Oc43532	X	X
Test Counts						1	1

## Internal Quality Control Review and Glossary

### General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follows guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013 and are included in this QC report where applicable. Additional QC data may be available on request.
2. All soil/sediment/solid results are reported on a dry basis, unless otherwise stated.
3. All biota/food results are reported on a wet weight basis on the edible portion, unless otherwise stated.
4. Actual LORs are matrix dependant. Quoted LORs may be raised where sample extracts are diluted due to interferences.
5. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds.
6. SVOC analysis on waters are performed on homogenised, unfiltered samples, unless noted otherwise.
7. Samples were analysed on an 'as received' basis.
8. Information identified on this report with blue colour, indicates data provided by customer, that may have an impact on the results.
9. This report replaces any interim results previously issued.

### Holding Times

Please refer to 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours prior to sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and regardless of any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the date of sampling, therefore compliance to these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether the holding time is 7 days however for all other VOCs such as BTEX or C6-10 TRH then the holding time is 14 days.

**\*\*NOTE:** pH duplicates are reported as a range NOT as RPD

### Units

**mg/kg:** milligrams per kilogram

**mg/L:** milligrams per litre

**ug/L:** micrograms per litre

**ppm:** Parts per million

**ppb:** Parts per billion

**%:** Percentage

**org/100mL:** Organisms per 100 millilitres

**NTU:** Nephelometric Turbidity Units

**MPN/100mL:** Most Probable Number of organisms per 100 millilitres

### Terms

<b>Dry</b>	Where a moisture has been determined on a solid sample the result is expressed on a dry basis.
<b>LOR</b>	Limit of Reporting.
<b>SPIKE</b>	Addition of the analyte to the sample and reported as percentage recovery.
<b>RPD</b>	Relative Percent Difference between two Duplicate pieces of analysis.
<b>LCS</b>	Laboratory Control Sample - reported as percent recovery.
<b>CRM</b>	Certified Reference Material - reported as percent recovery.
<b>Method Blank</b>	In the case of solid samples these are performed on laboratory certified clean sands and in the case of water samples these are performed on de-ionised water.
<b>Surr - Surrogate</b>	The addition of a like compound to the analyte target and reported as percentage recovery.
<b>Duplicate</b>	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
<b>USEPA</b>	United States Environmental Protection Agency
<b>APHA</b>	American Public Health Association
<b>TCLP</b>	Toxicity Characteristic Leaching Procedure
<b>COC</b>	Chain of Custody
<b>SRA</b>	Sample Receipt Advice
<b>QSM</b>	US Department of Defense Quality Systems Manual Version 5.3
<b>CP</b>	Client Parent - QC was performed on samples pertaining to this report
<b>NC</b>	Non-Client Parent - QC performed on samples not pertaining to this report, QC is representative of the sequence or batch that client samples were analysed within.
<b>TEQ</b>	Toxic Equivalency Quotient

### QC - Acceptance Criteria

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is 30% however the following acceptance guidelines are equally applicable:

Results <10 times the LOR : No Limit

Results between 10-20 times the LOR : RPD must lie between 0-50%

Results >20 times the LOR : RPD must lie between 0-30%

Surrogate Recoveries: Recoveries must lie between 20-130% Phenols & 50-150% PFASs

PFAS field samples that contain surrogate recoveries in excess of the QC limit designated in QSM 5.3 where no positive PFAS results have been reported have been reviewed and no data was affected.

WA DWER (n=10): PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

### QC Data General Comments

1. Where a result is reported as a less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch, but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown is not data from your samples.
3. Organochlorine Pesticide analysis - where reporting LCS data, Toxaphene & Chlordane are not added to the LCS.
4. Organochlorine Pesticide analysis - where reporting Spike data, Toxaphene is not added to the Spike.
5. Total Recoverable Hydrocarbons - where reporting Spike & LCS data, a single spike of commercial Hydrocarbon products in the range of C12-C30 is added and it's Total Recovery is reported in the C10-C14 cell of the Report.
6. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore laboratory analysis is unlikely to be completed within holding time. Analysis will begin as soon as possible after sample receipt.
7. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of Recovery the term "INT" appears against that analyte.
8. Polychlorinated Biphenyls are spiked only using Aroclor 1260 in Matrix Spikes and LCS.
9. For Matrix Spikes and LCS results a dash " - " in the report means that the specific analyte was not added to the QC sample.
10. Duplicate RPDs are calculated from raw analytical data thus it is possible to have two sets of data.

## Quality Control Results

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
<b>Method Blank</b>							
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>							
TRH C6-C9	mg/kg	< 20			20	Pass	
TRH C10-C14	mg/kg	< 20			20	Pass	
TRH C15-C28	mg/kg	< 50			50	Pass	
TRH C29-C36	mg/kg	< 50			50	Pass	
<b>Method Blank</b>							
<b>BTEX</b>							
Benzene	mg/kg	< 0.1			0.1	Pass	
Toluene	mg/kg	< 0.1			0.1	Pass	
Ethylbenzene	mg/kg	< 0.1			0.1	Pass	
m&p-Xylenes	mg/kg	< 0.2			0.2	Pass	
o-Xylene	mg/kg	< 0.1			0.1	Pass	
Xylenes - Total	mg/kg	< 0.3			0.3	Pass	
<b>Method Blank</b>							
<b>Total Recoverable Hydrocarbons - 2013 NEPM Fractions</b>							
Naphthalene	mg/kg	< 0.5			0.5	Pass	
TRH C6-C10	mg/kg	< 20			20	Pass	
TRH >C10-C16	mg/kg	< 50			50	Pass	
TRH >C16-C34	mg/kg	< 100			100	Pass	
TRH >C34-C40	mg/kg	< 100			100	Pass	
<b>Method Blank</b>							
<b>Heavy Metals</b>							
Arsenic	mg/kg	< 2			2	Pass	
Cadmium	mg/kg	< 0.4			0.4	Pass	
Chromium	mg/kg	< 5			5	Pass	
Copper	mg/kg	< 5			5	Pass	
Lead	mg/kg	< 5			5	Pass	
Mercury	mg/kg	< 0.1			0.1	Pass	
Nickel	mg/kg	< 5			5	Pass	
Zinc	mg/kg	< 5			5	Pass	
<b>LCS - % Recovery</b>							
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>							
TRH C6-C9	%	97			70-130	Pass	
TRH C10-C14	%	124			70-130	Pass	
<b>LCS - % Recovery</b>							
<b>BTEX</b>							
Benzene	%	104			70-130	Pass	
Toluene	%	104			70-130	Pass	
Ethylbenzene	%	105			70-130	Pass	
m&p-Xylenes	%	108			70-130	Pass	
o-Xylene	%	107			70-130	Pass	
Xylenes - Total	%	108			70-130	Pass	
<b>LCS - % Recovery</b>							
<b>Total Recoverable Hydrocarbons - 2013 NEPM Fractions</b>							
Naphthalene	%	94			70-130	Pass	
TRH C6-C10	%	91			70-130	Pass	
TRH >C10-C16	%	122			70-130	Pass	
<b>LCS - % Recovery</b>							
<b>Heavy Metals</b>							
Arsenic	%	94			70-130	Pass	
Cadmium	%	87			70-130	Pass	

Test			Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Chromium			%	94			70-130	Pass	
Copper			%	96			70-130	Pass	
Lead			%	96			70-130	Pass	
Mercury			%	93			70-130	Pass	
Nickel			%	96			70-130	Pass	
Zinc			%	90			70-130	Pass	
Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
<b>Spike - % Recovery</b>									
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>				Result 1					
TRH C6-C9	S19-Oc46824	NCP	%	98			70-130	Pass	
TRH C10-C14	S19-Oc38300	NCP	%	120			70-130	Pass	
<b>Spike - % Recovery</b>									
<b>BTEX</b>				Result 1					
Benzene	S19-Oc46824	NCP	%	98			70-130	Pass	
Toluene	S19-Oc46824	NCP	%	95			70-130	Pass	
Ethylbenzene	S19-Oc46824	NCP	%	96			70-130	Pass	
m&p-Xylenes	S19-Oc46824	NCP	%	99			70-130	Pass	
o-Xylene	S19-Oc46824	NCP	%	98			70-130	Pass	
Xylenes - Total	S19-Oc46824	NCP	%	99			70-130	Pass	
<b>Spike - % Recovery</b>									
<b>Total Recoverable Hydrocarbons - 2013 NEPM Fractions</b>				Result 1					
Naphthalene	S19-Oc46824	NCP	%	86			70-130	Pass	
TRH C6-C10	S19-Oc46824	NCP	%	98			70-130	Pass	
TRH >C10-C16	S19-Oc38300	NCP	%	118			70-130	Pass	
<b>Spike - % Recovery</b>									
<b>Heavy Metals</b>				Result 1					
Arsenic	S19-Oc41515	NCP	%	110			70-130	Pass	
Cadmium	S19-Oc41515	NCP	%	104			70-130	Pass	
Chromium	S19-Oc41515	NCP	%	108			70-130	Pass	
Copper	S19-Oc41515	NCP	%	116			70-130	Pass	
Lead	S19-Oc41515	NCP	%	114			70-130	Pass	
Mercury	S19-Oc41515	NCP	%	109			70-130	Pass	
Nickel	S19-Oc41515	NCP	%	104			70-130	Pass	
Zinc	S19-Oc41515	NCP	%	113			70-130	Pass	
Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
<b>Duplicate</b>									
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>				Result 1	Result 2	RPD			
TRH C6-C9	S19-Oc41514	NCP	mg/kg	< 20	< 20	<1	30%	Pass	
TRH C10-C14	S19-Oc46822	NCP	mg/kg	< 20	< 20	<1	30%	Pass	
TRH C15-C28	S19-Oc46822	NCP	mg/kg	< 50	< 50	<1	30%	Pass	
TRH C29-C36	S19-Oc46822	NCP	mg/kg	< 50	< 50	<1	30%	Pass	
<b>Duplicate</b>									
<b>BTEX</b>				Result 1	Result 2	RPD			
Benzene	S19-Oc41514	NCP	mg/kg	< 0.1	< 0.1	<1	30%	Pass	
Toluene	S19-Oc41514	NCP	mg/kg	< 0.1	< 0.1	<1	30%	Pass	
Ethylbenzene	S19-Oc41514	NCP	mg/kg	< 0.1	< 0.1	<1	30%	Pass	
m&p-Xylenes	S19-Oc41514	NCP	mg/kg	0.5	0.5	4.0	30%	Pass	
o-Xylene	S19-Oc41514	NCP	mg/kg	0.2	0.2	6.0	30%	Pass	
Xylenes - Total	S19-Oc41514	NCP	mg/kg	0.7	0.7	4.0	30%	Pass	

Duplicate								
Total Recoverable Hydrocarbons - 2013 NEPM Fractions				Result 1	Result 2	RPD		
Naphthalene	S19-Oc41514	NCP	mg/kg	< 0.5	< 0.5	<1	30%	Pass
TRH C6-C10	S19-Oc41514	NCP	mg/kg	< 20	< 20	<1	30%	Pass
TRH >C10-C16	S19-Oc46822	NCP	mg/kg	< 50	< 50	<1	30%	Pass
TRH >C16-C34	S19-Oc46822	NCP	mg/kg	< 100	< 100	<1	30%	Pass
TRH >C34-C40	S19-Oc46822	NCP	mg/kg	< 100	< 100	<1	30%	Pass
Duplicate								
Heavy Metals				Result 1	Result 2	RPD		
Arsenic	S19-Oc41514	NCP	mg/kg	2.6	2.3	11	30%	Pass
Cadmium	S19-Oc41514	NCP	mg/kg	< 0.4	< 0.4	<1	30%	Pass
Chromium	S19-Oc41514	NCP	mg/kg	15	16	7.0	30%	Pass
Copper	S19-Oc41514	NCP	mg/kg	17	16	8.0	30%	Pass
Lead	S19-Oc41514	NCP	mg/kg	12	12	3.0	30%	Pass
Mercury	S19-Oc41514	NCP	mg/kg	< 0.1	< 0.1	<1	30%	Pass
Nickel	S19-Oc41514	NCP	mg/kg	28	24	18	30%	Pass
Zinc	S19-Oc41514	NCP	mg/kg	44	44	1.0	30%	Pass
Duplicate								
				Result 1	Result 2	RPD		
% Moisture	S19-Oc43487	NCP	%	11	12	6.0	30%	Pass

## Comments

### Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

### Qualifier Codes/Comments

Code	Description
N01	F2 is determined by arithmetically subtracting the "naphthalene" value from the ">C10-C16" value. The naphthalene value used in this calculation is obtained from volatiles (Purge & Trap analysis).
N02	Where we have reported both volatile (P&T GCMS) and semivolatile (GCMS) naphthalene data, results may not be identical. Provided correct sample handling protocols have been followed, any observed differences in results are likely to be due to procedural differences within each methodology. Results determined by both techniques have passed all QAQC acceptance criteria, and are entirely technically valid.
N04	F1 is determined by arithmetically subtracting the "Total BTEX" value from the "C6-C10" value. The "Total BTEX" value is obtained by summing the concentrations of BTEX analytes. The "C6-C10" value is obtained by quantitating against a standard of mixed aromatic/aliphatic analytes.

## Authorised By

Ursula Long	Analytical Services Manager
Andrew Sullivan	Senior Analyst-Organic (NSW)
Gabriele Cordero	Senior Analyst-Metal (NSW)



### Glenn Jackson General Manager

Final report - this Report replaces any previously issued Report

- Indicates Not Requested

\* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please [click here](#).

Eurofins shall not be liable for loss, cost, damages or expenses incurred by the client, or any other person or company, resulting from the use of any information or interpretation given in this report. In no case shall Eurofins be liable for consequential damages including, but not limited to, lost profits, damages for failure to meet deadlines and lost production arising from this report. This document shall not be reproduced except in full and relates only to the items tested. Unless indicated otherwise, the tests were performed on the samples as received.



### CHAIN OF CUSTODY DESPATCH SHEET

<b>Project No:</b> 86874-01	<b>Suburb:</b> Ultimo	<b>To:</b> Envirolab Services
<b>Project Name:</b> Ultimo	<b>Order Number</b>	12 Ashley Street, Chatswood NSW 2067
<b>Project Manager:</b> David Walker	<b>Sampler:</b> Rob Keaveney	<b>Attn:</b>
<b>Emails:</b> david.walker@douglaspartners.com.au		<b>Phone:</b> 02 9910 6200
<b>Date Required:</b> Same day <input type="checkbox"/> 24 hours <input type="checkbox"/> 48 hours <input type="checkbox"/> 72 hours <input checked="" type="checkbox"/> Standard <input type="checkbox"/>		<b>Email:</b> sydney@envirolab.com.au
<b>Prior Storage:</b> <input type="checkbox"/> Esky <input checked="" type="checkbox"/> Fridge <input type="checkbox"/> Shelved	Do samples contain 'potential' HBM? Yes <input type="checkbox"/> No <input checked="" type="checkbox"/>	(If YES, then handle, transport and store in accordance with FPM HAZID)

Sample ID	Lab ID	Date Sampled	Sample Type	Container Type	Analytes										Notes/preservation
			S - soil W - water	G - glass P - plastic	Conso 8a	Conso 3a	VOC	BTEX only	Metals (8)	TRH BTEX					
202	1-2-1.3	1	26-11-19	S	G/P	✓		✓							OK to subsample for asbestos.
202	2-0-2.1	2	26-11-19	S	G/P		✓								
TS	3	↓		S	G				✓						
TB	A			S	G				✓						
BD 26/11/2019	-			S	G						✓	✓			* Send sample to Envirofins for analysis 3 day TAT
										</					

PQL = practical quantitation limit. If none given, default to Laboratory Method Detection Limit

ANZECC PQLs req'd for all water analytes ☐

Metals to Analyse: 8HM unless specified here:

Lab Report/Reference No: 231640

Total number of samples in container:

Relinquished by: DW

Transported to laboratory by:

Courier

**Send Results to:** Douglas Partners Pty Ltd

**Address:** 96 Hermitage Road, West Ryde

Phone:	98090666	Fax:	98094095
--------	----------	------	----------

**Signed:**

Received by: EZS, Trinidad Comptroller

Date & Time: 26.11.19, 17:15

Relinquished by: ELS Syd K. Core 27.11.2019 1200

Rec: Grave Eureka 27/11 2:40pm #690424

**Melbourne**

6 Monterey Road  
Dandenong South Vic 3175  
Phone : +61 3 8564 5000  
NATA # 1261  
Site # 1254 & 14271

**Sydney**

Unit F3, Building F  
16 Mars Road  
Lane Cove West NSW 2066  
Phone : +61 2 9900 8400  
NATA # 1261 Site # 18217

**Brisbane**

1/21 Smallwood Place  
Murarrie QLD 4172  
Phone : +61 7 3902 4600  
NATA # 1261 Site # 20794

**Perth**

2/91 Leach Highway  
Kewdale WA 6105  
Phone : +61 8 9251 9600  
NATA # 1261 Site # 23736

ABN – 50 005 085 521

e.mail : EnviroSales@eurofins.com

web : www.eurofins.com.au

## Sample Receipt Advice

Company name: **Douglas Partners (Syd)**  
Contact name: David Walker  
Project name: ULTIMO  
Project ID: 86874.01  
COC number: Not provided  
Turn around time: 3 Day  
Date/Time received: Nov 27, 2019 2:40 PM  
Eurofins reference: **690424**

### Sample information

- ☒ A detailed list of analytes logged into our LIMS, is included in the attached summary table.
- ☒ All samples have been received as described on the above COC.
- ☒ COC has been completed correctly.
- ☒ Attempt to chill was evident.
- ☒ Appropriately preserved sample containers have been used.
- ☒ All samples were received in good condition.
- ☒ Samples have been provided with adequate time to commence analysis in accordance with the relevant holding times.
- ☒ Appropriate sample containers have been used.
- ☐ Split sample sent to requested external lab.
- ☐ Some samples have been subcontracted.
- N/A Custody Seals intact (if used).

### Contact notes

If you have any questions with respect to these samples please contact:

Ursula Long on Phone : or by e.mail: UrsulaLong@eurofins.com

Results will be delivered electronically via e.mail to David Walker - david.walker@douglaspartners.com.au.

**Melbourne**

6 Monterey Road  
Dandenong South Vic 3175  
Phone : +61 3 8564 5000  
NATA # 1261  
Site # 1254 & 14271

**Sydney**

Unit F3, Building F  
16 Mars Road  
Lane Cove West NSW 2066  
Phone : +61 2 9900 8400  
NATA # 1261 Site # 18217

**Brisbane**

1/21 Smallwood Place  
Murarrie QLD 4172  
Phone : +61 7 3902 4600  
NATA # 1261 Site # 20794

**Perth**

2/91 Leach Highway  
Kewdale WA 6105  
Phone : +61 8 9251 9600  
NATA # 1261 Site # 23736

ABN – 50 005 085 521

e.mail : EnviroSales@eurofins.com

web : www.eurofins.com.au

## Sample Receipt Advice

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Contact name: David Walker  
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Ursula Long on Phone : or by e.mail: UrsulaLong@eurofins.com

Results will be delivered electronically via e.mail to David Walker - david.walker@douglaspartners.com.au.

**Company Name:** Douglas Partners (Syd)  
**Address:** 96 Hermitage Road  
West Ryde  
NSW 2114  
**Project Name:** ULTIMO  
**Project ID:** 86874.01

**Order No.:**  
**Report #:** 690424  
**Phone:** 02 9809 0666  
**Fax:**

**Received:** Nov 27, 2019 2:40 PM  
**Due:** Dec 2, 2019  
**Priority:** 3 Day  
**Contact Name:** David Walker

**Eurofins Analytical Services Manager : Ursula Long**

Sample Detail						Moisture Set	Eurofins   mgt Suite B6
Melbourne Laboratory - NATA Site # 1254 & 14271							
Sydney Laboratory - NATA Site # 18217						X	X
Brisbane Laboratory - NATA Site # 20794							
Perth Laboratory - NATA Site # 23736							
External Laboratory							
No	Sample ID	Sample Date	Sampling Time	Matrix	LAB ID		
1	BD26112019	Nov 26, 2019		Soil	S19-No38471	X	X
Test Counts						1	1

**Douglas Partners (Syd)**  
**96 Hermitage Road**  
**West Ryde**  
**NSW 2114**



**NATA Accredited**  
**Accreditation Number 1261**  
**Site Number 18217**

Accredited for compliance with ISO/IEC 17025 – Testing  
 The results of the tests, calibrations and/or  
 measurements included in this document are traceable  
 to Australian/national standards.

**Attention:** **David Walker**

**Report** **690424-S**  
 Project name **ULTIMO**  
 Project ID **86874.01**  
 Received Date **Nov 27, 2019**

<b>Client Sample ID</b>			G01 <b>BD26112019</b>
<b>Sample Matrix</b>			<b>Soil</b>
<b>Eurofins Sample No.</b>			<b>S19-No38471</b>
<b>Date Sampled</b>			<b>Nov 26, 2019</b>
Test/Reference	LOR	Unit	
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>			
TRH C6-C9	20	mg/kg	< 200
TRH C10-C14	20	mg/kg	< 200
TRH C15-C28	50	mg/kg	2000
TRH C29-C36	50	mg/kg	600
TRH C10-C36 (Total)	50	mg/kg	2600
<b>BTEX</b>			
Benzene	0.1	mg/kg	< 0.5
Toluene	0.1	mg/kg	< 0.5
Ethylbenzene	0.1	mg/kg	< 0.5
m&p-Xylenes	0.2	mg/kg	< 1
o-Xylene	0.1	mg/kg	< 0.5
Xylenes - Total	0.3	mg/kg	< 1.5
4-Bromofluorobenzene (surr.)	1	%	87
<b>Total Recoverable Hydrocarbons - 2013 NEPM Fractions</b>			
Naphthalene <sup>N02</sup>	0.5	mg/kg	26
TRH C6-C10	20	mg/kg	< 200
TRH C6-C10 less BTEX (F1) <sup>N04</sup>	20	mg/kg	< 200
TRH >C10-C16	50	mg/kg	< 500
TRH >C10-C16 less Naphthalene (F2) <sup>N01</sup>	50	mg/kg	< 500
TRH >C16-C34	100	mg/kg	2300
TRH >C34-C40	100	mg/kg	< 1000
TRH >C10-C40 (total)*	100	mg/kg	2300
<b>Heavy Metals</b>			
Arsenic	2	mg/kg	8.0
Cadmium	0.4	mg/kg	< 0.4
Chromium	5	mg/kg	45
Copper	5	mg/kg	120
Lead	5	mg/kg	1300
Mercury	0.1	mg/kg	3.9
Nickel	5	mg/kg	40
Zinc	5	mg/kg	240
% Moisture	1	%	11



## Sample History

Where samples are submitted/analysed over several days, the last date of extraction and analysis is reported.

A recent review of our LIMS has resulted in the correction or clarification of some method identifications. Due to this, some of the method reference information on reports has changed. However, no substantive change has been made to our laboratory methods, and as such there is no change in the validity of current or previous results.

If the date and time of sampling are not provided, the Laboratory will not be responsible for compromised results should testing be performed outside the recommended holding time.

Description	Testing Site	Extracted	Holding Time
Eurofins   mgt Suite B6			
Total Recoverable Hydrocarbons - 1999 NEPM Fractions	Sydney	Nov 28, 2019	14 Days
- Method: LTM-ORG-2010 TRH C6-C40			
BTEX	Sydney	Nov 28, 2019	14 Days
- Method: LTM-ORG-2010 TRH C6-C40			
Total Recoverable Hydrocarbons - 2013 NEPM Fractions	Sydney	Nov 28, 2019	14 Days
- Method: LTM-ORG-2010 TRH C6-C40			
Total Recoverable Hydrocarbons - 2013 NEPM Fractions	Sydney	Nov 28, 2019	
- Method: LTM-ORG-2010 TRH C6-C40			
Metals M8	Sydney	Nov 28, 2019	180 Days
- Method: LTM-MET-3040 Metals in Waters, Soils & Sediments by ICP-MS			
% Moisture	Sydney	Nov 27, 2019	14 Days
- Method: LTM-GEN-7080 Moisture			

**Company Name:** Douglas Partners (Syd)  
**Address:** 96 Hermitage Road  
West Ryde  
NSW 2114  
**Project Name:** ULTIMO  
**Project ID:** 86874.01

**Order No.:**  
**Report #:** 690424  
**Phone:** 02 9809 0666  
**Fax:**

**Received:** Nov 27, 2019 2:40 PM  
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**Priority:** 3 Day  
**Contact Name:** David Walker

**Eurofins Analytical Services Manager : Ursula Long**

Sample Detail						Moisture Set	Eurofins   mgt Suite B6
Melbourne Laboratory - NATA Site # 1254 & 14271							
Sydney Laboratory - NATA Site # 18217						X	X
Brisbane Laboratory - NATA Site # 20794							
Perth Laboratory - NATA Site # 23736							
External Laboratory							
No	Sample ID	Sample Date	Sampling Time	Matrix	LAB ID		
1	BD26112019	Nov 26, 2019		Soil	S19-No38471	X	X
Test Counts						1	1

## Internal Quality Control Review and Glossary

### General

1. Laboratory QC results for Method Blanks, Duplicates, Matrix Spikes, and Laboratory Control Samples follows guidelines delineated in the National Environment Protection (Assessment of Site Contamination) Measure 1999, as amended May 2013 and are included in this QC report where applicable. Additional QC data may be available on request.
2. All soil/sediment/solid results are reported on a dry basis, unless otherwise stated.
3. All biota/food results are reported on a wet weight basis on the edible portion, unless otherwise stated.
4. Actual LORs are matrix dependant. Quoted LORs may be raised where sample extracts are diluted due to interferences.
5. Results are uncorrected for matrix spikes or surrogate recoveries except for PFAS compounds.
6. SVOC analysis on waters are performed on homogenised, unfiltered samples, unless noted otherwise.
7. Samples were analysed on an 'as received' basis.
8. Information identified on this report with blue colour, indicates data provided by customer, that may have an impact on the results.
9. This report replaces any interim results previously issued.

### Holding Times

Please refer to 'Sample Preservation and Container Guide' for holding times (QS3001).

For samples received on the last day of holding time, notification of testing requirements should have been received at least 6 hours prior to sample receipt deadlines as stated on the SRA.

If the Laboratory did not receive the information in the required timeframe, and regardless of any other integrity issues, suitably qualified results may still be reported.

Holding times apply from the date of sampling, therefore compliance to these may be outside the laboratory's control.

For VOCs containing vinyl chloride, styrene and 2-chloroethyl vinyl ether the holding time is 7 days however for all other VOCs such as BTEX or C6-10 TRH then the holding time is 14 days.

**\*\*NOTE:** pH duplicates are reported as a range NOT as RPD

### Units

**mg/kg:** milligrams per kilogram

**mg/L:** milligrams per litre

**ug/L:** micrograms per litre

**ppm:** Parts per million

**ppb:** Parts per billion

**%:** Percentage

**org/100mL:** Organisms per 100 millilitres

**NTU:** Nephelometric Turbidity Units

**MPN/100mL:** Most Probable Number of organisms per 100 millilitres

### Terms

<b>Dry</b>	Where a moisture has been determined on a solid sample the result is expressed on a dry basis.
<b>LOR</b>	Limit of Reporting.
<b>SPIKE</b>	Addition of the analyte to the sample and reported as percentage recovery.
<b>RPD</b>	Relative Percent Difference between two Duplicate pieces of analysis.
<b>LCS</b>	Laboratory Control Sample - reported as percent recovery.
<b>CRM</b>	Certified Reference Material - reported as percent recovery.
<b>Method Blank</b>	In the case of solid samples these are performed on laboratory certified clean sands and in the case of water samples these are performed on de-ionised water.
<b>Surr - Surrogate</b>	The addition of a like compound to the analyte target and reported as percentage recovery.
<b>Duplicate</b>	A second piece of analysis from the same sample and reported in the same units as the result to show comparison.
<b>USEPA</b>	United States Environmental Protection Agency
<b>APHA</b>	American Public Health Association
<b>TCLP</b>	Toxicity Characteristic Leaching Procedure
<b>COC</b>	Chain of Custody
<b>SRA</b>	Sample Receipt Advice
<b>QSM</b>	US Department of Defense Quality Systems Manual Version 5.3
<b>CP</b>	Client Parent - QC was performed on samples pertaining to this report
<b>NC</b>	Non-Client Parent - QC performed on samples not pertaining to this report, QC is representative of the sequence or batch that client samples were analysed within.
<b>TEQ</b>	Toxic Equivalency Quotient

### QC - Acceptance Criteria

RPD Duplicates: Global RPD Duplicates Acceptance Criteria is 30% however the following acceptance guidelines are equally applicable:

Results <10 times the LOR : No Limit

Results between 10-20 times the LOR : RPD must lie between 0-50%

Results >20 times the LOR : RPD must lie between 0-30%

Surrogate Recoveries: Recoveries must lie between 20-130% Phenols & 50-150% PFASs

PFAS field samples that contain surrogate recoveries in excess of the QC limit designated in QSM 5.3 where no positive PFAS results have been reported have been reviewed and no data was affected.

WA DWER (n=10): PFBA, PFPeA, PFHxA, PFHpA, PFOA, PFBS, PFHxS, PFOS, 6:2 FTSA, 8:2 FTSA

### QC Data General Comments

1. Where a result is reported as a less than (<), higher than the nominated LOR, this is due to either matrix interference, extract dilution required due to interferences or contaminant levels within the sample, high moisture content or insufficient sample provided.
2. Duplicate data shown within this report that states the word "BATCH" is a Batch Duplicate from outside of your sample batch, but within the laboratory sample batch at a 1:10 ratio. The Parent and Duplicate data shown is not data from your samples.
3. Organochlorine Pesticide analysis - where reporting LCS data, Toxaphene & Chlordane are not added to the LCS.
4. Organochlorine Pesticide analysis - where reporting Spike data, Toxaphene is not added to the Spike.
5. Total Recoverable Hydrocarbons - where reporting Spike & LCS data, a single spike of commercial Hydrocarbon products in the range of C12-C30 is added and it's Total Recovery is reported in the C10-C14 cell of the Report.
6. pH and Free Chlorine analysed in the laboratory - Analysis on this test must begin within 30 minutes of sampling. Therefore laboratory analysis is unlikely to be completed within holding time. Analysis will begin as soon as possible after sample receipt.
7. Recovery Data (Spikes & Surrogates) - where chromatographic interference does not allow the determination of Recovery the term "INT" appears against that analyte.
8. Polychlorinated Biphenyls are spiked only using Aroclor 1260 in Matrix Spikes and LCS.
9. For Matrix Spikes and LCS results a dash " - " in the report means that the specific analyte was not added to the QC sample.
10. Duplicate RPDs are calculated from raw analytical data thus it is possible to have two sets of data.



## Quality Control Results

Test	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
<b>Method Blank</b>							
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>							
TRH C6-C9	mg/kg	< 20			20	Pass	
TRH C10-C14	mg/kg	< 20			20	Pass	
TRH C15-C28	mg/kg	< 50			50	Pass	
TRH C29-C36	mg/kg	< 50			50	Pass	
<b>Method Blank</b>							
<b>BTEX</b>							
Benzene	mg/kg	< 0.1			0.1	Pass	
Toluene	mg/kg	< 0.1			0.1	Pass	
Ethylbenzene	mg/kg	< 0.1			0.1	Pass	
m&p-Xylenes	mg/kg	< 0.2			0.2	Pass	
o-Xylene	mg/kg	< 0.1			0.1	Pass	
Xylenes - Total	mg/kg	< 0.3			0.3	Pass	
<b>Method Blank</b>							
<b>Total Recoverable Hydrocarbons - 2013 NEPM Fractions</b>							
Naphthalene	mg/kg	< 0.5			0.5	Pass	
TRH C6-C10	mg/kg	< 20			20	Pass	
TRH >C10-C16	mg/kg	< 50			50	Pass	
TRH >C16-C34	mg/kg	< 100			100	Pass	
TRH >C34-C40	mg/kg	< 100			100	Pass	
<b>Method Blank</b>							
<b>Heavy Metals</b>							
Arsenic	mg/kg	< 2			2	Pass	
Cadmium	mg/kg	< 0.4			0.4	Pass	
Chromium	mg/kg	< 5			5	Pass	
Copper	mg/kg	< 5			5	Pass	
Lead	mg/kg	< 5			5	Pass	
Mercury	mg/kg	< 0.1			0.1	Pass	
Nickel	mg/kg	< 5			5	Pass	
Zinc	mg/kg	< 5			5	Pass	
<b>LCS - % Recovery</b>							
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>							
TRH C6-C9	%	86			70-130	Pass	
TRH C10-C14	%	98			70-130	Pass	
<b>LCS - % Recovery</b>							
<b>BTEX</b>							
Benzene	%	89			70-130	Pass	
Toluene	%	90			70-130	Pass	
Ethylbenzene	%	96			70-130	Pass	
m&p-Xylenes	%	89			70-130	Pass	
o-Xylene	%	86			70-130	Pass	
Xylenes - Total	%	88			70-130	Pass	
<b>LCS - % Recovery</b>							
<b>Total Recoverable Hydrocarbons - 2013 NEPM Fractions</b>							
Naphthalene	%	112			70-130	Pass	
TRH C6-C10	%	87			70-130	Pass	
TRH >C10-C16	%	92			70-130	Pass	
<b>LCS - % Recovery</b>							
<b>Heavy Metals</b>							
Arsenic	%	101			70-130	Pass	
Cadmium	%	104			70-130	Pass	

Test			Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
Chromium			%	111			70-130	Pass	
Copper			%	113			70-130	Pass	
Lead			%	113			70-130	Pass	
Mercury			%	108			70-130	Pass	
Nickel			%	113			70-130	Pass	
Zinc			%	110			70-130	Pass	
Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
<b>Spike - % Recovery</b>									
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>				Result 1					
TRH C6-C9	S19-No41937	NCP	%	81			70-130	Pass	
TRH C10-C14	S19-No35398	NCP	%	95			70-130	Pass	
<b>Spike - % Recovery</b>									
<b>BTEX</b>				Result 1					
Benzene	S19-No41937	NCP	%	87			70-130	Pass	
Toluene	S19-No41937	NCP	%	85			70-130	Pass	
Ethylbenzene	S19-No41937	NCP	%	93			70-130	Pass	
m&p-Xylenes	S19-No41937	NCP	%	84			70-130	Pass	
o-Xylene	S19-No41937	NCP	%	83			70-130	Pass	
Xylenes - Total	S19-No41937	NCP	%	84			70-130	Pass	
<b>Spike - % Recovery</b>									
<b>Total Recoverable Hydrocarbons - 2013 NEPM Fractions</b>				Result 1					
Naphthalene	S19-No41937	NCP	%	81			70-130	Pass	
TRH C6-C10	S19-No41937	NCP	%	83			70-130	Pass	
TRH >C10-C16	S19-No35398	NCP	%	91			70-130	Pass	
<b>Spike - % Recovery</b>									
<b>Heavy Metals</b>				Result 1					
Arsenic	S19-No40801	NCP	%	100			70-130	Pass	
Cadmium	S19-No40801	NCP	%	108			70-130	Pass	
Chromium	S19-No40801	NCP	%	111			70-130	Pass	
Copper	S19-No40801	NCP	%	110			70-130	Pass	
Lead	S19-No40801	NCP	%	114			70-130	Pass	
Mercury	S19-No40801	NCP	%	121			70-130	Pass	
Nickel	S19-No40801	NCP	%	112			70-130	Pass	
Zinc	S19-No40801	NCP	%	105			70-130	Pass	
Test	Lab Sample ID	QA Source	Units	Result 1			Acceptance Limits	Pass Limits	Qualifying Code
<b>Duplicate</b>									
<b>Total Recoverable Hydrocarbons - 1999 NEPM Fractions</b>				Result 1	Result 2	RPD			
TRH C6-C9	S19-No37845	NCP	mg/kg	< 20	< 20	<1	30%	Pass	
TRH C10-C14	S19-No36589	NCP	mg/kg	< 20	< 20	<1	30%	Pass	
TRH C15-C28	S19-No36589	NCP	mg/kg	< 50	< 50	<1	30%	Pass	
TRH C29-C36	S19-No36589	NCP	mg/kg	< 50	< 50	<1	30%	Pass	
<b>Duplicate</b>									
<b>BTEX</b>				Result 1	Result 2	RPD			
Benzene	S19-No37845	NCP	mg/kg	< 0.1	< 0.1	<1	30%	Pass	
Toluene	S19-No37845	NCP	mg/kg	< 0.1	< 0.1	<1	30%	Pass	
Ethylbenzene	S19-No37845	NCP	mg/kg	< 0.1	< 0.1	<1	30%	Pass	
m&p-Xylenes	S19-No37845	NCP	mg/kg	< 0.2	< 0.2	<1	30%	Pass	
o-Xylene	S19-No37845	NCP	mg/kg	< 0.1	< 0.1	<1	30%	Pass	
Xylenes - Total	S19-No37845	NCP	mg/kg	< 0.3	< 0.3	<1	30%	Pass	

Duplicate								
Total Recoverable Hydrocarbons - 2013 NEPM Fractions				Result 1	Result 2	RPD		
Naphthalene	S19-No37845	NCP	mg/kg	< 0.5	< 0.5	<1	30%	Pass
TRH C6-C10	S19-No37845	NCP	mg/kg	< 20	< 20	<1	30%	Pass
TRH >C10-C16	S19-No36589	NCP	mg/kg	< 50	< 50	<1	30%	Pass
TRH >C16-C34	S19-No36589	NCP	mg/kg	< 100	< 100	<1	30%	Pass
TRH >C34-C40	S19-No36589	NCP	mg/kg	< 100	< 100	<1	30%	Pass
Duplicate								
Heavy Metals				Result 1	Result 2	RPD		
Arsenic	S19-No40799	NCP	mg/kg	11	8.3	31	30%	Fail Q15
Cadmium	S19-No40799	NCP	mg/kg	< 0.4	< 0.4	<1	30%	Pass
Chromium	S19-No40799	NCP	mg/kg	13	12	16	30%	Pass
Copper	S19-No40799	NCP	mg/kg	30	77	88	30%	Fail Q15
Lead	S19-No40799	NCP	mg/kg	26	25	1.0	30%	Pass
Mercury	S19-No40799	NCP	mg/kg	< 0.1	< 0.1	<1	30%	Pass
Nickel	S19-No40799	NCP	mg/kg	< 5	< 5	<1	30%	Pass
Zinc	S19-No40799	NCP	mg/kg	39	25	44	30%	Fail Q15
Duplicate								
				Result 1	Result 2	RPD		
% Moisture	S19-No38860	NCP	%	17	17	1.0	30%	Pass

## Comments

### Sample Integrity

Custody Seals Intact (if used)	N/A
Attempt to Chill was evident	Yes
Sample correctly preserved	Yes
Appropriate sample containers have been used	Yes
Sample containers for volatile analysis received with minimal headspace	Yes
Samples received within HoldingTime	Yes
Some samples have been subcontracted	No

### Qualifier Codes/Comments

Code	Description
G01	The LORs have been raised due to matrix interference
N01	F2 is determined by arithmetically subtracting the "naphthalene" value from the ">C10-C16" value. The naphthalene value used in this calculation is obtained from volatiles (Purge & Trap analysis).
N02	Where we have reported both volatile (P&T GCMS) and semivolatile (GCMS) naphthalene data, results may not be identical. Provided correct sample handling protocols have been followed, any observed differences in results are likely to be due to procedural differences within each methodology. Results determined by both techniques have passed all QAQC acceptance criteria, and are entirely technically valid.
N04	F1 is determined by arithmetically subtracting the "Total BTEX" value from the "C6-C10" value. The "Total BTEX" value is obtained by summing the concentrations of BTEX analytes. The "C6-C10" value is obtained by quantitating against a standard of mixed aromatic/aliphatic analytes.
Q15	The RPD reported passes Eurofins Environment Testing's QC - Acceptance Criteria as defined in the Internal Quality Control Review and Glossary page of this report.

### Authorised By

Ursula Long	Analytical Services Manager
Andrew Sullivan	Senior Analyst-Organic (NSW)
Gabriele Cordero	Senior Analyst-Metal (NSW)



**Glenn Jackson**

### General Manager

Final report - this Report replaces any previously issued Report

- Indicates Not Requested

\* Indicates NATA accreditation does not cover the performance of this service

Measurement uncertainty of test data is available on request or please [click here](#).

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## Appendix F

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ProUCL 5.1 Inputs & Outputs

	A	B
1	sample	copper
2	BH101	21
3	BH102	130
4	BH103	21
5	BD3_20191	4
6	BH104	66
7	BH104	280
8	BH104	29
9	BH105	42
10	BH105	51
11	BH105	45
12	BH106	44
13	BH106	45
14	BDA/201910	22
15	BH109	14
16	201	9
17	202	130
18	BD2611201	120

	A	B	C	D	E	F	G	H	I	J	K	L	
1	UCL Statistics for Uncensored Full Data Sets												
2													
3	User Selected Options												
4	Date/Time of Computation			ProUCL 5.12/12/2019 2:00:08 PM									
5	From File			copper fill input.xls									
6	Full Precision			OFF									
7	Confidence Coefficient			95%									
8	Number of Bootstrap Operations			2000									
9													
10													
11	copper												
12													
13	General Statistics												
14	Total Number of Observations				17		Number of Distinct Observations				14		
15							Number of Missing Observations				0		
16	Minimum				4		Mean				63.12		
17	Maximum				280		Median				44		
18	SD				68.96		Std. Error of Mean				16.72		
19	Coefficient of Variation				1.093		Skewness				2.206		
20													
21	Normal GOF Test												
22	Shapiro Wilk Test Statistic				0.74		Shapiro Wilk GOF Test						
23	5% Shapiro Wilk Critical Value				0.892		Data Not Normal at 5% Significance Level						
24	Lilliefors Test Statistic				0.276		Lilliefors GOF Test						
25	5% Lilliefors Critical Value				0.207		Data Not Normal at 5% Significance Level						
26	Data Not Normal at 5% Significance Level												
27													
28	Assuming Normal Distribution												
29	95% Normal UCL					95% UCLs (Adjusted for Skewness)							
30	95% Student's-t UCL			92.32		95% Adjusted-CLT UCL (Chen-1995)					100.2		
31						95% Modified-t UCL (Johnson-1978)					93.81		
32													
33	Gamma GOF Test												
34	A-D Test Statistic			0.389		Anderson-Darling Gamma GOF Test							
35	5% A-D Critical Value			0.763		Detected data appear Gamma Distributed at 5% Significance Level							
36	K-S Test Statistic			0.168		Kolmogorov-Smirnov Gamma GOF Test							
37	5% K-S Critical Value			0.214		Detected data appear Gamma Distributed at 5% Significance Level							
38	Detected data appear Gamma Distributed at 5% Significance Level												
39													
40	Gamma Statistics												
41	k hat (MLE)			1.165		k star (bias corrected MLE)					0.999		
42	Theta hat (MLE)			54.19		Theta star (bias corrected MLE)					63.21		
43	nu hat (MLE)			39.6		nu star (bias corrected)					33.95		
44	MLE Mean (bias corrected)			63.12		MLE Sd (bias corrected)					63.17		
45						Approximate Chi Square Value (0.05)					21.62		
46	Adjusted Level of Significance			0.0346		Adjusted Chi Square Value					20.6		
47													
48	Assuming Gamma Distribution												
49	95% Approximate Gamma UCL (use when n>=50)				99.09		95% Adjusted Gamma UCL (use when n<50)					104	
50													
51	Lognormal GOF Test												
52	Shapiro Wilk Test Statistic				0.978		Shapiro Wilk Lognormal GOF Test						
53	5% Shapiro Wilk Critical Value				0.892		Data appear Lognormal at 5% Significance Level						
54	Lilliefors Test Statistic				0.118		Lilliefors Lognormal GOF Test						

	A	B	C	D	E	F	G	H	I	J	K	L	
55	5% Lilliefors Critical Value					0.207	Data appear Lognormal at 5% Significance Level						
56	Data appear Lognormal at 5% Significance Level												
57													
58	Lognormal Statistics												
59	Minimum of Logged Data					1.386	Mean of logged Data					3.658	
60	Maximum of Logged Data					5.635	SD of logged Data					1.061	
61													
62	Assuming Lognormal Distribution												
63	95% H-UCL					141.7	90% Chebyshev (MVUE) UCL					120.8	
64	95% Chebyshev (MVUE) UCL					146.1	97.5% Chebyshev (MVUE) UCL					181.2	
65	99% Chebyshev (MVUE) UCL					250.1							
66													
67	Nonparametric Distribution Free UCL Statistics												
68	Data appear to follow a Discernible Distribution at 5% Significance Level												
69													
70	Nonparametric Distribution Free UCLs												
71	95% CLT UCL					90.63	95% Jackknife UCL					92.32	
72	95% Standard Bootstrap UCL					88.8	95% Bootstrap-t UCL					116.1	
73	95% Hall's Bootstrap UCL					148.5	95% Percentile Bootstrap UCL					91.76	
74	95% BCA Bootstrap UCL					102.2							
75	90% Chebyshev(Mean, Sd) UCL					113.3	95% Chebyshev(Mean, Sd) UCL					136	
76	97.5% Chebyshev(Mean, Sd) UCL					167.6	99% Chebyshev(Mean, Sd) UCL					229.5	
77													
78	Suggested UCL to Use												
79	95% Adjusted Gamma UCL					104							
80													
81	Note: Suggestions regarding the selection of a 95% UCL are provided to help the user to select the most appropriate 95% UCL.												
82	Recommendations are based upon data size, data distribution, and skewness.												
83	These recommendations are based upon the results of the simulation studies summarized in Singh, Maichle, and Lee (2006).												
84	However, simulations results will not cover all Real World data sets; for additional insight the user may want to consult a statistician.												
85													



	A	B
1	sample	lead
2	BH101	100
3	BH102	170
4	BD2_20191022	47
5	BH104	36
6	BH104	120
7	BH104	1100
8	BH104	160
9	BH105	74
10	BH105	130
11	BH105	95
12	BH106	15
13	BH106	99
14	BDA/20191022	14
15	BH109	5
16	201	25
17	202	30
18	BD26112019	1300

	A	B	C	D	E	F	G	H	I	J	K	L
1	UCL Statistics for Uncensored Full Data Sets											
2												
3	User Selected Options											
4	Date/Time of Computation			ProUCL 5.12/12/2019 2:03:04 PM								
5	From File			lead fill input.xls								
6	Full Precision			OFF								
7	Confidence Coefficient			95%								
8	Number of Bootstrap Operations			2000								
9												
10												
11	lead											
12												
13	General Statistics											
14	Total Number of Observations				17		Number of Distinct Observations				17	
15							Number of Missing Observations				0	
16	Minimum				5		Mean				207.1	
17	Maximum				1300		Median				95	
18	SD				378.9		Std. Error of Mean				91.89	
19	Coefficient of Variation				1.83		Skewness				2.562	
20												
21	Normal GOF Test											
22	Shapiro Wilk Test Statistic				0.521		Shapiro Wilk GOF Test					
23	5% Shapiro Wilk Critical Value				0.892		Data Not Normal at 5% Significance Level					
24	Lilliefors Test Statistic				0.421		Lilliefors GOF Test					
25	5% Lilliefors Critical Value				0.207		Data Not Normal at 5% Significance Level					
26	Data Not Normal at 5% Significance Level											
27												
28	Assuming Normal Distribution											
29	95% Normal UCL					95% UCLs (Adjusted for Skewness)						
30	95% Student's-t UCL				367.5		95% Adjusted-CLT UCL (Chen-1995)				419.2	
31							95% Modified-t UCL (Johnson-1978)				377	
32												
33	Gamma GOF Test											
34	A-D Test Statistic				1.19		Anderson-Darling Gamma GOF Test					
35	5% A-D Critical Value				0.789		Data Not Gamma Distributed at 5% Significance Level					
36	K-S Test Statistic				0.267		Kolmogorov-Smirnov Gamma GOF Test					
37	5% K-S Critical Value				0.219		Data Not Gamma Distributed at 5% Significance Level					
38	Data Not Gamma Distributed at 5% Significance Level											
39												
40	Gamma Statistics											
41	k hat (MLE)				0.599		k star (bias corrected MLE)				0.532	
42	Theta hat (MLE)				345.9		Theta star (bias corrected MLE)				389.1	
43	nu hat (MLE)				20.35		nu star (bias corrected)				18.09	
44	MLE Mean (bias corrected)				207.1		MLE Sd (bias corrected)				283.8	
45						Approximate Chi Square Value (0.05)				9.459		
46	Adjusted Level of Significance				0.0346		Adjusted Chi Square Value				8.81	
47												
48	Assuming Gamma Distribution											
49	95% Approximate Gamma UCL (use when n>=50))				396.1		95% Adjusted Gamma UCL (use when n<50)				425.3	
50												
51	Lognormal GOF Test											
52	Shapiro Wilk Test Statistic				0.954		Shapiro Wilk Lognormal GOF Test					
53	5% Shapiro Wilk Critical Value				0.892		Data appear Lognormal at 5% Significance Level					
54	Lilliefors Test Statistic				0.162		Lilliefors Lognormal GOF Test					

	A	B	C	D	E	F	G	H	I	J	K	L	
55	5% Lilliefors Critical Value					0.207	Data appear Lognormal at 5% Significance Level						
56	Data appear Lognormal at 5% Significance Level												
57													
58	Lognormal Statistics												
59	Minimum of Logged Data					1.609	Mean of logged Data					4.3	
60	Maximum of Logged Data					7.17	SD of logged Data					1.433	
61													
62	Assuming Lognormal Distribution												
63	95% H-UCL					688.9	90% Chebyshev (MVUE) UCL					409	
64	95% Chebyshev (MVUE) UCL					510.4	97.5% Chebyshev (MVUE) UCL					651.1	
65	99% Chebyshev (MVUE) UCL					927.6							
66													
67	Nonparametric Distribution Free UCL Statistics												
68	Data appear to follow a Discernible Distribution at 5% Significance Level												
69													
70	Nonparametric Distribution Free UCLs												
71	95% CLT UCL					358.2	95% Jackknife UCL					367.5	
72	95% Standard Bootstrap UCL					353.4	95% Bootstrap-t UCL					1187	
73	95% Hall's Bootstrap UCL					1365	95% Percentile Bootstrap UCL					362.6	
74	95% BCA Bootstrap UCL					424.3							
75	90% Chebyshev(Mean, Sd) UCL					482.7	95% Chebyshev(Mean, Sd) UCL					607.6	
76	97.5% Chebyshev(Mean, Sd) UCL					780.9	99% Chebyshev(Mean, Sd) UCL					1121	
77													
78	Suggested UCL to Use												
79	95% Chebyshev (Mean, Sd) UCL					607.6							
80													
81	Note: Suggestions regarding the selection of a 95% UCL are provided to help the user to select the most appropriate 95% UCL.												
82	Recommendations are based upon data size, data distribution, and skewness.												
83	These recommendations are based upon the results of the simulation studies summarized in Singh, Maichle, and Lee (2006).												
84	However, simulations results will not cover all Real World data sets; for additional insight the user may want to consult a statistician.												
85													